New Strategies for Cultivation and Detection of Previously Uncultured Microbes

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An integrative approach was used to obtain pure cultures of previously uncultivated members of the divisions Acidobacteria and Verrucomicrobia from agricultural soil and from the guts of wood-feeding termites. Some elements of the cultivation procedure included the following: the use of agar media with little or no added nutrients; relatively long periods of incubation (more than 30 days); protection of cells from exogenous peroxides; and inclusion of humic acids or a humic acid analogue (anthraquinone disulfonate) and quorum-signaling compounds (acyl homoserine lactones) in growth media. The bacteria were incubated in the presence of air and in hypoxic (1 to 2% O2 [vol/vol]) and anoxic atmospheres. Some bacteria were incubated with elevated concentrations of CO2 (5% [vol/vol]). Significantly more Acidobacteria were found on isolation plates that had been incubated with 5% CO2. A simple, high-throughput, PCR-based surveillance method (plate wash PCR) was developed. This method greatly facilitated detection and ultimate isolation of target bacteria from as many as 1,000 colonies of nontarget microbes growing on the same agar plates. Results illustrate the power of integrating culture methods with molecular techniques to isolate bacteria from phylogenetic groups underrepresented in culture.

Cultivation-independent molecular techniques have illuminated the enormous microbial diversity that exists on our planet and have served to define nearly 40 phylum-level divisions within the Bacteria domain alone (23). Most of these divisions, however, are poorly represented by cultured organisms, and at least 13 remain candidate divisions represented only by environmental gene sequences (23). The Acidobacteria and Verrucomicrobia divisions are among those divisions of the domain Bacteria represented by a large diversity of 16S rRNA genes, which occur in particular abundance in soils, but contain few cultured members (3, 11, 12, 17, 20, 21, 23, 36, 48). Hence, our appreciation of the physiological diversity of Acidobacteria and Verrucomicrobia is limited, as is our knowledge of their role in global biogeochemical cycles. Clearly, a better understanding of these divisions would be attained by having a greater diversity of their members available in pure culture for detailed study.

The intrinsic selectivity of any given medium and incubation condition imposes limits on the nature, number, and diversity of microbes recovered from natural samples. It follows, then, that the application of isolation procedures that better mimic conditions existing in the habitat from which the samples were obtained could increase the likelihood of retrieving previously uncultured organisms. Recent efforts to accomplish this have met with some success by using the following: (i) relatively low concentrations of nutrients (1, 13–15, 19, 43, 45, 50); (ii) nontraditional sources of nutrients, signaling molecules, or inhibitors (of undesired organisms) (9, 10, 13, 31); and (iii) relatively lengthy periods of incubation (19, 22, 24–26, 33, 39, 40), sometimes directly in the natural environment from which the inoculum was obtained (26).

For soil microbes, some of which may have become adapted to elevated concentrations of CO2 and concentrations of O2 lower than the atmospheric O2 concentrations (38), the composition of the incubation atmosphere may be an important consideration. Elevated CO2 concentrations are rarely used in incubation atmospheres for isolation of soil microorganisms, yet CO2 could be important for metabolic processes other than pure autotrophy. Likewise, for soil microbes, the transition to fully aerobic conditions on plating in air may be a stressful event. This would be especially true if cells were not immediately equipped to cope with reactive oxygen species such as hydrogen peroxide (H2O2), superoxide (O2·−), or hydroxyl radical (OH·) produced by their own metabolism or present in media as a result of autoclaving (reviewed in reference 29). Even with facultative anaerobes such as Escherichia coli, an abrupt transition of anaerobically grown cells to aeration can severely retard growth of certain mutants (27). It is also noteworthy that the cultivability (in air) of E. coli and Vibrio vulnificus after starvation is greatly improved if plating media are supplemented with catalase or pyruvate, two compounds known to eliminate H2O2 (5, 34). Such observations suggest that incubation atmospheres enriched with CO2 and/or limited in O2, as well as the incorporation of agents to detoxify reactive oxygen species in the plating media, should be included among treatments seeking to recover previously uncultured microbes.

Whatever cultivation approach is tried, however, one is ultimately confronted with the need to evaluate its success. This is a potentially arduous task if, as in this study, many different media and incubation conditions are being tested and little or nothing is known about the microbes sought other than their 16S rRNA gene sequences. Accordingly, some high-through-

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put screening method is desirable. To deal with this, we developed a simple, high-throughput, PCR-based procedure, plate wash PCR (PWPCR), that facilitated the surveillance of isolation plates for the presence of target organisms and the ultimate recognition of colonies made up of target organisms. The results of this endeavor constitute the substance of the present paper.

