

Plasmid-Encoded Anthranilate Synthase (TrpEG) in *Buchnera aphidicola* from Aphids of the Family Pemphigidae

ROELAND C. H. J. VAN HAM,[†] DAVID MARTÍNEZ-TORRES, ANDRÉS MOYA,
AND AMPARO LATORRE*

Department of Genetics, University of Valencia, 46100 Burjassot, Valencia, Spain

Received 20 July 1998/Accepted 9 October 1998

Buchnera aphidicola is an obligate intracellular symbiont of aphids. One of its proposed functions is the synthesis of essential amino acids, nutrients required by aphids but deficient in their diet of plant phloem sap. The genetic organization of the tryptophan pathway in *Buchnera* from proliferous aphids of the family Aphididae has previously been shown to reflect a capacity to overproduce this essential amino acid (C.-Y. Lai, L. Baumann, and P. Baumann, Proc. Natl. Acad. Sci. USA 91:3819–3823, 1994). This involved amplification of the genes for the first enzyme in the pathway, anthranilate synthase (TrpEG), on a low-copy-number plasmid. Here we report on the finding and molecular characterization of TrpEG-encoding plasmids in *Buchnera* from aphids of the distantly related family Pemphigidae. *Buchnera* from *Tetraneura caerulea* contained a 3.0-kb plasmid (pBTc2) that carried a single copy of *trpEG* and resembled *trpEG* plasmids of *Buchnera* from the Aphididae. The second plasmid (pBPs2), isolated from *Buchnera* of *Pemphigus spyrothecae*, contained a different replicon. It consisted of a putative origin of replication containing iterons and an open reading frame, designated *repAC*, which showed a high similarity to the gene encoding the replication initiation protein RepA of the RepA/C replicon from the broad-host-range IncA/C group of plasmids. The plasmid population was heterogeneous with respect to the number of tandem repeats of a 1.8-kb unit carrying *repAC*, *trpG*, and remnants of *trpE*. The two principal forms consisted of either five or six copies of this repeat and a single-copy region carrying *repAC*, the putative origin of replication, and *trpE*. The unexpected finding of elements of the RepA/C replicon in previously characterized *trpEG* plasmids from *Buchnera* of the Aphididae suggests that a replacement of replicons has occurred during the evolution of these plasmids, which may point to a common ancestry for all *Buchnera trpEG* amplifications.

Aphids are dependent on an intracellular symbiont (*Buchnera aphidicola*, Proteobacteria) for normal growth and reproduction (7, 19, 45). The bacteria reside in specialized cells in the aphid hemocoel and are transmitted maternally through infection of eggs or embryos (11, 26). Phylogenetic studies have revealed two major characteristics of the evolutionary history of the association (37, 39); (i) the symbiosis had a single origin, dated about 150 million to 250 million years ago; and (ii) host and symbiont lineages have since diverged strictly in parallel. The association, like other symbioses in insects feeding on restricted and unbalanced diets, is thought to have a nutritional basis (5–7, 20). Aphids feed on plant phloem sap, a diet rich in carbohydrates but deficient in nitrogenous compounds, including most essential amino acids (16, 18, 27, 41). *Buchnera* has been proposed as the source of essential amino acids for the aphid (14), which has been supported by evidence from nutritional and physiological studies (17, 20–22, 45) and, more recently, by the finding of genetic modifications in the tryptophan and leucine biosynthetic pathways in *Buchnera* from several aphid species. In both cases, genes encoding key enzymes in the respective pathways were found to be amplified and relocated to plasmids (10, 30).

Lai et al. (30) found that the genes for the two subunits of anthranilate synthase (*trpE* and *trpG*), the first enzyme in the pathway leading to tryptophan, are contained on a low-copy-

number plasmid in *Buchnera* from the aphid *Schizaphis graminum* (Aphididae). The plasmid consisted of four identical tandem repeats of a 3.6-kb *trpEG*-containing unit. *trpEG* was amplified about 16-fold over the remaining genes of the pathway, which reside in a single locus [*trpDC(F)BA*] on the *Buchnera* chromosome (38). *trpEG*-encoding plasmids have since been found in *Buchnera* from various species of the Aphididae (4, 32, 42, 43), and their overall similarity suggests that the amplification is ancestral to this lineage (32). The Aphididae is the largest and evolutionarily most successful family of aphids. Many of its species have high growth and reproductive rates, and it includes a number of major agricultural pests (8).

In contrast, *Buchnera* from the aphid *Schlechtendalia chinensis*, a member of the distantly related family Pemphigidae, was found to carry all the genes of the tryptophan pathway on the chromosome, organized into two single-copy linkage groups [*trpEG* and *trpDC(F)BA*] (31). This difference in organization, which is assumed to reflect a difference in the capacity to overproduce tryptophan, has been linked to potentially varying requirements for the amino acid by aphid hosts. *S. chinensis* has a long development time and a low reproductive rate, and its demand for tryptophan may therefore be lower than in the highly prolific aphids of the Aphididae (5–7, 9, 31).

Here we report on the finding and molecular characterization of *trpEG*-containing plasmids in *Buchnera* from the aphids *Tetraneura caerulea* and *Pemphigus spyrothecae*, both belonging to the Pemphigidae. We propose a scenario for the evolution of *trp* in *Buchnera* in which there was a single ancestral transfer of *trpEG* to a RepA/C-like replicon followed by independent events of replicon replacement and back-transfer of *trpEG* to the chromosome in different lineages.

* Corresponding author. Mailing address: Department of Genetics, University of Valencia, c/Dr. Moliner 50, 46100 Burjassot, Valencia, Spain. Phone: 34-96-3864505. Fax: 34-96-3983029. E-mail: amparo.latorre@uv.es.

[†] Present address: Department of Biology, University of York, York, YO10 5YW, United Kingdom.

MATERIALS AND METHODS

Aphid material and *B. aphidicola*. Leaves carrying galls produced by the aphid *P. spyrothecae* were collected from poplar trees (La Yesa, Spain). *T. caeruleus* was collected from galls on elm trees (Bugarra, Spain). The galls were kept at 4°C until needed for further manipulation of the animals.

General methods. *Buchnera* plasmid and genomic DNA was isolated as previously described (49). DNA was manipulated by standard methods (2, 44). Southern hybridizations were done with either digoxigenin-labeled probes (Boehringer Mannheim) or the enhanced chemiluminescence (ECL) system (Amersham), as specified by the manufacturers.

