Acid-Adapted Strains of *Escherichia coli* K-12 Obtained by Experimental Evolution

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Enteric bacteria experience a wide range of pHs throughout the human intestinal tract: from pH 1.5 to 3.5 in the stomach, then a rise to pH 4 to 7 in the duodenum and pH 7 to 9 in the jejunum, and then a decline to pH 5 to 6 in the cecum (1, 2). *Escherichia coli* has evolved to grow over a relatively wide range of intestinal pHs from pH 4.5 to 9.0 (3–5) and to survive without growth for several hours at pH 2 (6–8; for reviews of acid stress and extreme-acid survival, see references 9 to 11). The ability to survive the low pH of the stomach—and enter rapid growth as the pH increases in the lower digestive tract—may contribute to the low infectious doses characteristic of enteric pathogens, such as *E. coli* O157:H7 (12).

The adaptations of *E. coli* to growth in mild acid (pH 5 to 6) have been studied previously (9–11). Growth in mild acid may be enhanced by components of metabolism and transport, such as the lysine and arginine decarboxylases (13, 14), and by cyclopropane fatty acid synthase (8, 15, 16). A transcriptomic study of growth at pH 5 versus pH 7 shows that acid upregulates proton pumps of the electron transport system and outer membrane proteins, the chemotaxis regulon for proton-driven motility, periplasmic acid chaperones *hdeA* and *hdeB*, the *mar* multidrug resistance regulon, oxidative stress genes, and numerous genes with unidentified functions (17).

Nevertheless, we still know little about how *E. coli* grows below pH 5, at the lower limit of its pH range, just above the range of nongrowth survival (pH 1.5 to 4.0). The range of pHs occurring during the transition from nongrowth to growth conditions is experienced by bacteria as they pass through the stomach and enter the intestine. Understanding growth in and adaptation to this lower range of growth pH is particularly important, because the gut microbiome may include hundreds of competing species (18, 19) and the ability of bacteria to outgrow their neighbors at pH 4 to 5 could influence which strains colonize the gut. The same pH range also occurs in many fruit juices vulnerable to bacterial contamination (20).

To investigate genes that impact growth below pH 5, we used the approach of experimental evolution (21, 22). In an evolution experiment, multiple replicates of a single microbial lineage are repeatedly diluted and cultured under specific regulated conditions (23, 24). If the specified environment deviates from the optimal conditions for peak fitness, then there exists strong selective pressure for mutant alleles that enhance fitness in the new environment. *E. coli* has been the subject of a number of evolution studies involving adaptation to high temperatures (25), high ethanol concentrations (26), growth in complex media (27), and resistance to antibiotics (28).

Experimental evolution offers several advantages for the study of bacterial stress responses (29, 30). First, the approach allows the researcher to tightly regulate the microbe’s environment for thou-
TABLE 1 Strains isolated after 2,000 generationsa

<table>
<thead>
<tr>
<th>Strain</th>
<th>Population</th>
<th>Isolate no.</th>
</tr>
</thead>
<tbody>
<tr>
<td>JLSE0079</td>
<td>H9</td>
<td>1</td>
</tr>
<tr>
<td>JLSE0080</td>
<td>H9</td>
<td>2</td>
</tr>
<tr>
<td>JLSE0083</td>
<td>B11</td>
<td>1</td>
</tr>
<tr>
<td>JLSE0084</td>
<td>B11</td>
<td>2</td>
</tr>
<tr>
<td>JLSE0091</td>
<td>F11</td>
<td>1</td>
</tr>
<tr>
<td>JLSE0092</td>
<td>F11</td>
<td>2</td>
</tr>
<tr>
<td>JLSE0137</td>
<td>F9</td>
<td>2</td>
</tr>
<tr>
<td>JLSE0138</td>
<td>F9</td>
<td>3</td>
</tr>
</tbody>
</table>

a The sequence data are available in the NCBI SRA under accession number SRP041420.

sands of generations, which allows multiple selective sweeps of potentially unpredicted mutations to spread through the population. The evolving populations can be frozen regularly and stored indefinitely, allowing the researcher to set up direct comparisons between the ancestral strain and the adapted strains. Whole-genome resequencing can then permit identification of mutations in genes relevant to acid stress. Such an analysis might reveal surprising mechanisms of the stress response that differ from those identified by traditional gene-knockout investigation. Finally, experimental evolution allows us to determine whether acid adaptation comes at the price of a fitness trade-off in other environments.

Previous experimental evolution studies of acid stress are limited. E. coli experimental evolution in moderate acid (pH 5.3) yields an acid-dependent fitness increase, but genome sequence analysis has not been used to identify the underlying genetic changes (31). In another study, populations that evolved for 1,000 generations with 20 rounds of exposure to pH 2.5 showed 20 new mutations, including mutations in etg alleles that relieve log-phase repression of extreme-acid resistance (32). Here we report on the isolation and characterization of eight E. coli strains from 4 of 24 populations cultured at pH 4.8 (for 730 generations) and then at pH 4.6 (2,000 generations in total). We found putative novel contributors to the acid stress response, including a strong selection for altered RNA polymerase.