**MATERIALS AND METHODS**

**Sample collection and manipulation.** Soil samples were collected between August 2001 and October 2002 from the Long Term Ecological Research (LTER) site located at the Michigan State University W. K. Kellogg Biological Station (KBS) in Hickory Corners, Mich. The KBS-LTER site includes a large-scale replicate field experiment with treatments representing different cropping systems and types of management, several successional sites, and unmanaged forested sites (http://www.lter.kbs.msu.edu). Soil core (2-cm diameter) samples (to a depth of 10 cm) were taken from each of five permanent sampling stations distributed across one of four replicate fields (replicate 1) of the never cultivated successional treatment, which is representative of “native” soil. Soil core wells were stored at 4°C (usually for less than 48 h) until they were homogenized under a hypoxic, CO2-enriched atmosphere (2% O2, 5% CO2, 93% N2) contained within a flexible vinyl hypoxic chamber fitted with an oxygen sensor and controller (Coy Laboratory Products, Grass Lake, Mich.). Approximately 30 g of soil was added to 100 ml of phosphate-buffered saline (pH 7.0) containing 224 mM sodium pyrophosphate as a dispersal agent and 1 mM dithiothreitol as a reducing agent (47). The suspension was stirred vigorously for 30 min and allowed to settle for 30 min. An aliquot of the supernatant was serially diluted in the same buffer and spread onto various media with at least three replicate plates per dilution.

Termite, *Reticulitermes flavipes* (Kollar) (Rhinotermitidae), were collected near Dansville, Mich., and either used immediately or maintained in the laboratory as described previously (8, 35). The guts from 25 to 50 worker larvae were extracted under a hypoxic atmosphere (described above) with sterile forceps and pooled in a glass tissue homogenizer containing 2 ml of a sterile basal salts solution based on the freshwater medium of Widdel and Bak (49). The basal salts solution contained the following (per liter): KH2PO4, 0.2 g; NH4Cl, 0.25 g; KCl, 0.5 g; CaCl2·2H2O, 0.15 g; NaCl, 1.0 g; MgCl2·6H2O, 0.62 g; Na2SO4, 2.84 g; 5-(4,6-dichloro-2-yl)-aminobenzoic acid, 40 mg; D-(+)-biotin, 10 mg; nicotinic acid, 100 mg; calcium (D)-pantothenate, 50 mg; pyridoxamine dihydrochloride, 100 mg; and thiamine dihydrochloride, 100 mg (49).

All solutions were heat sterilized, except for the trace element and mixed vitamin solutions, which were passed through a 0.22-μm-pore-size filter and autoclaved. The mixed vitamin stock solution contained the following (per liter): 4-aminoquinolinate, 50 mg; 3-aminobenzoinic acid, 40 mg; 3-aminobenzoic acid, 10 mg; nicotinic acid, 100 mg; calcium (D)-pantothenate, 50 mg; pyridoxamine dihydrochloride, 100 mg; and thiamine dihydrochloride, 100 mg (49).

**Cultivation conditions and screening.** The basal medium used for cultivation of soil bacteria was a modification of the basal salts solution described above and contained the following (per liter): KH2PO4, 0.2 g; NH4Cl, 0.25 g; KCl, 0.5 g; CaCl2·2H2O, 0.15 g; NaCl, 1.0 g; MgCl2·6H2O, 0.62 g; Na2SO4, 2.84 g; HEPES (pH 6.8), 10 mM; trace element solution (discussed below); 1 ml; vitamin B12 solution (50 mg/liter); 1 ml; mixed vitamin solution (discussed below); 1 ml; and Bacto Agar (Becton Dickinson and Company, Franklin Lakes, N.J.), 15 g. The trace element solution contained the following (per liter): MgCl2·6H2O, 1.5 g; CoCl2·6H2O, 190 mg; MnCl2·4H2O, 100 mg; ZnCl2, 70 mg; H3BO3·6H2O, 6 mg; Na2MoO4·2H2O, 36 mg; NiCl2·6H2O, 24 mg; CaCl2·2H2O, 2 mg; and HCl (25% [vol/vol]), 10 ml (49). The mixed vitamin stock solution contained the following (per liter): 4-aminoquinolinate, 50 mg; D(+)-biotin, 10 mg; nicotinic acid, 100 mg; calcium (D)-pantothenate, 50 mg; pyridoxamine dihydrochloride, 100 mg; and thiamine dihydrochloride, 100 mg (49).