Cloning and sequencing. Cloning of *B. aphidicola* (*T. caeruleus*) plasmid DNA has been described previously (49). One clone with an *EcoRI* insert of 3.0 kb was completely sequenced. Populations of aphids from the Pemphigidae are often small, and only a limited number of *P. spyrothecae* animals could be collected. Consequently, the total amount of plasmid DNA available for experiments was reduced and a shotgun-cloning approach was taken to maximize cloning efficiency. Approximately 30 ng of *B. aphidicola* (*P. spyrothecae*) plasmid DNA was digested with *XbaI* and ligated to *XbaI*-digested, phosphatase-treated pBluescript II SK (Stratagene). After transformation of *Escherichia coli* DH5 α with the ligation and screening of recombinant colonies, clones carrying inserts of 2.1 and 1.8 kb were analyzed by restriction enzyme digestion and end sequencing (see Fig. 4A). All the clones were identical, and one clone of each size was completely sequenced. Sequence information obtained from these clones was used to design oligonucleotide primers complementary to *repAC* and *trpE* that would allow the PCR amplification of a region spanning a 1.5-kb *XbaI* fragment that had remained uncloned during shotgun cloning (*repAC*P_{1,5'}-AGA GCA ATG AAA AAC GCT TCT CG-3'; and *trpE*P_{1,5'}-TCA GGT GAC GCT CCA AAT AAG G-3'). Cycling was performed with the GeneAmp 2400 System (Perkin Elmer) and consisted of 5 cycles of 94°C for 30 s, 62°C for 1 min, and 72°C for 1 min followed by 30 cycles of 94°C for 30 s, 58°C for 30 s, and 72°C for 30 s, with a final 10-min extension at 72°C. The PCR yielded fragments of 1.53 and 1.70 kb, which were ligated into a T vector (36) and transformed into *E. coli* XL1Blue (Stratagene). Two clones with inserts corresponding to the sizes of the two PCR fragments were completely sequenced (see Fig. 4A). Nested deletions of all selected cloned fragments were generated in both directions by using the nested deletions kit (Pharmacia). Nucleotide sequencing was performed with the AmpliTaqF Dye Deoxy Terminator cycle-sequencing kit (Perkin Elmer) and the 373 automated DNA sequencer (Perkin-Elmer) as recommended by the manufacturer.

Quantitative hybridization. The ratio between the copy number of the 1.8- and 2.1-kb *XbaI* fragments per plasmid was determined by densitometric scans of Southern blots containing eight *XbaI* digests of *B. aphidicola* (*P. spyrothecae*) total DNA and hybridized with a probe containing *trpG* and *repAC*. Scanning was performed with a Gelstation (TDI) image analyzer, and images were analyzed with the Intelligent Quantifier Bioimage software (Bio Systems Corp.).

Serial partial digestion of *B. aphidicola* (*P. spyrothecae*) total DNA. *B. aphidicola* (*P. spyrothecae*) total DNA was first digested to completion with *AccI* and *SacI* to remove most of the single-copy region from the plasmid that contained an *XbaI* site. Partial digestions with serial dilutions of *XbaI* were then performed by the method of Ausubel et al. (2), starting with 5 U of enzyme and using approximately 200 ng of DNA per digest. Controls included DNAs digested to completion with *XbaI*, *AccI*, *SacI*, and *AccI* plus *SacI*. Electrophoresis was performed overnight on 0.5% agarose gels in 0.5 \times Tris-borate-EDTA at 1.2 V/cm and was followed by Southern blotting. Hybridization was done with the cloned 1.8-kb *XbaI* fragment as a probe. We used 1-kb ladder DNA (Gibco-BRL) and *HindIII*-digested λ DNA as molecular weight markers.

Computer analysis of the DNA sequences. DNA sequence data were assembled and analyzed with the Genetics Computer Group (GCG) program package 8.0 for the VAX/VMS (15). BLASTP searches (1) were done at the network server of the National Center for Biotechnology Information. Phylogenetic analysis of amino acid sequences was performed with the PROTDIST (Dayhoff PAM distances) and the NEIGHBOR (neighbor-joining method) programs as implemented in PHYLIP version 3.5 (23).

Nucleotide sequence accession numbers. The nucleotide sequence data reported in this paper have been deposited in the GenEMBL databases under accession no. AJ012333 and AJ012334 (PBTC2 and PBPs2, respectively).

RESULTS

Detection of plasmids. *B. aphidicola* (*T. caeruleus*) has previously been shown to carry a small, cryptic plasmid (pBTc1, 1.74 kb) that contained the *repA*₁ replicon (related to IncFII plasmids) found in leucine plasmids (49). Restriction enzyme analysis of plasmid DNA preparations from this species indicated the presence of a second plasmid of about 3 kb. Similar observations were made with plasmid DNA isolated from *B. aphidicola* (*P. spyrothecae*) (48). Plasmid DNA preparations from both species were here further analyzed by Southern hybridization, cloning, and nucleotide sequencing. The two natu-

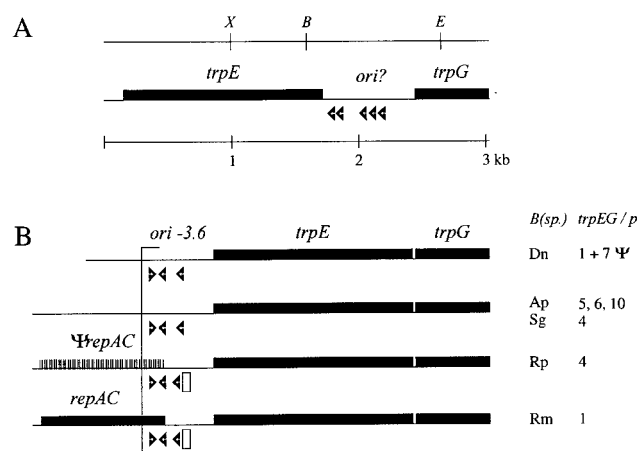


FIG. 1. Linearized physical maps of *B. aphidicola* *trpEG* plasmids. All genes are transcribed in the rightward direction. Open arrowheads indicate the approximate position and orientation of DnaA boxes. (A) Restriction site and genetic map of pBTc2 from *B. aphidicola* (*T. caeruleus*). The region denoted *ori?* contains the putative origin of replication. Restriction enzyme sites: X, *XbaI*; B, *BglII*; E, *EcoRI*. (B) Genetic maps of the repeated units of *B. aphidicola* (Aphididae) *trpEG* plasmids (adapted from reference 42 with permission from the publisher). *B.(sp.)*, species names (see below); *trpEG/p*, number of repeat units per plasmid (42); *ori-3.6*, putative origin of replication; Ψ , repeat units containing *trpEG* pseudogenes (32); striped box and Ψ *repAC*, location of a putative *repAC* pseudogene; open rectangle, 19-bp element similar to consensus sequence of *RepA/C* iterons. Species abbreviations: Dn, *Diuraphis noxia*; Ap, *Acyrthosiphon pisum*; Sg, *Schizaphis graminum*; Rp, *Rhopalosiphum padi*; Rm, *Rhopalosiphum maidis*.

ral plasmids described below are designated pBTc2 and pBPs2, after their source *B. aphidicola* (*T. caeruleus*) and *B. aphidicola* (*P. spyrothecae*), respectively.

Characterization of pBTc2. An *EcoRI* shotgun cloning of *B. aphidicola* (*T. caeruleus*) plasmid DNA yielded four recombinants with an insert of 3.0 kb. Restriction fragment analysis showed these to be identical, and one clone was completely sequenced. A physical map of plasmid pBTc2 is presented in Fig. 1A. The G+C content of the 3.0-kb fragment is 20.74 mol%, which is similar to the 21.2 mol% of plasmid pBTc1 from this species (49) and is consistent with the low G+C content (28 mol%) of *Buchnera* genomic DNA (28). Two open reading frames (ORFs) were found, which correspond to *trpE* and *trpG*. The deduced amino acid sequences are 52 and 53% identical to TrpE and TrpG, respectively, from *Buchnera* of the phylogenetically treated aphid *S. chinensis* (Pemphigidae). Potential regulatory elements for gene expression (33) could not be identified.