MATERIALS AND METHODS

Bacterial strains and media. Escherichia coli K-12 W3110 was the ancestral strain of all acid-adapted populations (33). Additional strains derived from E. coli K-12 W3110 were isolated during the course of the experiment (Table 1). For competition assays assessing the relative fitness of ancestral and adapted strains, a lacZ:kan insertion allele was introduced into each strain by P1 phage transduction from Keio collection strain GI1886 (Coli Genetic Stock Center, Yale University) (34). The lacZ:kan insertion strains were maintained with 50 μg/ml kanamycin.

All bacterial cultures were grown in media based on LBK medium (10 g/liter tryptone, 5 g/liter yeast extract, 7.45 g/liter KCl) (17). The culture media were buffered with either 100 mM homopiperazine-N,N’-bis-2-(ethanolsulfonic acid) (HOMOPIPES; pKa = 4.55, 8.12), 100 mM 2-(N-morpholino)ethanesulfonic acid (MES; pKa = 5.96), or 100 mM 3-morpholinopropane-1-sulfonic acid (MOPS; pKa = 7.20), depending upon the desired pH range of 4.4 to 4.8, 5.5 to 6.5, or 7.0, respectively (17). The pKa values presented above are those found at 25°C. For media buffered to pH 4.5 to 4.8, the LBK medium was supplemented with 10 g/liter l-malic acid (pKa = 3.40, 5.11) (LBKmal). The pH of the medium was adjusted as necessary with either 5 M HCl or 5 M KOH. For all experiments, the pH of the medium was tested immediately before and after bacterial growth.

Experimental evolution. A protocol based on successful evolution protocols described in the literature (22, 25, 27) was devised for experimental evolution of E. coli growth at external pH values below 5. Twenty-four populations were established from the same parental freezer stock of strain W3110. Evolving populations (200 μl each) were maintained in 96-well plates in a microplate reader. In each plate, 24 evolving populations occupied three columns of eight wells, with the columns being separated by an empty column to minimize the possibility of cross contamination. The initial inoculation of each well was with a 1:100 dilution of an overnight culture of the ancestral strain W3110. Populations were evolved in LBKmal with 100 mM HOMOPIPES at pH 4.8 (with daily dilutions of 1:4,000) for approximately 730 generations. The number of generations was estimated using the assumption that a dilution of 1:4,000 allows 12 generations of binary fission per day. After generation 730, the pH of the dilution medium was adjusted to 4.6 and the populations evolved for a further 1,270 generations (daily dilution, 1:100). A dilution of 1:100 allows 100-fold growth daily, producing 6.64 generations. The well plate was maintained at 37°C in a SpectraMax Plus384 MicroPlate reader (Molecular Devices). For the initial 22 h of the incubation, the well plate was shaken for 3 s every 15 min, after which the optical density at 450 nm (OD450) of each population was recorded. After 22 h, the plate was maintained at 37°C without shaking for 1 to 4 h until the next daily dilution.

For the first 730 generations, at the end of each daily cycle the endpoint cultures were supplemented with 100 μl glycerol (50%, vol/vol) and stored at −80°C. In the event that the cultures needed to be restarted from the freezer, aliquots of the most recent frozen cultures were diluted 1:50 into sterile growth medium on the first day and were then diluted as normal on the following days. After 730 generations (61 daily dilutions) of continuous growth in moderate acid, the pH of the LBKmal growth medium was lowered to 4.6. The daily dilution was decreased to 1:100, to decrease the population bottleneck (23). The glycerol solution for storing cultures was buffered with 100 mM MES, and the pH was adjusted to 6.5, to decrease the stress of the storage condition. All other aspects of the protocol remained unchanged.

After a cumulative 2,000 generations of evolution (61 dilutions at 1:4,000, 190 dilutions at 1:100), four populations were selected at random and streaked out on LBK agar. From each population, two clones were selected for genome sequencing (Table 1).

Endpoint growth measurement at pH 4.6. As one measure of evolving fitness, the endpoint growth level (culture density at stationary phase) was measured for cultures derived from each of the 24 experimental populations. The reference populations were chosen after approximately 25, 730, and 2,000 generations. Reference populations were thawed, diluted 1:50 in LBKmal at pH 4.8 (100 mM HOMOPIPES), and placed into rows of wells (volume, 200 μl) in the same microplate. The cultures were then incubated in a well plate at 37°C in a SpectraMax Plus384 MicroPlate reader (Molecular Devices) for 22 h. Every 15 min for the duration of the growth period, the well plate was shaken for 3 s, after which the OD450 of each population was recorded. After 22 h, each population was diluted 1:100 into pH 4.6 medium for growth. A total of three serial dilutions at pH 4.6 (1:100) and 22-h incubations were performed. After each growth period, the endpoint values of the OD450 were recorded and averaged.