**FIG. 1. PWPCR method to detect growth and monitor isolation of targeted bacteria.** Of the three medium and incubation conditions shown in this diagram (conditions A, B, and C), growth of targeted bacteria (+) is represented only in condition C.
Watertown, Mass.) for the following amplification schedule: (i) 3 min at 95°C; (ii) 30 cycles, with 1 cycle consisting of 30 s at 95°C, 30 s at the annealing temperature shown in Table 1, and 45 s at 72°C; and (iii) 10 min at 72°C.

Preliminary experiments to determine the optimum PCR conditions with the Acidobacteria-targeting (Acd31F-1492R) and Verrucomicrobia-targeting (Ver53F-1492R) primer pairs were performed by using template DNA from Acidobacterium capsulatum (ATCC 51196) and Verrucomicrobiurn spinosum (ATCC 43997), respectively. Optimum reaction conditions were determined across a gradient of annealing temperatures (50 to 65°C) and MgCl₂ concentrations (1 to 2.5 mM) by using a PTC-200 DNA Engine gradient thermocycler (MJ Research, South San Francisco, Calif.). Determining the sensitivity of target gene detection was performed by the PCRIs with Acidobacteria-targeting primers and decreasing amounts of A. capsulatum DNA mixed with nontarget DNA (E. coli K-12) to yield 50 ng of total DNA per reaction mixture. Sensitivity was also determined by using the Verrucomicrobiurn-targeting primer set with decreasing amounts of genomic DNA from a termite-associated Verrucomicrobiurn diasse isolate TAV1 (described below) in a 1:2 mass ratio with E. coli K-12 DNA. Direct, group-specific PCR amplification of 16S rRNA genes in environmental samples was performed with group-specific primers (see above) and 50 ng of DNA from soil or from 50 termite guts. Genomic DNA was extracted using the Ultraclean soil or fecal DNA kits per the manufacturer (MoBio Laboratories, Carlsbad, Calif.).

PWPCR was simply the PCR in which template DNA was obtained from the aggregate of colonies present on an isolation plate (Fig. 1). To obtain template DNA from the aggregate of colonies present on an isolation plate, the surface of the agar medium was flooded with 2 ml of bead solution from the UltraTec soil or fecal DNA kits per the manufacturer's protocols (MoBio Laboratories, Carlsbad, Calif.).

PWPCR results were compared for each treatment and used in a chi-square test with Bonferroni corrections for comparison of overall cultivability or the cultivability of specific primers in control reaction mixtures run in the same plate. The ability of PWPCR to detect a colony of a target organism from many nontarget organisms was examined by performing PWPCR on isolated isolation plates. A laboratory collection of 26 different bacterial isolates obtained from KBS-LTER soil (various α-, β-, and γ-Proteobacteria, Bacillace spp. [pLumifermics], Arthrobacter spp. [pLumifermics], Cytophaga- and Flavobacterium-like strains [pLumifermics] and Cytophaga- and Flavobacterium-like strains [pLumifermics] were each inoculated onto three or four sites (94 total) on plates of R2A agar (Difco, Detroit, Mich.), a medium commonly used for isolating environmental heterotrophs. Some plates were also inoculated with V. spinosum (ATCC 43997) at one site, and then all plates were inoculated in air, at room temperature, and in the dark for 6 days. At the end of the incubation period, plates with (positive) and without (negative) V. spinosum were used for PWPCR individually, and after dilution of template DNA extracted from V. spinosum-positive plates, the plates were inoculated with DNA extracted from V. spinosum-negative plates.