Most *trpEG*-containing plasmids of *Buchnera* from aphids of the family Aphididae share a conserved 500- to 600-nucleotide (nt) sequence immediately upstream of *trpE* which has features characteristic of an origin of replication (29). The region has been designated *ori-3.6* and contains three DnaA boxes (consensus 5'-TTATCCACA-3') and regions of low G+C content (Fig. 1B) (43). A similar region, 717 nt in length, is present downstream of *trpE* on pBTc2 and may therefore contain the origin of replication of the plasmid. It has a very low G+C content (11 mol%) and contains five DnaA boxes in direct orientation, only one of which deviates by a single nucleotide from the consensus sequence. Beyond a high A+T content, the region has no further sequence similarity to *ori-3.6*.

B. aphidicola (Aphididae) *trpEG* plasmids are typically composed of tandemly repeated, identical 3.2- to 3.6-kb units, each carrying a copy of *trpEG* and *ori-3.6* (42, 43). To determine the composition of pBTc2, a Southern hybridization analysis was performed of various *EcoRI*-digested and undigested *B. aphid-*

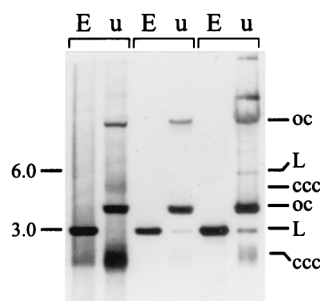


FIG. 2. Southern blot analysis of *EcoRI*-digested (E) and undigested (u) *B. aphidicola* (*T. caerulescens*) DNA preparations. Cloned pBTc2 was used as a hybridization probe. The different forms of the putative plasmids are marked: open circular (oc), linear (L), and covalently closed circular (ccc).

icola (*T. caerulescens*) DNA preparations, using cloned pBTc2 as a probe. For each preparation, two strongly hybridizing bands were detected in lanes with undigested DNA and a single strong band was detected in lanes with *EcoRI*-digested DNA (Fig. 2). These three bands correspond to, respectively, the open-circular, covalently closed supercoiled, and linear forms of a plasmid of 3.0 kb. In addition, three larger, more weakly hybridizing bands were detected in undigested DNAs. Their sizes correspond to the different forms of a plasmid of 6.0 kb. *EcoRI* digestion converted all the different forms into a single linear fragment of 3.0 kb. These findings suggest that pBTc2 consists mainly of one copy of the sequenced 3.0-kb unit but that a fraction of the plasmid population is present as a dimer. In one preparation, molecules of an even higher molecular weight were detected (Fig. 2, right lane), indicating that further multimerization of the plasmid occurs.

Identification of pBPs2. Restriction enzyme analysis of a plasmid DNA preparation from *B. aphidicola* (*P. spyrothecae*) indicated the presence of at least two plasmids. One of these (pBPs2) was larger than 12 kb as estimated from the migration of bands of undigested plasmid DNA in ethidium bromide-stained agarose gels. Analogous to *trpEG* plasmids of *B. aphidicola* (Aphididae), it seemed to be composed of repeated elements, because summation of three restriction fragments generated with *XbaI* (2.1, 1.8, and 1.5 kb) added up to only 5.4 kb. Moreover, the 1.8-kb band stained more intensely than the other two fragments. Southern hybridization with a *trpG*-containing probe revealed that the plasmid carried sequences homologous to *trpG* and reinforced the assumption that it was composed of repeated elements (Fig. 3). *AccI* and *PvuII* each yielded a single, slightly blurred band of 13 to 15 kb, while *EcoRI*, *PstI*, *ScaI*, and *XhoI* yielded two bands, one of which also migrated at about 13 kb and the second of which migrated at more than 23 kb. Digests with *EcoRV*, *ClaI*, and *XbaI* yielded either one or two bands migrating at 1.5 to 2.1 kb. These hybridization patterns were at first explained in terms of the 13- to 15-kb *AccI* and *PvuII* bands representing the linear form of the putative plasmid. The fact that the plasmid could be linearized, in contrast to most previously characterized *trpEG* plasmids (42), implied that it contained a single-copy region in addition to repeated elements. Recognition sites for *EcoRI*, *PstI*, *ScaI*, and *XhoI* are absent from the plasmid, and hence the two detected bands correspond to the open-circular (at 23 kb) and the comigrating linear and closed-circular forms of a 13- to 15-kb plasmid. Finally, recognition sites for *EcoRV*, *ClaI*, and *XbaI* are present in the repeated part of the plasmid and hence convert it into the constituent repeat units.

Cloning and sequencing. *B. aphidicola* (*P. spyrothecae*) plasmid DNA was shotgun cloned with *XbaI*. Screening of 108

recombinant colonies showed that 10% of plasmids had an insert of 2.1 kb and 20% had an insert of 1.8 kb, corresponding in size to the two *XbaI* fragments detected in the Southern hybridization (Fig. 3). The 1.5-kb *XbaI* fragment, initially detected in agarose gels, was not found. Restriction enzyme analysis and end sequencing of the recombinant plasmids suggested that all clones of the same size were identical and one of each group was completely sequenced (Fig. 4B).

The 2.1-kb fragment contained three ORFs, the first two of which corresponded to the 3' half of *trpE* and a complete copy of *trpG*, with the stop and start codons of the two genes overlapping. BLASTP homology searches with the putative product of the third ORF yielded a high probability score (i.e., 893) with the N-terminal part of replication initiation protein RepA encoded by *E. coli* plasmid RA1, which belongs to the broad-host-range IncA/C incompatibility group (34). The amino acid sequences were 69% identical over the first 243 positions. The *B. aphidicola* (*P. spyrothecae*) gene is designated *repAC₁*. The 1.8-kb fragment also contained three ORFs, the first of which was similar to the gene encoding the C-terminal part of the same RepA protein that was lacking from the 2.1-kb fragment. Because the coding sequences are in frame, the two fragments are most probably contiguous on the plasmid. The second ORF corresponded to a complete copy of the *trpG* gene, and the third ORF, again, corresponds to the gene encoding the N-terminal part of RepA. Both sequences were completely identical to the homologous sequences in the 2.1-kb fragment.

The 2.1-kb *XbaI* fragment contained single *AccI* and *SacI* restriction sites (Fig. 4A). Since the former enzyme was at first assumed to linearize the plasmid, this implied that the fragment contained part of a single-copy region of the plasmid. However, neither of the two sequenced fragments contained a restriction site for *PvuII*, suggesting that this site had to be present in the uncloned 1.5-kb *XbaI* fragment and, hence, that the 1.8-kb fragment represented the repeated unit of the plasmid. In addition, the uncloned 1.5-kb *XbaI* fragment was expected to contain the missing 5' end of the *trpE* gene and possibly the 3' end of a *repAC* gene. Oligonucleotide primers complementary to *trpE* and *repAC₁* that would allow the amplification of this fragment were designed (Fig. 4A).