Lysine decarboxylase assays. Isolated colonies were tested for lysine decarboxylase and other catabolic enzyme activities using an Enterol-Plurist indicator battery (Liofilchem). For large-scale screening, lysine decarboxylase was assayed by spotting an isolated colony into 200 μl Moeller decarboxylase broth, modified from the procedure of the company BD (BBL, 2011). The medium contained 5 g/liter L-lysine, 1 g/liter glucose, 3 g/liter yeast extract, and 15 mg/liter bromocresol purple. Samples were incubated at 37°C in the wells of a microplate; the plate was sealed for anaerobiosis. After 24 h, the plate was inspected visually. A purple color (alkalization) indicated positive lysine decarboxylase activity, whereas a yellow color indicated low or absent activity. Visual inspection was confirmed by analysis of the absorbance readings recorded for
each well at 340 nm and 590 nm using the SpectraMax Plus384 microplate reader (Molecular Devices).

**Whole-genome sequencing.** The genomes of the ancestral and evolved strains were sequenced. For each strain, the total DNA was extracted using a DNasey DNA extraction kit (Qiagen). The DNA purity was confirmed by measuring the ratios of the absorbance at 260 nm/280 nm and 260 nm/230 nm. The genomes were sequenced with an Illumina MiSeq sequencer at the Michigan State University Research Technology Support Facility. Sequence reads were mapped to the *E. coli* K-12 W3110 reference genome (GenBank accession number NC_007779.1) (35). The quality scores (Q) for the sequences ranged from 25 to 33, with the average Q value being 30.8; this value corresponds to an error rate of 1/1,000. Sequences with 30-fold coverage or greater were accepted for mutation calls. Mutations were predicted with the breseq computational pipeline (http://barricklab.org/breseq) (22).

**Competition assays.** Evolved clones were assayed for changes in fitness relative to that of the ancestral strain (36). Competitors were differentiated in coculture on the basis of lac marker states. Each evolved strain (lac-phenotype [lac-] phenotype) was cocultured with the lac-negative construct of the ancestral wild type (W3110 lacZ::kan) at pH 4.6. To ensure that the lac-negative phenotype did not confer a fitness advantage or disadvantage in the relevant environment, a competition assay between the lac-negative ancestor (W3110 lacZ::kan) and the original ancestor (W3110) was carried out simultaneously with each experiment. The two strains were cultured separately overnight at 37°C for 24 h with aeration in LBKmad at pH 4.8 (100 mM HOMOPIPES). The cultures were reprinted 1:100 into pH 4.8 media and incubated for an additional 16 to 18 h with aeration.

To commence competition, 125 μl of an overnight culture from a lac-negative strain and 125 μl of an overnight culture from a lac+ strain were combined and diluted (100-fold) in 25 ml of LBKmad at pH 4.6 (100 mM HOMOPIPES). Samples were immediately serially diluted in M63 and incubated for 24 h of growth, which was used to ensure the cultures were not confluent. The OD600 was measured after 1 h and then subsequently at approximately 20-min intervals until the cultures began to enter stationary phase. The growth rate of each culture was calculated with respect to the time period corresponding to log-phase growth (37).

**Cotransductant strains and competition assays at pH 4.6.** A thiG761::kan insertion (80 to 90% linked to rpoC) was introduced into an evolved clone containing an rpoC mutation (clone B11-1) by P1 phage transduction using strain JW5549-1 (Keio collection). Cotransduction with the linked kan marker replaced the mutated region of the gene with the ancestral sequence. The replacement was confirmed by PCR amplification and sequencing of the gene. The forward primer sequence used was 5’-CACCCGAGATGATTTCGTG-3’, and the reverse primer sequence used was 5’-GAGCCTGTCAACCGGTTTAC-3’ (844 bp between primers). The cotransductant strain was competed against the parental clone in LBKmad at pH 4.6 following the same protocol described above.

**Nucleotide sequence accession number.** Sequence data have been deposited in the NCBI Sequence Read Archive (SRA) under accession number SRP041420.

## RESULTS

### Adaptation of 24 experimental *E. coli* populations at pH 4.6

We established culture conditions for maintaining experimental *E. coli* lineages below pH 5. In LBK medium, *E. coli* catabolizes diverse metabolites to optimize growth at low pH (9). The added components, HOMOPIPES and d/l-malate (LBKmad), buffered the acidity below pH 5. Both d- and l-malate may be catabolized by reactions that consume acid (38, 39) and thus potentially supply the experimental lineages with a novel means of enhancing acid tolerance. Using this medium, LBKmad, we established that our laboratory strain of *E. coli* K-12 W3110 could grow consistently in a microplate with the medium adjusted to pH 4.8.

Twenty-four bacterial populations were evolved under a daily dilution regime in 96-well microtiter plates in LBK medium supplemented with HOMOPIPES and d/l-malate, buffered initially at pH 4.8, as described in Materials and Methods. After 730 generations of growth, trial experiments of three successive 1:100 dilutions revealed that cultures of all experimental populations could be maintained at pH 4.6. We therefore decreased the pH of the culture conditions to pH 4.6 to provide even greater selective pressure on the experimental populations. We also decreased the daily dilution factor to 1:100, in order to decrease the daily population bottleneck and thus sample a greater number of possible mutations.