PCR products were analyzed by electrophoresis of 5-μl samples of reaction mixtures on 1% agarose gels at 100 V in 0.5X Tris-borate-EDTA. PCR products were visualized by UV illumination after staining with 1% Gelstar nucleic acid stain (Cambrex, East Rutherford, N.J.), and images were captured by using a Kodak electrophoresis documentation and analysis system 290 (Eastman Kodak). Sequence analysis of PWPCR-amplified 16S rRNA genes from environmental samples, PWPCR, or bacterial isolates were cloned directly into E. coli using the plasmid vector pCR2.1 or pCR4.0 (TOPO TA cloning kit; Invitrogen). Restriction fragment length polymorphism analyses were used to identify commonly cultured and unique clones. The partial sequence of each clone was determined with Applied Biosystems cycle sequencing technology (Applied Biosystems, Foster City, Calif.), the 16S rRNA gene primer 531R (5′-TAC CGC GGC TGC TGG CAC-3′), and/or vector primers. Preliminary phylogenetic affiliation of each clone was determined by sequence comparison to the GenBank nucleotide database using BLAST (2) or to the Ribosomal Database Project II database using the sequence match tool (18). Nearly full-length sequence (at least fourfold coverage) of the 16S rRNA gene from isolates and selected clones was obtained by using primers complementary to the multiple cloning site of pCR2.1 or pCR4.0 (F2 and R4* [see Table 1]); Acd31F, Ver53F, and 1492R (Table 1); and 338F (5′-CTC CTA CGG GAC GGA GCA GGA GT-3′), 531R (above), 767F (5′-AGC AAA CAG GAT TAG ATA CAC TGG-3′), 810R (5′-GGC GTG GAC TTC CAG GGT ATC T-3′), and 1087F (5′-GGT TAA GTC GGG CAA GCA-3′) with the Applied Biosystems cycle sequencing technology and either an ABI Prism 3100 genetic analyzer or ABI Prism 3700 DNA analyzer (Applied Biosystems).

Contiguous sequences for each isolate were assembled with the Vector NTI software package (Informax). These sequences were inserted into and aligned against a 16S rRNA gene sequence database in the ARB software package (http://www.arb-home.de/) (32), along with any other available phylum-specific sequences (>500 nucleotides [nt]) from GenBank (http://www.ncbi.nlm.nih.gov), the Ribosomal Database Project II (http://rdp.cme.msu.edu/) (18), or our own environmental clones. Aligned Acidobacteria and Verrucomicrobia sequence lengths were more than 1,250 nt to generate phylogenetic trees using maximum-likelihood analysis based on 1,097 shared nucleotides for the Acidobacteria and 1,050 nucleotides for the Verrucomicrobia. The minimum evolution-ary distance method in PAUP* was used for bootstrap analyses of the same data (46).

**Treatment effects on cultivability.** In order to determine which, if any, treatments had a significant impact on overall cultivability or the cultivability of Acidobacteria from soil, colony counts (in CFU per gram [dry weight] of soil) and PWPCR results were compared for each treatment and used in a chi-square test for goodness of fit with Bonferroni’s error rate adjustment (37, 42). Colonies used to determine colony counts had a minimum diameter of 0.2 mm and were visible using a colony counter fitted with a 1.5× magnifying lens. For overall cultivability, the average colony count was used as the expected value and that for a particular treatment was used as the observed value. For Acidobacteria culti-vability, the expected value was the probability of detection using PWPCR among all treatments multiplied by the number of agar plates used for a given treatment, where as the observed value was the number of times Acidobacteria were detected for a particular treatment. A total of 63 treatments were screened for this analysis.

**Nucleotide sequence accession numbers.** Partial 16S rRNA gene sequences (ca. 1,400 bases) from isolates KBS89, TAA43, TAA48, TAA166, TAV1, TAV2, TAV3, and TAV4 have been deposited in the EMBL, GenBank, and DDBI nucleotide sequences database under accession numbers AY587227 through AY587234.

## Results and Discussion

**Specificities and sensitivities of PCR and PWPCR.** Only targeted 16S rRNA genes were amplified during PCR with group-specific primers in control reaction mixtures run in the presence of *E. coli* DNA or after PWPCR of a diverse collec-
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Amplification with the PCR mixture and as little as 16 pg of A. capsulatum DNA and 93.75 fg of V. spinosum DNA yielded a visible amplimer. The specificity of each primer set was also confirmed by sequence analysis of clones obtained after PCR amplification using soil or termite gut community DNA and after PWPCR of simulated or experimental isolation plates. Of more than 100 such clones examined, all corresponded to the 16S rRNA gene targeted by the primer pair.

By PWPCR, the equivalent of a single V. spinosum colony could be detected on plates from a background of at least 940 nontarget colonies composed of 26 different soil bacteria from six major phyllogenetic groups (Fig. 2). Considering the small amount of V. spinosum colony material relative to that of the other bacteria, it should be possible to detect colonies of targeted microbes among a much larger number of nontarget colonies of similar size. Restriction fragment length polymorphism analysis revealed that only Verrucomicrobia-specific ribosomal DNA was amplified despite the diversity of nonspecific DNA in each sample (data not shown).