PCR yielded two products, which were 1.53 and 1.70 kb in size. After cloning, one recombinant plasmid from each size group was completely sequenced on both strands (Fig. 4B). Both fragments contained the 3' end of a *repAC* gene and the missing 5' part of *trpE*, as well as a single *PvuII* site. However, the 3' end of the *repAC* sequences differed slightly from the homologous sequence of the 1.8-kb *XbaI* fragment. The latter contained only one copy of a 19-bp element (5'-CAACAAAC

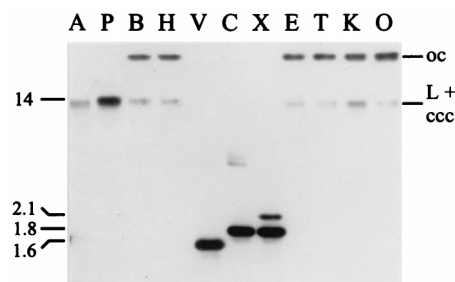


FIG. 3. Southern blot analysis of *B. aphidicola* (*P. spyrothecae*) plasmid DNA. Hybridizations were performed with a probe complementary to *trpG*. Restriction enzymes: A, *AccI*; P, *PvuII*; B, *BamHI*; H, *HindIII*; V, *EcoRV*; C, *ClaI*; X, *XbaI*; E, *EcoRI*; T, *PstI*; K, *ScaI*; O, *XhoI*. oc, open circular; L, linear; ccc, covalently closed circular.

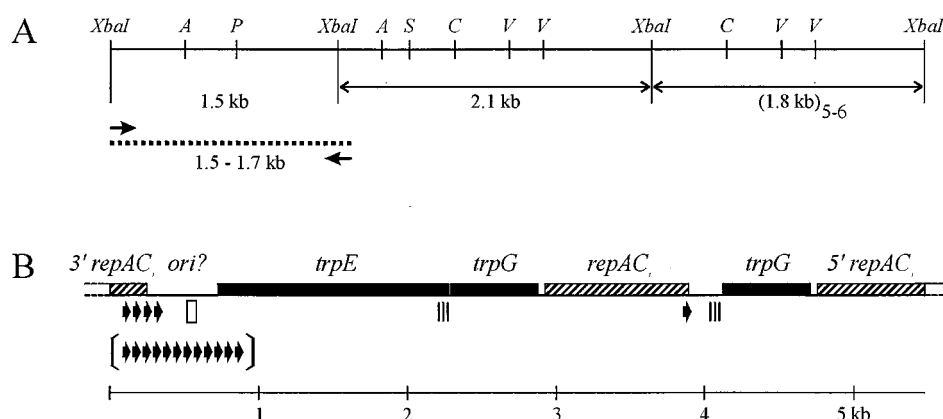


FIG. 4. Physical map of cloned and sequenced portion of pBPs2 from *B. aphidicola* (*P. spyrothecae*). (A) Restriction site and clone map. Double-headed arrows, cloned *XbaI* restriction fragments; arrows, oligonucleotide primers used for PCR; dotted line, cloned PCR product. Restriction enzyme sites: A, *AccI*; P, *PvuII*; S, *SacI*; C, *ClaI*; V, *EcoRV*. (B) Genetic map. All genes are transcribed in the rightward direction. The region denoted *ori?* contains the putative origin of replication. Arrows, 19-bp iteron unique to pBPs2; arrows between brackets illustrate the copy number of the same iteron in the 1.70-kb PCR fragment; open rectangle, 19-bp element differing in one nucleotide from consensus sequence of RepA/C iterons; triple strip, 129-bp sequence identical to the 3' end of *trpE*.

TATAAAAAAAAA-3'), located 30 nt upstream of the stop codon TAA (Fig. 4B). In the smaller of the two PCR products, this element occurred at exactly the same position within the coding sequence but was directly followed by three additional copies, the second of which contained the stop codon (TAA) of *repAC*. The 1.7-kb PCR product contained 12 copies of the element. Since the third copy again included the stop codon, the *repAC* coding sequences of the two PCR products were identical, and this variant of the gene is designated *repAC*₂. The presence of a repeated 19-bp element resembles the iterons found in the origin of replication of the RepA/C replicon of plasmid RA1 (34). The difference of eight copies of this element in the region downstream of *repAC*₂ explains the size difference of about 160 bp between the two PCR products. This length variation most probably reflects a heterogeneous plasmid population within our DNA preparation (see below).

The total length of the three assembled contiguous fragments, including the sequence derived from the smaller, 1.53-kb PCR product, is 5,477 bp (Fig. 4B). The sequence has a G+C content of 25.6 mol% and 85% is occupied by putative ORFs. In contrast to pBTc2, putative -35 (TTGAC) and -10 (AGTTAA) promoter elements could be identified, 125 bp upstream of *trpE*. The elements are separated by 16 bp and resemble previously identified *Buchnera* promoters (32). A second possible -10 box (TAACATA), which is identical to the *E. coli trp* -10 sequences (33), was found overlapping 3 bp with the first. The intergenic region between *repAC*₁ and *trpG* contained a 129-nt sequence that was identical to the 3' end of *trpE*. Similar "remnants" of *trpE* have been described for the *trpEG* plasmid of *B. aphidicola* (*Uroleucon sonchi*) (4). The intergenic region between *repAC*₂ and *trpE* was highly AT rich (85 mol%) and contained one copy of a 19-bp element (5'-TATATGGGAATGTTGCACA-3') that differs in only one position from the consensus sequence (5'-AT-TGGG-G-TG-CACG-3') of a 13-copy iteron described for the origin of replication of *E. coli* plasmid RA1 (Fig. 4B) (34, 35). In addition, *ori* of this replicon contains one copy of a DnaA box. This element was absent from our sequence, but it may not be required for replication (40). The high A+T content and similarity with respect to the presence of iterons suggest that the *repAC*₂-*trpE* intergenic region contains the origin of replication of pBPs2.

Structure of pBPs2. To establish the overall structure of the plasmid, we performed the following experiments. First, quantitative hybridizations were carried out to determine the relative abundance of the two *XbaI* fragments detected in the initial hybridizations (Fig. 3). Densitometric assays of Southern blots containing *XbaI* digested plasmid DNA indicated that the 1.8-kb fragment was 5.19 ± 0.97 times more abundant than the 2.1-kb fragment (data not shown). A copy number of 5 or 6 of the 1.8-kb fragment relative to one copy each of the 2.1- and 1.5-kb fragments adds up to a plasmid size of either 12.8 or 14.6 kb, which was in good agreement with the initial estimations. Second, partial *XbaI* digestions of plasmid DNA predigested with *AccI* plus *SacI* resulted in a ladder of bands in which each step consisted of two bands differing 200 bp in size (Fig. 5A). The size difference between the double steps corresponded exactly to multiples of 1.8 kb plus 1.6 kb and multiples of 1.8 kb only. These findings were in agreement with the expectation based on the restriction site map (Fig. 4A), the 1.6-kb fragment resulting from the *SacI* site in the 2.1-kb *XbaI* fragment.