Over the course of the experiment, we observed significant increases in the endpoint culture density of all evolving populations, consistent with adaptation to low pH (Fig. 1). We measured the endpoint culture density of experimental populations sampled from populations frozen at several different generation numbers (generations 25, 730, and 2,000). The 25-generation plate (the second 4,000-fold dilution plate after the ancestor) was chosen instead of the ancestral strain, in order to test cultures frozen in plates under equivalent conditions. By generation 730, the acid-evolved populations showed an increased ability to grow at pH 4.6, although their endpoint densities declined by the third daily dilution. Comparison of the results by the t test to determine statistically significant differences (*P* < 0.001) verified the increase in endpoint growth between generations 25 and 730 and between generations 730 and 2,000 (evolved at pH 4.6). By generation 2,000, all populations reached endpoint densities higher than
enzyme activity (Table 2). For each generation number (760, 2,000), screening three generations of our 24 evolving populations for min. The final OD450 was measured after 22 h. Error bars represent SEMs for three subsequent days by daily dilution 100-fold at pH 4.6. On each day, cultures were conducted in a microplate reader for 22 h. Culture was conducted for three subsequent days by daily dilution 100-fold at pH 4.6. On each day, cultures were incubated at 37°C in a microplate reader with shaking every 15 min. The final OD_{450} was measured after 22 h. Error bars represent SEMs (n = 24).

FIG 1 Growth of evolved populations at pH 4.6. The endpoint growth of the 24 populations from generations 25, 730, and 2,000 was compared. Frozen populations were thawed and diluted 1:50 in LBKmal, pH 4.8, for initial recovery and then incubated in a microplate reader for 22 h. Culture was conducted for three subsequent days by daily dilution 100-fold at pH 4.6. On each day, cultures were incubated at 37°C in a microplate reader with shaking every 15 min. The final OD_{450} was measured after 22 h. Error bars represent SEMs (n = 24).

Those at generations 25 and 730. Furthermore, the 2,000-genera-
tion populations grew consistently over three serial dilutions at pH 4.6; a t test showed no difference among the endpoint reads for the three successive dilutions (P = 0.54). Similar experiments conducted at pH 7.0 and at pH 9.0 showed no fitness advantage or disadvantage (data not shown). Thus, the growth advantage of the evolved populations was specific to pH 4.6, and the evolved pop-
ulations incurred no fitness disadvantage at high pH.

Downregulation of lysine decarboxylase activity. E. coli bac-
teria respond to acid stress through various shifts in metabo-
lism, such as upregulation of the lysine and arginine decar-
boxylases (13, 40). We therefore screened isolates from our
evolving populations for several metabolic enzyme activities, using the Enterol-Pluri-Test. Several of our 2,000-genera-
tion clones tested negative for the activity of the degradative lysine
decarboxylase (CadA). This was surprising, given that in the
ancestor, the gene encoding CadA is one of the genes that is the
most highly induced by acid (13, 41).

We investigated the loss of lysine decarboxylase activity by
screening three generations of our 24 evolving populations for
enzyme activity (Table 2). For each generation number (760, 1,400, 2,000), three isolates from each of the 24 populations were
screened for lysine decarboxylase activity after 24 h growth in
Moeller broth with lysine. In this assay, the ancestor W3110 con-
sistently tested positive (purple), and isolates with a cadA deletion consistently tested negative (yellow). Among the sampled pop-
ulations, we observed a progressive decline in the prevalence of
decarboxylase-positive clones. At generation 760, only one isolate
(one of three isolates from population B7) tested negative. At gen-
eration 1,400, 26 isolates from 15 populations tested negative for
lysine decarboxylase function, and by generation 2,000, all but one
isolate (an isolate from population A11) tested negative. Further
screening of the populations revealed a minority of decarboxy-
lase-positive isolates remaining in several populations, such as
population F9 (data not shown).

Upon further incubation at 37°C, all sampled clones subse-
quently tested positive after 2 or 3 days. This observation suggests
that the enzyme was present but downregulated. Indeed, genome
sequencing subsequently confirmed that the cadA gene and its
own regulators showed no mutations in the downregulated
strains (see “Genomic analysis of acid-evolved isolates” below).
The basis of cadA downregulation remains to be elucidated, but
the occurrence of cadA downregulation in all 24 populations sug-
gests a beneficial fitness effect under our acid stress conditions.

Genomic analysis of acid-evolved isolates. To identify the
genic basis of the improved fitness of the evolved clones at low pH,
we sequenced the genomes of eight clones from generation 2,000
(SRA accession number SRP041420). Two clones were isolated
from each of four experimental populations, populations H9,
B11, F11, and F9. Among the sequenced clones, two (clones F9-2
and F9-3 from population F9) displayed lysine decarboxylase ac-
tivity comparable to that of the ancestor; the other six evolved
cloned tested negative.

For comparison with our acid-evolved clones, we resequenced
the ancestral strain W3110 from our original reference freeze-
er stock (D8-W), as well as a clone from a second freezer stock in our
collection (D13-W) (see Table S1 in the supplemental material).
Our two W3110 clones showed nearly identical sequences. How-
ever, several differences appeared between the genome of our lab
strain and the sequenced E. coli K-12 W3110 reference genome
ASM1024v1). For example, our lab strain showed deletions of the
insD-insC and insH-alsK regions. Also, a number of single nucleo-
tide polymorphisms (SNPs) appeared in the icd coding sequence.
The source of the unusual icd variability is unknown.