**Treatments effects and isolation of Acidobacteria and Verrucomicrobia.** On the basis of direct microscopic counts, 1.41 × 10^9 ± 0.16 × 10^9 DTAF-stainable microbes (n = 3) were present in each gram (dry weight) of soil. In cultivation experiments, recoveries ranged from 4.0 × 10^7 to 9.7 × 10^7 CFU/g (dry weight) of soil, or roughly 4.0 to 7.0% of the total microbial community on the basis of direct counts. These recoveries were higher than the “1% or less” recoveries commonly cited but similar to those from other studies that have used low nutrient concentrations and long incubation times (19, 24). No single treatment significantly increased the overall recovery of soil bacteria (Fig. 3A), which suggests that the longer incubation times used for all experiments may be responsible for our higher recoveries of bacteria.

When PWPCR results were compiled from the same experiments, however, one treatment, used individually or in combination with other treatments, had a significant positive effect on the occurrence of soil Acidobacteria on plates: this was the presence of 5% CO_2 in incubation atmospheres (Fig. 3B). Incubating the media in atmospheres with 5% CO_2 resulted in a slight acidification (about half of a pH unit) and, therefore,
could also be responsible for the increase in cultivation of Acidobacteria. Incubation of plates under hypoxia or supplementation of media with catalase or acyl-HSLs also tended to elicit a greater occurrence of Acidobacteria, whereas supplementation of media with an organic nutrient mixture appeared to have the opposite effect. The addition of humic acids in the form of soil extract or the humic acid analogue AQDS had no apparent effect on the occurrence of Acidobacteria (data not shown). While the increased values seen with these latter treatments were not statistically significant in this study, they may ultimately prove to be so if examined individually and systematically in a large-scale experiment.

PWPCR-based identification of primary isolation plates containing Acidobacteria enabled us to make an informed selection of companion treatment plates from which to prepare subcultures for additional PCR-based screening (see Materials and Methods). Ultimately, soil Acidobacteria strain KBS89 was isolated from soil that was plated on basal medium supplemented with catalase, acyl-HSLs, and a mixture of organic carbon substrates (described above) and incubated in CO₂-enriched air.

A PWPCR-based strategy, similar to that used for the isolation of Acidobacteria from soil, was used for the isolation of previously uncultivated microbes from termite guts. The overall recovery of viable prokaryotes from termite guts (9.7% of the direct microscopic count) was marginally higher than that from soil, with an estimated 4.5 × 10⁵ CFU per gut equivalent. As with primary isolation plates from soil inocula, PWPCR revealed the presence of Acidobacteria on some of the plates, and by using analogous procedures, Acidobacteria strains TAA43, TAA48, and TAA166 were subsequently isolated on plates containing basal medium supplemented with yeast extract and peptone (0.1% [wt/vol] each) and incubated in CO₂-enriched air.

When Verrucomicrobia-specific primers were used, Verrucomicrobia were also detected by PWPCR on primary isolation plates of all compositions inoculated with termite gut homogenate and incubated in air and in a hypoxic atmosphere enriched with 5% CO₂. Media used for cultivation of termite-associated microorganisms contained yeast extract and peptone with or without acetate and/or catalase. From such plates, four termite gut Verrucomicrobia (strains TAV1 through
TAV4) were isolated, assisted by PWPCR surveillance of subcultures. TAV1 and TAV2 were isolated from plates containing basal medium with yeast extract, peptone, and acetate. TAV3 and TAV4 were isolated from the same medium without acetate. All TAV isolates were obtained from plates incubated in CO₂-enriched air.

Properties of Acidobacteria and Verrucomicrobia isolates. All of the Acidobacteria isolates belong to subdivision 1 of the Acidobacteria (Fig. 4) (23). On the basis of 16S rRNA gene sequence similarity, the nearest cultivated relatives to strains KBS89 and TAA166 are Ellin351 (97%) and Ellin337 (98%), respectively (49). TAA166 is the nearest cultivated relative to strains TAA43 and TAA48 (96.1%), which possess identical 16S rRNA sequences. All Acidobacteria isolates are short rods (0.5 by 1 µm) that divide by binary fission and form slightly opaque colonies after 4 or 5 days, which reach a maximum diameter of 1 mm in 14 to 16 days. Soil Acidobacteria isolate KBS89 and, to a lesser extent, the termite gut Acidobacteria isolates produce copious amounts of an extracellular (apparently capsular) material (Fig. 4), which made colonies hard to disrupt and was presumably responsible for their flocculent growth in liquid cultures.