Control digestions with *AccI* and *SacI* revealed that the plasmid population was in fact heterogeneous and was composed of molecules of different sizes. A single digestion with *SacI*, which linearizes the plasmid, yielded a short ladder of bands, differing in size by steps of 1.8 kb (Fig. 5A). The two most strongly hybridizing bands were 12.8 and 14.6 kb, corresponding to molecules containing either five or six copies of the 1.8-kb *XbaI* repeat, respectively. Four more weakly hybridizing bands measured approximately 18.2, 16.4, 11.0, and 9.2 kb, corresponding to molecules containing 8, 7, 4, and 3 copies of the same unit. A similar ladder of bands was observed for *AccI*, although all the bands were 1.3 kb smaller, which is consistent with the presence of two sites, 1.3 kb apart, in the single-copy region of the plasmid (Fig. 4A). Superimposed on the length variation due to differences in repeat number is the length variation accounted for by the difference in copy number of the 19-bp direct repeat downstream of *repAC*₂. The inferred overall structure of a plasmid containing five tandem repeats of the 1.8-kb *XbaI* unit is presented in Fig. 5B.

Similarity between pBPs2 *repAC* and *B. aphidicola* (*Rhopalosiphum*) *trpEG* plasmids. The single-copy region between *repAC*₂ and *trpE* of pBPs2 was found to share similarities with

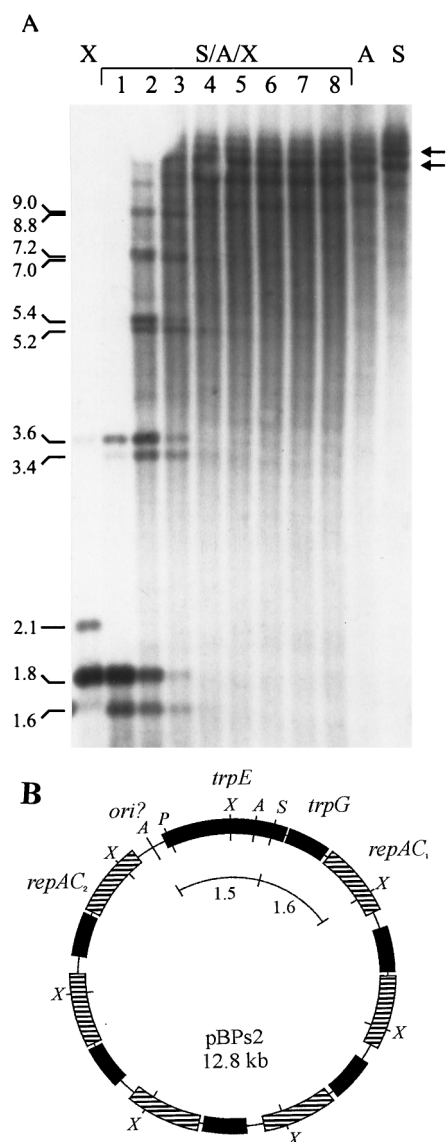


FIG. 5. (A) Southern blot analysis of partial digestion of *AccI*-*SacI*-digested *B. aphidicola* (*P. spyrothecae*) DNA by using serial dilutions of *XbaI*. Lanes: 1 to 8, digestions with 5.0, 2.5, 1.24, 0.63, 0.31, 0.16, 0.08, and 0.04 U of *XbaI*, respectively, for 15 min. X, A, S, control digestions performed to completion (X, *XbaI*; S, *SacI*; A, *AccI*). Arrows indicate the two principal forms of the plasmid. (B) Proposed structure of pBPs2 from *B. aphidicola* (*P. spyrothecae*) containing five tandem repeats of a 1.8-kb unit. Boxes with a similar pattern represent identical genes. All genes are transcribed in a clockwise direction. The region denoted *ori?* contains the putative origin of replication. Arcs correspond to restriction fragments removed from molecules with *AccI* and *SacI* before partial digestion experiment with *XbaI*. The restriction enzyme sites shown are as in Fig. 4A.

the origin of replication of plasmid RA1, but no significant similarities to the 500- to 700-nt putative *ori* of pBTc2 or the *B. aphidicola* (Aphididae) *trpEG* plasmids were observed. However, a comparison of the *repAC* sequence to the latter group of plasmids revealed a striking similarity (73% identical positions in a 955-bp overlap) to the region immediately upstream of *ori*-3.6 of the *B. aphidicola* (*R. maidis*) plasmid (Fig. 1B). Analysis of the recent update of this sequence (provided by P. Baumann) showed that this plasmid in fact carries a potentially intact *repAC* gene, whose deduced amino acid se-

quence was 71% identical to the two pBPs2 *RepAC* sequences and 64% identical to the plasmid RA1 homologue (Fig. 6). In addition, the 19-bp element (5'-TATATGGGAATGTTGCA CA-3') present in a single copy in the putative *ori* of pBPs2 and in 13 copies in the *ori* of RA1 (34, 35) also occurs in the *ori* of *B. aphidicola* (*R. maidis*) plasmid, with only a 1-bp difference from the pBPs2 sequence.

A subsequent comparison of *B. aphidicola* (*R. maidis*) *repAC* to the region upstream of *ori*-3.6 from the closely related *B. aphidicola* (*R. paid*) *trpEG* plasmid yielded a similarity of 71% in an 843-bp overlap. The latter sequence, however, did not contain an uninterrupted reading frame. A putative remnant of the 19-bp element (5'-TATACGAGAATAGTGCAC A-3') was also found in exactly the same position as in the *B. aphidicola* (*R. maidis*) sequence. The similarity of *repAC* to the upstream region of *ori* in *trpEG* plasmids from other species was much lower (44 to 59%), and the 19-bp element or remnants thereof could not be identified. Nevertheless, the high sequence similarity in *B. aphidicola* (*Rhopalosiphum*) suggests that at least the *B. aphidicola* (*R. padi*) sequence contains an *repAC* pseudogene. This mode of gene silencing has a well-documented precedent in the *trpEG* plasmids of *Buchnera* from the aphids *Diruaphis noxia* (32) and *Uroleucon sonchi* (4). Both plasmids carry only one functional copy of *trpEG*, in addition to a variable number of repeat units containing *trpEG* or *trpE* pseudogenes.

The multiple alignment further illustrates differences among the termini of the RepA sequences (Fig. 6). RepA from plasmid RA1 has an additional 33 residues at its N-terminal end, while the *B. aphidicola* (*P. spyrothecae*) sequences are longer at their C-terminal end. Remarkably, however, a high similarity to the RA1 sequence is retained at the nucleotide level after the stop codon, in the second reading frame of the RA1 sequence. The similarity drops immediately after the stop codon of the pBPs2 *repAC* sequences. Because it is unlikely that *Buchnera* would have retained such a high similarity in a non-coding sequence, it is possible that the published RA1 sequence contains a sequencing error. A reestimation of the size of RepA in the sodium dodecyl sulfate-polyacrylamide gel electrophoresis analysis of Fig. 2 of Llanes et al. (35) yielded a molecular mass of 42 kDa, in contrast to the 35-kDa estimate reported by the authors. Our estimate corresponds exactly to a RepA protein of 366 residues (Fig. 6).