Mutations were identified in the sequenced genomes by align-
ing each isolate’s raw reads with the E. coli K-12 W3110 reference
genome (35) using the breseq computational pipeline (http://
barricklab.org/breseq) (24). Mutation calls were accepted for
sites showing 30-fold or greater coverage. Each acid-adapted
strain had between two and six nonsynonymous mutations.
Among the evolved genomes, we identified 24 mutations in cod-
ing regions (Table 3), which included 20 missense mutations, 2
single-base insertions (hepA, mppA), 1 9-bp insertion (yejM), and

TABLE 2 Lysine decarboxylase activity of clones from acid-evolved populations

<table>
<thead>
<tr>
<th>No. of isolates in the following population testing negative for lysine decarboxylase activity:</th>
</tr>
</thead>
<tbody>
<tr>
<td>No. of generations</td>
</tr>
<tr>
<td>760</td>
</tr>
<tr>
<td>1,400</td>
</tr>
<tr>
<td>2,000</td>
</tr>
</tbody>
</table>

*For each generation, three isolates from each population were assayed. The number of isolates of three isolates tested that lost lysine decarboxylase activity (indicated by a yellow color in Moeller broth) after 24 h of culture in a microplate is indicated.*
1 silent mutation (yjilK). The high ratio of nonsynonymous mutations to synonymous mutations across the evolved isolates suggests that these mutations were likely to be adaptive under the experimental conditions (42). An additional 12 mutations in intergenic regions were also identified (Table 4). Pairs of clones isolated from the same population typically displayed both shared and unshared mutations; the shared mutations presumably arose earlier in the ancestor common to that population. Only one mutation (yesJ insertion) that was clearly absent from the ancestor occurred in two acid-evolved populations. Multiple mutations were found in icd, but numerous SNPs in icd were found in the ancestral stock, suggesting a mutation frequency unrelated to the evolution experiment. The diversity of our evolved clones suggests that our remaining populations, of the original 24, contained further mutations of interest. Interestingly, only one of the sequenced mutations occurred in a gene with a well-known role in acid stress tolerance, adiY, the activator of the acid-inducible arginine decarboxylase (AdiA). The adiY mutation appeared in the two isolates of population B11 (B11-1 and B11-2). The other mutations that we found are likely

<table>
<thead>
<tr>
<th>Gene</th>
<th>Function</th>
<th>Mutation</th>
<th>Identification of a mutation in clone:</th>
</tr>
</thead>
<tbody>
<tr>
<td>flu</td>
<td>AG43 transport (biofilm)</td>
<td>Missense (A162E)</td>
<td>X        X</td>
</tr>
<tr>
<td>ftsX</td>
<td>Cell division (IM transporter)</td>
<td>Missense (A101V)</td>
<td>X        X</td>
</tr>
<tr>
<td>fucL</td>
<td>3-Fucose isomerase</td>
<td>Missense (R363S)</td>
<td>X        X</td>
</tr>
<tr>
<td>gcvP</td>
<td>Glycine decarboxylase</td>
<td>Missense (R854A)</td>
<td>X</td>
</tr>
<tr>
<td>hepA</td>
<td>RNA polymer recycling factor</td>
<td>Insertion (1139, +G)</td>
<td>X        X</td>
</tr>
<tr>
<td>icd</td>
<td>Isocitrate dehydrogenase</td>
<td>Missense (D398E)</td>
<td>X        X        X        X</td>
</tr>
<tr>
<td>menH</td>
<td>Menaquione biosynthesis</td>
<td>Missense (D79Y)</td>
<td>X        X</td>
</tr>
<tr>
<td>metE</td>
<td>Homocysteine transmethylyse</td>
<td>Missense (R337S)</td>
<td>X</td>
</tr>
<tr>
<td>mhpB</td>
<td>Phenylpropionate degradation</td>
<td>Missense (R226Y)</td>
<td>X</td>
</tr>
<tr>
<td>msbA</td>
<td>Lipid transport IM to OM</td>
<td>Missense (G150C)</td>
<td>X</td>
</tr>
<tr>
<td>rbmE</td>
<td>23S rRNA methyltransferase</td>
<td>Missense (P157I)</td>
<td>X</td>
</tr>
<tr>
<td>rpoB</td>
<td>RNA polymer beta subunit</td>
<td>Missense (A679V)</td>
<td>X</td>
</tr>
<tr>
<td>rpoC</td>
<td>RNA polymer beta prime subunit</td>
<td>Missense (I774S)</td>
<td>X</td>
</tr>
<tr>
<td>rpoD</td>
<td>RNA polymer sigma 70</td>
<td>Missense (M273I)</td>
<td>X</td>
</tr>
<tr>
<td>yjiM</td>
<td>IM protein Silent (TCG)</td>
<td>Missense (Q73K)</td>
<td>X</td>
</tr>
<tr>
<td>yjiK</td>
<td>IM protein Silent (TCG)</td>
<td>Missense (P314Q)</td>
<td>X        X</td>
</tr>
</tbody>
</table>