The termite-associated Verrucomicrobia isolates (TAV1 to TAV4) belong to subdivision 4 of the phylum Verrucomicrobia (Fig. 5A) (23). On the basis of 16S rRNA gene sequence similarity, the nearest cultivated relative to TAV1 is Opitutus terrae strain VeSm13 (94.2%). Strains TAV2, TAV3, and TAV4 have 16S rRNA gene sequences virtually identical to each other, and the nearest cultivated relative to these isolates is Opitutus terrae strain PB90-1 (93%). TAV1 shares only 92.7% sequence similarity to the other TAV isolates. All of the termite-associated Verrucomicrobia isolates are facultative anaerobes, obtaining significantly higher population densities in liquid culture in CO₂-enriched air and in an hypoxic atmosphere than under a CO₂-enriched anoxic atmosphere. TAV cells are 0.25 to 0.50 µm in diameter and occur almost exclusively in pairs (Fig. 5B). Additionally, TAV1 produces an abundance of extracellular (apparently capsular) material (Fig. 5C). The TAV isolates were detected on primary isolation plates after 30 days and on subculture plates after 14 days. On the original isolation media, they formed very small (<0.5 mm in diameter), white, round, mucoid colonies that were visible only with a dissecting microscope. After isolation and several passages in the laboratory, however, all TAV isolates formed larger colonies (2- to 4-mm diameter) in 2 to 5 days on R2A agar in air. Preliminary results from studying the distribution and abundance of these targeted phylogenetic groups suggest that Verrucomicrobia are autochthonous to the guts of R. flavipes and not allochthonous contaminants derived from soil, whereas the opposite is true for the Acidobacteria (J. T. Wertz, B. S. Stevenson, and J. A. Breznak, Abstr. 103rd Gen. Meet. Am. Soc. Microbiol. 2003, abstr. N-223, 2003).

Overview of the PWPCR-based isolation procedure. Given the variety of individual cultivation treatments and treatment combinations used in this study, as well as the various sources of inocula, the detection and isolation of Acidobacteria and Verrucomicrobia would have been extremely difficult without the PWPCR method. One of the most time-consuming aspects of any isolation procedure is the screening, selecting, and subculture of colonies from primary isolation plates, and if low nutrient conditions are used to prevent overgrowth by nondesired organisms, most colonies on such plates will be fairly
small. Indeed, colonies of the *Acidobacteria* and *Verrucomicrobia* strains isolated in this study would have been easily overlooked without the aid of a dissecting microscope. However, the PWPCR procedure economizes on time by directing one to treat plates known to contain the target organism(s). Hence, owing to its simplicity, utility, and relatively low cost, we anticipate that PWPCR will become widely used as an adjunct to creative approaches for isolation of novel, sought-after organisms. The only requirement is at least one specific and reliable primer in the pair of primers used for PCR.

As with any method, PWPCR also has some limitations. For PWPCR, the sought-after organisms must be capable of growth on plates solidified with agar (or an agar substitute) and also capable of being harvested from such plates. This would eliminate organisms that either cannot grow on solid media or that, like certain spirochetes (7) and spirilla (16), form largely subsurface colonies difficult to harvest by simple plate washing. However, the key element of PWPCR is PCR with a specific primer pair, so as long as sufficient cell material can be obtained to make a DNA template, either by harvesting cells from liquid cultures or removing subsurface colony material by coring, surveillance of cultures is possible. Thus, our results underscore the power of integrating various cultivation conditions with molecular biology to retrieve some of the “not-yet-cultured majority” of microbes on our planet (6, 39).

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REFERENCES

14. Fox, P. O., and D. R. E. Godfrey. 1970. Coring, surveillance of cultures is possible. Thus, our results underscore the power of integrating various cultivation conditions with molecular biology to retrieve some of the “not-yet-cultured majority” of microbes on our planet (6, 39).

REFERENCES

distributed soil bacteria from phylogenetic lineages previously only detected in cultivation-independent surveys. Environ. Microbiol. 4:654–666.