TrpEG phylogenetic relationships. A phylogenetic analysis of *Buchnera* TrpE and TrpG sequences was conducted to assess the relationships of the newly sequenced genes. In accordance with previous 16S rDNA phylogenies (39, 49), the TrpG sequences from the *B. aphidicola* (Pemphigidae) form a cluster (Fig. 7). In the TrpE analysis, the outgroup sequences (*E. coli* and *Salmonella typhimurium*) rooted the tree within the *B. aphidicola* (Pemphigidae). The branch lengths of the groupings around the root were relatively short, particularly in the TrpE phylogeny, while the longer branches leading to the *B. aphidicola* (*T. caerulescens*) sequences indicate that the substitution rate in this lineage has been higher than in any of the other lineages.

DISCUSSION

Recent genetic studies have substantiated the importance of essential amino acid biosynthesis by *B. aphidicola* in its symbiotic association with aphids (7, 10, 30, 49). To date, rearrangements in the pathways leading to tryptophan and leucine remain the sole genetic modifications found in *Buchnera* that are indicative of a specific, enhanced biosynthetic capability (5-7, 12). However, amplification of the genes (*trpEG*) encoding

of one 3.2-kb unit encoding a functional copy of *TrpEG* and a variable number of repeat units carrying *trpEG* pseudogenes (4, 32). The plasmid population in *B. aphidicola* (*T. caerulescens*) was composed mainly of molecules containing a single 3.0-kb *trpEG* unit, but Southern hybridization indicated the presence of multimers (Fig. 2).

The molecular basis of concatenation in these plasmids is unknown, but it is likely to be initiated by multimerization, which involves homologous recombination between plasmid monomers (25, 46). An indication that recombination (and mismatch repair) occurs in the *trpEG* plasmids comes from the observation by Lai et al. of gene conversion among *trpEG* pseudogenes (32). Multimerization of low-copy-number plasmids increases the chance of their loss at cell division (46). Several maintenance systems are known to counteract this risk, including multimer resolution and partitioning systems (46, 51). There is no evidence for the presence of such systems on the *trpEG* plasmids. However, stable inheritance may not be essential if selection for *trpEG* copy number does not act primarily on individual cells. In its intracellular environment, the slowing dividing *Buchnera* (3) may be viable with a reduced number of *trpEG* plasmids or even with none. This could shift the need for plasmid stability in individual cells to the need for partitioning bacteria with an adequate genetic composition to the next generation of aphids. We speculate that selection acting at this higher level underlies the observed structural variability of *Buchnera trpEG* plasmids.

Phylogenetic analysis of *TrpEG*. The fact that pBTc2 was more similar to the *B. aphidicola* (Aphididae) *trpEG* plasmids than to pBP2s raised the possibility that *B. aphidicola* (*T. caerulescens*) acquired its plasmid through horizontal transfer. Phylogenetic analysis, however, showed that the plasmid-borne sequences of *B. aphidicola* (*T. caerulescens*) and *B. aphidicola* (*P. spyrothecae*) are most closely related to the chromosomal *TrpEG* sequence of *B. aphidicola* (*S. chinensis*) (Fig. 7). This refutes horizontal acquisition of pBTc2 by *B. aphidicola* (*T. caerulescens*). The slight incongruence at the root between the *TrpE* and *TrpG* topologies can be attributed to an accelerated substitution rate in *B. aphidicola* (*T. caerulescens*), causing its long branch to be "artificially attracted" (24) by the *B. aphidicola* (Aphididae) sequences in the case of the *TrpE* phylogeny.

Replicons in *Buchnera trpEG* plasmids. Two replicons are possibly linked to the amplification of *trpEG*: (i) a RepA/C-like replicon, in which replication is most probably initiated by plasmid-encoded RepAC, and (ii) an *ori-3.6*-like replicon, which is characterized by the presence of multiple DnaA boxes in the *ori* and for which DnaA is thought to be involved in initiation of replication (30, 42, 43). After the discovery of RepA/C in *B. aphidicola* (*P. spyrothecae*), computer analysis of previously characterized *trpEG* plasmids (42) revealed the presence of a putative intact *repAC* gene in *B. aphidicola* (*R. maidis*) and a *repAC* pseudogene in *B. aphidicola* (*R. padi*) (Fig. 1B and 6). In addition, *ori-3.6* of both plasmids contained a single copy of the 19-bp iteron of the RepA/C replicon. None of these elements could be identified unequivocally in any of the other *B. aphidicola* (Aphididae) plasmids or in pBTc2. The phylogenetic distribution of the two replicons is summarized in Fig. 8.

The presence of conserved elements of both replicons in the *B. aphidicola* (*R. maidis*) plasmid and a *repAC* pseudogene in its closest relative *B. aphidicola* (*R. padi*) demonstrates that *repAC* can be silenced in the presence of an *ori-3.6*-like replicon. The silenced *repAC* gene of *B. aphidicola* (*R. padi*) apparently diverged rapidly from the *B. aphidicola* (*R. maidis*) sequence through the accumulation of substitutions (only 71% similarity at the nucleotide level). This observation, together

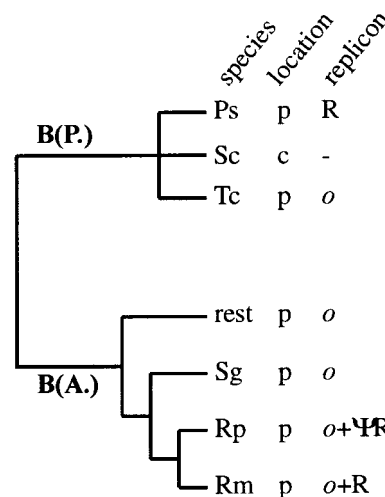


FIG. 8. Phylogenetic distribution of *trpEG*-associated replicons in *Buchnera* from aphids of the families Pemphigidae [B(P.)] and Aphididae [B(A.)]. Species designations are as in Fig. 7. Location: p, plasmid; c, chromosomal. Replicon designation: R, RepA/C; ΨR, RepAC pseudogene; o, *ori-3.6* like.

with the fact that the noncoding regions between *trpG* and *ori-3.6* are often unusually long [up to 1 kb in *B. aphidicola* (*A. pisum*) (Fig. 1)], raises the possibility that a RepA/C-like replicon was ancestral to all *B. aphidicola* (Aphididae) plasmids but that the respective *repAC* genes have been silenced in parallel lineages and have diverged beyond recognition.

