

ERRATA

Cloning of the *Alcaligenes latus* Polyhydroxyalkanoate Biosynthesis Genes and Use of These Genes for Enhanced Production of Poly(3-hydroxybutyrate) in *Escherichia coli*

JONG-IL CHOI, SANG YUP LEE, AND KYUBOEM HAN

Department of Chemical Engineering and BioProcess Engineering Research Center, Korea Advanced Institute of Science and Technology, 373-1 Kusong-dong, Yusong-gu, Taejeon 305-701, and Biotech Research Institute II, LG Chemicals, Ltd., Science Town, Taejeon 305-380, Korea

Volume 64, no. 12, p. 4897–4903, 1998. Page 4898, column 2, Results, line 16: “6.3-kb” should read “6.4-kb.”

Line 19: “6,286 bp” should read “6,433 bp.”

Page 4899, Fig. 1. Figure 1 should appear as shown below.

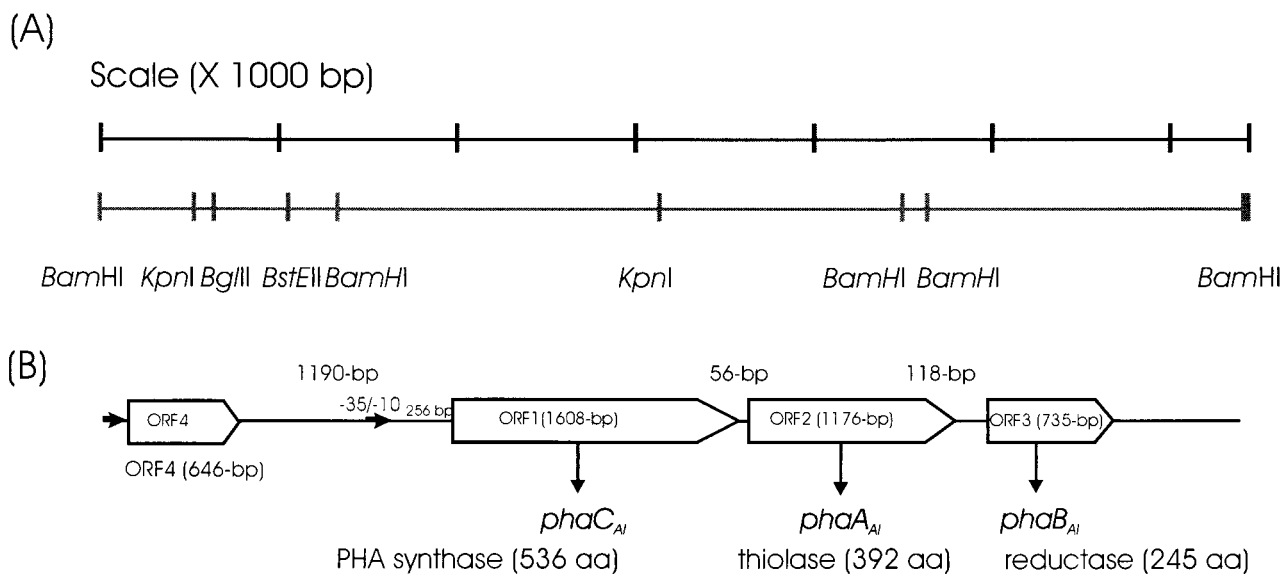


FIG. 1.

Column 1, line 4: “1,029 bp” should read “1,176 bp.”

Line 5: “343 amino acids” should read “392 amino acids.”

Line 6: “35,406 Da” should read “40,519 Da.”

Page 4900, column 1, line 21: “6.3-kb” should read “6.4-kb.”

Line 31: “5.3-kb” should read “5.4-kb.”

Column 2, Discussion, line 25: “6.3-kb” should read “6.4-kb.”

Page 4901, Fig. 2: “pJC1 (9-kbp),” “pJC2 (9.7-kbp),” “pJC3 (8-kbp),” and “pJC4 (8.7-kbp)” should read “pJC1 (9.1-kbp),” “pJC2 (9.8-kbp),” “pJC3 (8.1-kbp),” and “pJC4 (8.8-kbp),” respectively.

Inactivation of *Cryptosporidium parvum* Oocysts by Ammonia

MICHAEL B. JENKINS, DWIGHT D. BOWMAN, AND WILLIAM C. GHIORSE

Section of Microbiology, Division of Biological Sciences, and Department of Microbiology and Immunology,
College of Veterinary Medicine, Cornell University, Ithaca, New York 14853

Volume 64, no. 2, p. 784–788, 1998. p. 786, Table 1. Table 1 should read as shown below.

TABLE 1. Inactivation rates of *C. parvum* oocysts exposed to measured concentrations of ammonia^a

[NH ₃] (mol/liter)	$K \pm 95\% \text{ CI/h}^b$	Days to reach 99.999% inactivation ^c
0.007	0.014 \pm 0.004	34.3
0.026	0.027 \pm 0.007	17.8
0.039	0.050 \pm 0.005	9.6
0.060 ^d	0.047 \pm 0.014	10.2
0.104	0.059 \pm 0.034	8.1
0.148	0.066 \pm 0.030	7.3
5.8 ^e	0.479	1

^aBased on data from the dye permeability assay after a 24-h exposure.

^bIt was assumed that oocyst inactivation was a first-order process. The coefficient of inactivation was determined by regressing $\ln(P_0/P_t)$ against time (derived from the equation $P_t = P_0 \cdot e^{-Kt}$, where P_0 is the initial percentage of viable oocysts, P_t is the percentage of viable oocysts at time t , in hours, and K is the coefficient of inactivation). The 95% confidence intervals (CI) were determined by multiplying the Student t value at the appropriate degree of freedom and at an α level (two-sided) of 0.025 by the standard deviations of K .

^cCalculated by the equation $t = \ln(P_0/P_t)/K$.

^dThis concentration of NH₃ and exposure time were used in the validation experiment shown in Table 2.

^eA power function, $y = 2.523x^{-0.525}$ ($r^2 = 0.993$), that fit the regression of [NH₃] against days to reach 99.999% inactivation was used to determine the concentration of ammonia that would reduce the viability of oocysts by 99.999% in 1 day. The K value for this concentration of ammonia was then derived.

Page 787, column 2, line 9: “26.5 days” should read “55.1 days.”