**TABLE 3** Mutations identified in coding regions in the 2,000th generation clones<sup>a</sup>

<table>
<thead>
<tr>
<th>Gene</th>
<th>Function</th>
<th>Mutation</th>
<th>Identification of a mutation in clone:</th>
</tr>
</thead>
<tbody>
<tr>
<td>TABLE 4</td>
<td>Intergenic mutations identified in the 2,000th generation acid-adapted clones&lt;sup&gt;a&lt;/sup&gt;</td>
<td></td>
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</tr>
</tbody>
</table>

**TABLE 4** Intergenic mutations identified in the 2,000th generation acid-adapted clones<sup>a</sup>

<table>
<thead>
<tr>
<th>Proximal gene(s)</th>
<th>Mutation (positions, nucleotide change)</th>
<th>Identification of a mutation in clone:</th>
</tr>
</thead>
<tbody>
<tr>
<td>matP ←→ ompA</td>
<td>Intergenic (+40/−36, C to A)</td>
<td>X</td>
</tr>
<tr>
<td>racR ←→ ydaS</td>
<td>Intergenic (−61/−63, C to A)</td>
<td>X</td>
</tr>
<tr>
<td>Y75_p4289 ←→ eaeH</td>
<td>Intergenic (−545/−1035, C to A)</td>
<td>X</td>
</tr>
<tr>
<td>proL ←→</td>
<td>Noncoding (57/77 nt, C to A)</td>
<td>X</td>
</tr>
<tr>
<td>yobT ←→ bhaA</td>
<td>Intergenic (−87/−544, C to A)</td>
<td>X</td>
</tr>
<tr>
<td>yflM ←→ yfM</td>
<td>Intergenic (−258/+102) +8 bp (GCACATG)</td>
<td>X</td>
</tr>
<tr>
<td>rhaA ←→ rhaD</td>
<td>Intergenic (+308/−143, C to A)</td>
<td>X</td>
</tr>
<tr>
<td>rrsC ←→</td>
<td>Noncoding (1417/1542 nt, +A)</td>
<td>X</td>
</tr>
<tr>
<td>pabA ←→ argD</td>
<td>Intergenic (+34/−52, A to G)</td>
<td>X</td>
</tr>
<tr>
<td>yfiI ←→ ubiC</td>
<td>Intergenic (+84/−139, G to T)</td>
<td>X</td>
</tr>
<tr>
<td>dcaU ←→ aspA</td>
<td>Intergenic (−202/+104, G to T)</td>
<td>X</td>
</tr>
<tr>
<td>aces ←→ mraA</td>
<td>Intergenic (−228/+165, G to A)</td>
<td>X</td>
</tr>
</tbody>
</table>

<sup>a</sup>The genomes of the experimental clones and the ancestral strain were sequenced and aligned with the sequence of the E. coli K-12 W3110 reference genome (35) using the breseq computational pipeline ([http://barricklab.org/breseq](http://barricklab.org/breseq)). The mutations listed represent differences between the genome of the ancestor and the adapted clones, excluding intergenic mutations. An X signifies that the mutation is present in that clone. IM, inner membrane; OM, outer membrane; ECA, enterobacterial common antigen.
to yield previously unrecognized mechanisms of acid stress response.

Every evolved genome had one mutation in a subunit of the RNA polymerase (RNAP) holoenzyme (43). The specific RNAP component that was affected differed among the populations. Population H9 isolates shared a mutation in \( rpoB \) (beta subunit; A679V), whereas B11 and F9 isolates each showed mutations in \( rpoC \) (beta prime subunit; V507L and I774S, respectively.) The \( rpoC \) beta prime subunit I774S mutation maps near the beta/beta prime interface and confers resistance to the CBR703 series of RNAP inhibitors (44). The F9 isolates shared a mutation in \( rpoD \) (housekeeping sigma 70; M273I). In each population, the RNAP mutation was shared by both clones, a result that would be consistent with early fixation in the evolution experiment. Besides the RNAP subunit mutations, the B11 strains also had a frameshift mutation in the HepA (RapA) helicase, which is involved in recycling RNAP and in UV resistance (45, 46).

**Relative fitness of 2,000th generation isolates at pH 4.6.** Fitness competitions were performed on our eight sequenced clones, in order to measure their relative fitness gains under acidic conditions. We cocultured the evolved clones against the ancestor over a 24-h growth period in \( \text{LBK}_{\text{mal}} \) buffered to pH 4.6. Competitor colonies were counted before and after growth by plating on X-Gal agar, on which the different \( lac \) marker states could be discerned. The relative fitness of each evolved isolate was measured by determining the number of generations that each strain accumulated during the 24-h period and then dividing the isolate’s number of doublings by that of the ancestor. Thus, a value of \( W \) of greater than 1 indicates that the \( lac^+ \) isolate had a fitness advantage, whereas a value of \( W \) of less than 1 indicates that the \( lac^- \) negative ancestor had a fitness advantage. For each isolate, equal numbers of competitions in which the \( lac \) phenotypes were reversed (\( lac^- \) isolate versus \( lac^+ \) negative ancestor) were performed. In order to confirm the neutrality of the \( lac^- \) negative marker, the \( lac^- \) negative ancestor was also competed with the \( lac^+ \) ancestor. The mean relative fitness of W3110 versus W3110 \( lac^+ :\text{kan} \) was not significantly different (one-sample t test) from 1.0 and was a mean of 1.045 ± 0.07 with 99% confidence.