Evolution of *trp* in *Buchnera*. Lai et al. (32) have previously proposed a scenario for the evolution of *trp* in *Buchnera*. In brief, they hypothesized that the ancestor of all present-day *Buchnera* species had the *trp* genes arranged in two linkage groups [*trpEG* and *trpDC(F)BA*] on its chromosome, an organization still found in *B. aphidicola* (*S. chinensis*) (31). *trpEG* was relocated to a plasmid in the ancestor of the *B. aphidicola* (Aphididae), and the resulting enhanced capacity of tryptophan production promoted increased growth and reproductive rates of the insect host. This scenario must be amended in the light of two significant findings made in the present study. Firstly, *trpEG* plasmids are found in *Buchnera* species from the two most divergent lineages of aphids, i.e., the Aphididae and Pemphigidae. This suggests that selection for amplification of *trpEG* in *Buchnera* may be as ancient as the symbiosis itself. Second, similar replicons are found in both lineages (Fig. 8). On the basis of these findings, we hypothesized that the transfer of *trpEG* to a plasmid carrying a RepA/C-like replicon occurred in the ancestor of all present-day *Buchnera* species. Like the related *E. coli* RA1 plasmid, the origin of replication may have contained a single DnaA box. An *ori-3.6* like origin of replication evolved de novo on this plasmid through amplification of the resident DnaA box. (Note that, in contrast to the RepA/C-like replicon and the *repA*, replicon linked to leucine plasmids [49], there are no reports on bacterial replicons related to the *ori-3.6*-like replicon.) De novo evolution of *ori-3.6* must have occurred independently in the lineages leading to *B. aphidicola* (*T. caerulescens*) and *B. aphidicola* (Aphididae) but not in *B. aphidicola* (*P. spyrothecae*). In the presence of *ori-3.6* and its hypothesized involvement in a regulatory mechanism of initiation of chromosome replication (32), control over replication of the *trpEG* plasmid was more advantageously exerted by the DnaA protein than by RepAC. This led to silencing of the ancestral RepA/C replicon in most lineages. Although it has yet to be established whether *repAC* is functional in *B. aphidicola* (*R. maidis*), it remains difficult to explain

why the elements of RepA/C are much more highly conserved in this species than in any of the other *B. aphidicola* (Aphididae) strains.

Our scenario implies that *trpEG* in the lineage leading to *B. aphidicola* (*S. chinensis*) must have been transferred back into the chromosome (Fig. 8) rather than that its organization represents the ancestral state (31). In *E. coli*, the closest-known relative of *Buchnera* (47), all the genes of the *trp* pathway are organized into a single operon (13). If a similar organization was present in the presymbiotic ancestor of *Buchnera*, then the proposed reversal in *B. aphidicola* (*S. chinensis*) provides an explanation for the organization of its *trp* genes into two separate linkage groups.

Finally, our scenario would predict that (i) *trpEG* plasmids, containing elements of either RepA/C or *ori-3.6* like replicons (or both), are likely to be found in *Buchnera* strains from other families of aphids and (ii) in lineages where *trpEG* is chromosomally encoded, its location is expected to be different from the one found in *B. aphidicola* (*S. chinensis*).

ACKNOWLEDGMENTS

We are indebted to Fernando González-Candelas for support and to the Servei de Bioinformàtica and the S.C.S.I.E. (Universitat de València) for providing computing and sequencing facilities, respectively. We gratefully thank J. M. Michelena for identification of the aphid species.

This work was supported by an EU HCM research grant (CHRX-CT94-0660) to R.V. and grant PB96-0793 C04-01 from DGES to A.M.