All eight evolved clones outcompeted the ancestor and displayed relative fitness values of 1.5 or higher (Fig. 2A). The magnitude of the fitness advantage varied among the clones, with a somewhat lower fitness advantage being noted in the population F11 isolates, but all isolates outcompeted the ancestor. These fitness values are notably larger than those typically reported for fitness changes, in addition to displaying variation among replicates greater than that observed at low pH. This result shows that the improved fitness of our acid-evolved clones is specific to growth at low pH, with little to no correlated fitness change occurring at high pH.

**Fitness at pH 4.6**. Fitness improvement in one environment often comes at the cost of reduced fitness in another (23). We sought to determine if the improved fitness of the evolved clones at low pH carried the trade-off of reduced fitness at high pH. To test this possibility, we measured the fitness of four of the evolved clones relative to that of the ancestor in a medium buffered to pH 9 (Fig. 2C). The evolved clones showed insignificant to slightly negative fitness changes, in addition to displaying variation among replicates greater than that observed at low pH. This result shows that the improved fitness of our acid-evolved clones is specific to growth at low pH, with little to no correlated fitness change occurring at high pH.

**Early-log-phase growth of 2,000th generation isolates at pH 4.6.** The relative fitness of bacterial strains evolved in our microplates could be due to various factors, including the culture density reached by stationary phase (Fig. 1) as well as the rate of growth in log phase. To separate these factors, we measured the growth rates of our acid-evolved isolates under conditions conducive to optimal log-phase growth (rotary shaking of baffled flasks). We cultured four isolates (representing the four populations) in \( \text{LBK}_{\text{mal}} \) buffered to pH 4.5 and compared their early-log-phase growth rate to that of their ancestor (Fig. 3). The four isolates showed a significant increase in the log-phase growth rate compared with their ancestors, albeit with little to no correlated fitness change occurring at low pH.
The cotransductant strain was competed against its parental ancestral sequence. The sequence reversion was confirmed by the linked W3110 thiG761 mutation in the evolved strain in which the mutated rpoC gene (Table 3). In order to investigate the effect of the rpoC mutation on fitness, we transduced the linked W3110 thiG761::kan allele into strain B11-1 to produce a cotransductant strain in which the mutated rpoC is reverted to the ancestral sequence. The sequence reversion was confirmed by PCR. The cotransductant strain was competed against its parental mutant strain in LBK<sub>mal</sub> pH 4.6. The mean fitness of B11-1 lacZ::kan (containing rpoC V507L) relative to that of B11-1 thiG761::kan (containing a reverted rpoC) was 1.358 ± 0.03 (99% confidence, n = 3, one-sample t test, P = 0.008). The decreased fitness of B11-1 thiG761::kan with reverted rpoC suggests that the rpoC mutation provides a competitive advantage for acid growth.

**DISCUSSION**

In this study, we examined the adaptation of *E. coli* to growth below pH 5, a condition relevant to understanding both enteric bacterial passage into the intestine (1, 18) and fruit juice contamination (20). Following 2,000 generations of experimental evolution at the lower limit of *E. coli*’s tolerable pH range, we found that eight evolved clones isolated from 4 of our 24 populations displayed a substantially higher fitness than the ancestral clone under growth conditions of pH 4.6 (Fig. 2). Indeed, we observed a fitness improvement much greater than that which has been seen in previous long-term evolution experiments at pH 5.3 (31). Our experiment was performed in a complex medium, LBK<sub>mal</sub> which broadened the range of possible metabolic responses and, hence, targets of selection to the acidic conditions (9, 27). Evolved populations showed both a significantly higher endpoint culture density (Fig. 1) and significantly higher early-log-phase growth rates (Fig. 3), suggesting that these conditions selected for improvements in both the early and late parts of the growth cycle.

**Mutations in the RNA polymerase holoenzyme of acid-adapted strains.** All acid-adapted evolved clones that we examined have a missense mutation in a component of the RNA polymerase holoenzyme (43, 47, 48). Our fitness competition assays suggest that the altered rpoC allele of evolved clone B11-1 confers a competitive advantage for acidic growth. In other reports, RNA polymerase mutations arise in some but not all laboratory evolution experiments; the mechanism by which they alter fitness is poorly understood (23, 30).

For our acid-evolved strains, we proposed three possible mechanisms by which the observed RNA polymerase mutations might have enhanced fitness at low pH. (i) The RNA polymerase mutations could increase the stability of the complex at low pH, when the cytoplasm is slightly acidified by proton leakage from the external medium. In this case, we would expect the acid-evolved strains to show increased fitness in the presence of benzoic acid, which acidifies the cytoplasm (49). However, our acid-evolved clones showed decreased fitness in benzoate (Fig. 2B). Thus, under our conditions, acid-stressed bacteria did not evolve resistance to permeant acids.