REFERENCES

- Altschul, S. F., W. Gish, W. Miller, E. W. Myers, and D. J. Lipman. 1990. Basic local alignment search tool. *J. Mol. Biol.* **215**:403–410.
- Ausubel, F. M., R. Brent, R. E. Kingston, D. D. Moore, J. G. Seidman, J. A. Smith, and K. Struhl. 1989. *Current protocols in molecular biology*. Green Publishing, New York, N.Y.
- Baumann, L., and P. Baumann. 1994. Growth kinetics of the endosymbiont *Buchnera aphidicola* in the aphid *Schizaphis graminum*. *Appl. Environ. Microbiol.* **60**:3440–3443.
- Baumann, L., M. A. Clark, D. Rouhakhsh, P. Baumann, N. A. Moran, and D. J. Voegtlin. 1997. Endosymbionts (*Buchnera*) of the aphid *Uroleucon sonchi* contain plasmids with *trpEG* and remnants of *trpE* pseudogenes. *Curr. Microbiol.* **35**:18–21.
- Baumann, P., and N. A. Moran. 1997. Non-cultivable microorganisms from symbiotic associations of insects and other hosts. *Antonie Leeuwenhoek* **72**: 39–48.
- Baumann, P., N. A. Moran, and L. Baumann. 1997. The evolution and genetics of aphid endosymbionts. *BioScience* **47**:12–20.
- Baumann, P., L. Baumann, Lai C. Y., D. Rouhakhsh, N. A. Moran, and M. A. Clark. 1995. Genetics, physiology, and evolutionary relationships of the genus *Buchnera*: intracellular symbionts of aphids. *Annu. Rev. Microbiol.* **49**:55–94.
- Blackman, R. L., and V. F. Eastop. 1984. *Aphids on the world's crops*. John Wiley & Sons, Ltd., Chichester, United Kingdom.
- Blackman, R. L., and V. F. Eastop. 1994. *Aphids on the world's crops*. CAB International, Wallingford, United Kingdom.
- Bracho, A. M., D. Martínez-Torres, A. Moya, and A. Latorre. 1995. Discovery and molecular characterization of a plasmid localized in *Buchnera* sp. bacterial endosymbiont of the aphid *Rhopalosiphum padi*. *J. Mol. Evol.* **41**: 67–73.
- Brough, C. N., and A. F. G. Dixon. 1990. Ultrastructural features of egg development in oviparae of the vetch aphid, *Megoura viciae* Buckton. *Tissue Cell* **22**:51–63.
- Clark, M. A., L. Baumann, and P. Baumann. 1998. Sequence analysis of a 34.7-kb DNA segment from the genome of *Buchnera aphidicola* (endosymbiont of aphids) containing *groEL*, *dnaA*, the *atp* operon, *gidA*, and *rho*. *Curr. Microbiol.* **36**:158–163.
- Crawford, I. P. 1989. Evolution of a biosynthetic pathway: the tryptophan paradigm. *Annu. Rev. Microbiol.* **43**:567–600.
- Dadd, R. H. 1985. Nutrition: organisms, p. 315–319. In G. A. Kerkut and L. I. Gilbert (ed.), *Comprehensive insect physiology, biochemistry and pharmacology*, vol. 4. Pergamon Press, Elmsford, N.Y.
- Devereux, J., P. Haerberli, and O. Smithies. 1984. A comprehensive set of sequence analysis programs for the VAX. *Nucleic Acids Res.* **12**:387–395.
- Douglas, A. E. 1989. Mycetocyte symbiosis in insects. *Biol. Rev.* **69**:409–434.
- Douglas, A. E. 1990. Nutritional interactions between *Myzus persicae* and its symbionts, p. 319–327. In R. K. Campbell and R. D. Eikenbary (ed.), *Aphid-plant genotype interactions*. Elsevier Biomedical Press, Amsterdam, The Netherlands.
- Douglas, A. E. 1993. The nutritional quality of phloem sap utilized by natural aphid populations. *Ecol. Entomol.* **18**:31–38.
- Douglas, A. E. 1996. Reproductive failure and the amino acid pools in pea aphids (*Acyrtosiphon pisum*) lacking symbiotic bacteria. *J. Insect Physiol.* **42**:247–255.
- Douglas, A. E. 1998. Nutritional interactions in insect-microbial symbioses: aphids and their symbiotic bacteria *Buchnera*. *Annu. Rev. Entomol.* **43**:17–37.
- Douglas, A. E., and W. A. Prosser. 1992. Synthesis of the essential amino acid tryptophan in pea aphid (*Acyrtosiphon pisum* symbiosis). *J. Insect Physiol.* **38**:565–568.
- Febvay, G., I. Liadouze, J. Guillaud, and G. Bonnet. 1995. Analysis of energetic amino acid metabolism in *Acyrtosiphon pisum*—a multidimensional approach to amino acid metabolism in aphids. *Arch. Insect Biochem. Physiol.* **29**:45–69.
- Felsenstein, J. 1993. *Phylogenetic Inference Package (PHILIP) version 3.5*. University of Washington, Seattle.
- Felsenstein, J. 1998. Phylogenies from molecular sequences: inference and reliability. *Annu. Rev. Genet.* **22**:521–565.
- Gubathakurta, A., I. Viney, and D. Summers. 1996. Accessory proteins impose site selectivity during ColE1 dimer resolution. *Mol. Microbiol.* **20**: 613–620.
- Hinde, R. 1971. Maintenance of aphid cells and the intracellular symbionts of aphids in vitro. *J. Invertebr. Pathol.* **17**:333–338.
- Houk, E. J., and G. W. Griffiths. 1980. Intracellular symbionts of the Homoptera. *Annu. Rev. Entomol.* **24**:161–187.
- Ishikawa, H. 1987. Nucleotide composition and kinetic complexity of the genomic DNA of an intracellular symbiont of the pea aphid *Acyrtosiphon pisum*. *J. Mol. Evol.* **24**:205–211.
- Kornberg, A., and T. A. Baker. 1992. *DNA replication*, 2nd ed. W. H. Freeman & Co., New York, N.Y.
- Lai, C.-Y., L. Baumann, and P. Baumann. 1994. Amplification of *trpEG*: adaptation of *Buchnera aphidicola* to an endosymbiotic association with aphids. *Proc. Natl. Acad. Sci. USA* **91**:3819–3823.
- Lai, C.-Y., P. Baumann, and N. A. Moran. 1995. Genetics of the tryptophan biosynthetic pathway of the prokaryotic endosymbiont (*Buchnera*) of the aphid *Schlechtendalia chinensis*. *Insect Mol. Biol.* **4**:47–59.
- Lai, C.-Y., P. Baumann, and N. A. Moran. 1996. The endosymbiont (*Buchnera* sp.) of the aphid *Diuraphis noxia* contains plasmids consisting of *trpEG* and tandem repeats of *trpEG* pseudogenes. *Appl. Environ. Microbiol.* **62**: 332–339.
- Lisser, S., and H. Margalit. 1993. Compilation of *E. coli* mRNA promoter sequences. *Nucleic Acids Res.* **21**:1507–1516.
- Llanes, C., P. Gabant, M. Couturier, and Y. Michel-Briand. 1994. Cloning and characterization of the Inc A/C plasmid RA1 replicon. *J. Bacteriol.* **176**: 3403–3407.
- Llanes, C., P. Gabant, M. Couturier, L. Bayer, and P. Plesiat. 1996. Molecular analysis of the replication elements of the broad-host-range RepA/C replicon. *Plasmid* **36**:26–35.
- Marchuk, D., M. Drumm, A. Saulino, and F. S. Collins. 1992. Construction of T-vectors, a rapid general system for direct cloning of unmodified PCR products. *Nucleic Acids Res.* **19**:1154.
- Moran, N. A., M. A. Munson, P. Baumann, and H. Ishikawa. 1993. A molecular clock in endosymbiotic bacteria is calibrated using the insect hosts. *Proc. R. Soc. London Ser. B* **243**:167–171.
- Munson, M. A., and P. Baumann. 1993. Molecular cloning and nucleotide sequence analysis of a putative *trpDC(F)BA* operon in *Buchnera aphidicola* (endosymbiont of the aphid *Schizaphis graminum*). *J. Bacteriol.* **173**:6321–6324.
- Munson, M. A., P. Baumann, M. A. Clark, L. Baumann, and N. A. Moran. 1991. Evidence for the establishment of aphid-eubacterium endosymbiosis in an ancestor of four aphid families. *J. Bacteriol.* **173**:6321–6324.
- Nieto, C., R. Giraldo, E. Fernandez-Tresguerres, and R. Diaz. 1992. Genetic and functional analysis of the basic replicon of pPS10, a plasmid specific for *Pseudomonas* isolated from *Pseudomonas syringae* pathovar savastanoi. *J. Mol. Biol.* **223**:415–426.
- Raven, J. A. 1983. Phytophages of xylem and phloem: a comparison of plant sap-feeders. *Adv. Ecol. Res.* **13**:136–234.
- Rouhakhsh D., C.-Y. Lai, C. von Dohlen, M. A. Clark, L. Baumann, P. Baumann, N. A. Moran, and D. J. Voegtlin. 1996. The tryptophan biosynthetic pathway of aphid endosymbionts (*Buchnera*): genetics and evolution of plasmid-associated anthranilate synthase (*trpEG*) within the Aphididae. *J. Mol. Evol.* **42**:414–421.
- Rouhakhsh D., M. A. Clark, L. Baumann, N. A. Moran, and P. Baumann. 1997. Evolution of the tryptophan biosynthetic pathway in *Buchnera* (aphid endosymbionts): studies of plasmid-associated *trpEG* within the genus *Uroleucon*. *Mol. Phylogenet. Evol.* **8**:167–176.
- Sambrook, J., E. F. Fritsch, and T. Maniatis. 1989. *Molecular cloning: a*

- laboratory manual. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.
45. **Sasaki, T., H. Hayashi, and H. Ishikawa.** 1991. Growth and reproduction of symbiotic and aposymbiotic pea aphids, *Acyrtosiphon pisum*, maintained on artificial diets. *J. Insect Physiol.* **37**:85–92.
 46. **Summers, D. K.** 1991. The kinetics of plasmid loss. *Trends Biotechnol.* **9**: 273–278.
 47. **Untermann, B. M., P. Baumann, and D. L. McLean.** 1989. Pea aphid symbiont relationships established by analysis of 16S rRNAs. *J. Bacteriol.* **171**: 2970–2974.
 48. **van Ham, R. C. H. J., and A. Latorre.** Unpublished data.
 49. **van Ham, R. C. H. J., A. Moya, and A. Latorre.** 1997. Putative evolutionary origin of plasmids carrying the genes involved in leucine biosynthesis in *Buchnera aphidicola* (endosymbiont of aphids). *J. Bacteriol.* **179**:4768–4777.
 50. **Viswanathan, V. K., J. M. Green, and B. P. Nichols.** 1995. Kinetic characterization of 4-amino-4-deoxychorismate synthase from *Escherichia coli*. *J. Bacteriol.* **177**:5918–5923.
 51. **Williams, D. R., and C. M. Thomas.** 1992. Active partitioning of bacterial plasmids. *J. Gen. Microbiol.* **138**:1–16.