(ii) The RNA polymerase mutations could destabilize the complex and thus moderate the overexpression of acid stress genes. Such overexpression at the edge of the permissive pH range could be deleterious for growth, for example, by catabolizing amino acids needed for protein synthesis. Destabilizing mutations of RNA polymerase have been described, although the details of their mechanism remain poorly understood (50). Downregulation via RNA polymerase destabilization could be consistent with our observed downregulation of lysine decarboxylase in all evolving populations (Table 2). The gene *cadA* is one of the most highly expressed acid stress genes in *E. coli* (13, 17, 51).

(iii) The RNA polymerase mutations could have caused the rapid adjustment of multiple regulons in small ways that cumulatively increase fitness under the selected condition. For example, under an experimental evolution involving adaptation to minimal glycerol medium, Herrig et al. found that early RNA polymerase substitutions conferred the greatest fitness advantage of all mutations isolated (32). Moreover, many other SNPs confer a fitness advantage only when combined with RNA polymerase. The authors hypothesized that the RNA polymerase mutations fixed very early in the experiment due to fitness benefits stemming from their manifold effects on gene regulation. They further hypothesized that subsequent mutations might have functioned to compensate for the inevitable negative side effects of altered RNA polymerase function (47, 53).

**Putative acid stress response genes.** In our analysis of acid-evolved genome sequences, we identified mutations in several genes known to be important for *E. coli*’s acid stress response. The most notable example of these was in adiY (mutated in the B11 population of isolates). The adiY gene encodes an AraC-like regulatory protein that can induce expression of AdiA (arginine decarboxylase) and AdiC (arginine-agmatine antiporter), the essential components of the arginine-dependent acid resistance system (6, 40). The effect of the adiY mutation could be mediated by either an increased or a decreased rate of adiAC transcription.

Two other strains, F9-2 and F9-3, had a mutation in a regulatory upstream of the *nuo* operon, which codes for NADH:ubiquinone oxidoreductase (respiratory complex I), a component of the electron transport chain (47, 48). The authors hypothesized that the RNA polymerase mutations fixed very early in the experiment due to fitness benefits stemming from their manifold effects on gene regulation. They further hypothesized that subsequent mutations might have functioned to compensate for the inevitable negative side effects of altered RNA polymerase function (47, 53).
electron transport chain (54, 55). The respiratory complex I is a proton pump of the electron transport chain known to be upregulated at low external pH (17). If this mutation upstream of nuoA were to increase the rate of transcription of the nuo operon, it could potentially increase acid stress by increasing the bacterium’s net proton efflux.

In one clone, H9-2, we identified a missense mutation in gcvP, which encodes glycine decarboxylase P, part of the glycine cleavage system (56). Transcription of gcvP is positively regulated by glycine (57) and is responsible for the reversible breakdown of glycine into carbon dioxide, ammonia, and a cofactor-bound methylene group (58). The glutamate, arginine, lysine, and ornithine decarboxylase reactions have been demonstrated to play a role in maintaining neutral cytoplasmic pH. It is possible that the catabolic glycine decarboxylase pathway similarly serves to buffer the cytoplasm via the release of ammonia during glycine decarboxylation. Indeed, Lu et al. recently demonstrated that the conversion of glutamine to glutamate, with the release of ammonia, can serve a protective role for E. coli during exposure at pH 2.5 (59).

The protein Ag43, encoded by flu, was mutated in clones isolated from population H9. Ag43 is an autotransporter protein involved in cellular aggregation and biofilm formation, and production of Ag43 in a planktonic culture is associated with bacterial autoaggregation (forming clumps) (60, 61). Expression of flu is subject to phase variation, in which individuals in the same clonal population vary qualitatively in their production of the protein (60, 61). The missense mutation (A162E) that we identified is located in the protein’s passenger domain, α52, which protrudes from the outer membrane into the extracellular space (60). This suggests that the mutant strains might have an altered ability to adhere to other cells, perhaps by binding to each other more tightly within the culture. Aggregation could benefit acid-stressed cells by reducing the total area of the cell surface exposed to the acidic medium, thereby reducing the rate of proton influx from the environment across the membrane.

Another possible acid stress gene is msbA, which encodes an ATP-dependent inner membrane flipase responsible for lipid transport from the inner leaflet to the outer leaflet of the bacterium’s inner membrane. Loss of function of MsbA causes phospholipids and lipopolysaccharides (LPSs) to collect at the cell’s inner membrane, thereby compromising the cell’s structural integrity (62). The two evolved clones from population F11 showed the presence of a G150C missense mutation in one of MsbA’s transmembrane domains. It is possible that this mutation could alter membrane integrity, which could in turn impact acid sensitivity.

This study has established eight acid-adapted E. coli clones for future study, each of which shows significant, though varying, fitness improvements relative to their ancestor. Many exciting genetic and evolutionary questions about these strains remain to be addressed. In future studies, we intend to investigate the role of RNAS mutations in gene expression and relative fitness, as well as the putative contributions of the other potentially beneficial mutant alleles that we have identified.

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