

Chlorine Inactivation of Bacterial Bioterrorism Agents

Laura J. Rose,^{1*} Eugene W. Rice,² Bette Jensen,¹ Ricardo Murga,^{1†}
Alicia Peterson,¹ Rodney M. Donlan,¹ and Matthew J. Arduino¹

Centers for Disease Control and Prevention, Atlanta, Georgia,¹ and U.S. Environmental
Protection Agency, Cincinnati, Ohio²

Received 20 April 2004/Accepted 29 August 2004

Seven species of bacterial select agents were tested for susceptibility to free available chlorine (FAC). Under test conditions, the FAC routinely maintained in potable water would be sufficient to reduce six species by 2 orders of magnitude within 10 min. Water contaminated with spores of *Bacillus anthracis* spores would require further treatment.

The contamination of the U.S. mail system with anthrax spores in 2001 has heightened concern about the safety of other public services and utilities, including our water distribution systems. Currently, chlorination is the most common method of disinfecting drinking water in the United States (2, 3). The U.S. Environmental Protection Agency and the water treatment industry use Ct values ("C" is the chlorine concentration in milligrams/liter, and "t" is the exposure time in minutes) to calculate microbial inactivation and to evaluate the effectiveness of a water treatment system. Ct tables have been developed for some waterborne pathogens to indicate conditions necessary for a 2-log₁₀ (99%) or 3-log₁₀ (99.9%) inactivation. The present study determined Ct values for 2-log and 3-log inactivation of seven species (11 isolates) on the Centers for Disease Control and Prevention (CDC) list of category A and B potential bioterrorism agents (12).

The bacterial strains *Bacillus anthracis* Ames, *Brucella melitensis* ATCC 23456, *Brucella suis* EAM562, *Burkholderia mallei* M-9, *Burkholderia mallei* M-13, *Burkholderia pseudomallei* ATCC 1688, *Francisella tularensis* LVS, *Francisella tularensis* NY98, *Yersinia pestis* A1122, and *Yersinia pestis* Harbin were obtained from CDC laboratories. *B. anthracis* Sterne 34F2 was obtained from Colorado Serum Co., Denver, Colo.

The effect of each chlorine concentration was tested in triplicate by using chlorine demand-free buffer (0.05 M KH₂PO₄; pH 7) and maintained at 5 and 25°C. Testing methods are described elsewhere (4). Free available chlorine (FAC) and total chlorine were monitored by using DPD colorimetric analysis (1).

Decay curves were generated for each organism by using the log₁₀-transformed data of the mean CFU counts at each time, temperature, and chlorine concentration. Linear regressions of the appropriate segments of the decay curves were performed to estimate the time needed for a 99 or 99.9% reduction in viable counts. The Ct values were calculated by multiplying inactivation times for a given temperature and percent inactivation by the chlorine concentration at that time. The reported

Ct values represent the mean of the Ct values calculated for each chlorine concentration.

The results of the chlorine challenge and the calculated Ct values are shown in Table 1 for the gram-negative bacteria and in Table 2 for *Bacillus anthracis* spores. *Burkholderia*, *Brucella*, and *Yersinia* strains were more susceptible to chlorine treatment than *Francisella tularensis*, as shown by Ct values ≤0.7 for a 3-log inactivation of these organisms. Ct values for a 3-log inactivation of *Francisella tularensis* ranged from 1.0 to 10.3. The slightly greater resistance of *Francisella tularensis* to chlorine was also observed by Foote et al. (6) as determined by injection into guinea pigs.

The *Bacillus anthracis* spores were less susceptible to chlorine disinfection than the gram-negative organisms. The Ames strain was slightly less susceptible to the chlorine than the Sterne strain, requiring more than 2 h for a 2-log reduction when exposed to 0.8 mg of FAC/liter at 25°C, whereas the Sterne strain underwent a >4-log reduction in counts after 2 h under similar conditions (Table 2). The Ct values determined in the present study for *B. anthracis* spores are comparable to the data obtained by Brazis et al. (5), from which Ct values can be calculated to be 458 at 4°C and 113 at 22°C for a 4-log reduction (99.99%). Differences between the Brazis findings and the results of the present study may be attributed to variability between strains, slight pH or temperature differences, or methods of spore preparation.

A 1992 survey of samples from 283 water utilities reported that of those that use chlorine, a median residual of ~1.1 mg/liter, and a median contact time to the first point of use (from treatment facility to first access point in the water distribution system) of 45 min was reported in the utilities responding to the survey (13). Using the survey median result as a guide, we can estimate a median Ct value of 49.5 (1.1 mg/liter × 45 min) for the 283 water utilities surveyed. Our study shows that viable *Burkholderia mallei*, *Burkholderia pseudomallei*, *Brucella melitensis*, *Brucella suis*, *Francisella tularensis*, and *Yersinia pestis* would be reduced by more than 3 orders of magnitude under these median conditions if pH and temperatures were similar to those in the present study. The *Bacillus anthracis* spores, however, would not be inactivated by 2 log₁₀ or 3 log₁₀ under these median treatment conditions.

The Ct value can provide some indication of the efficacy of

* Corresponding author. Mailing address: Centers for Disease Control and Prevention, 1600 Clifton Rd., C16, Atlanta, GA 30333. Phone: (404) 639-4984. Fax: (404) 639-3822. E-mail: lrose@cdc.gov.

† Present address: Fort Defiance Indian Hospital, Fort Defiance, AZ 86504.

TABLE 1. Free chlorine inactivation of select agents at pH 7

Isolate	Temp (°C)	Initial inoculum (log ₁₀ CFU)	FAC (mg/liter)	Viable cells after exposure time [mean log ₁₀ (CFU + 1)/ml, n = 3] ^a of:			Ct (mg min/liter)	
				1 min	5 min	10 min	2 log	3 log
<i>Burkholderia mallei</i> M-9	5	5.8	0.14	5.1	0.5	0.0	0.2	0.2
			0.39	0.0	0.0			
	25	6.0	0.15	0.8	0.0	0.0	0.1	0.2
			0.46	0.0	0.0	0.0		
<i>Burkholderia mallei</i> M-13	5	5.8	0.13	5.7	2.1	2.1	0.2	0.2
			0.47	0.0	0.0	0.0		
	25	5.7	0.21	0.0	0.0	0.0	0.1	0.2
			0.60	0.0	0.0	0.0		
			1.01	0.0	0.0	0.0		
<i>Burkholderia pseudomallei</i> ATCC 1688	5	5.5	0.21	3.2	1.6	0.0	0.5	0.7
			0.65	2.3	0.0	0.0		
	25	5.6	1.43	0.0	0.0	0.0	0.4	0.6
			0.08	4.4	2.6	0.9		
			0.43	1.5	0.0	0.0		
			0.85	0.0	0.0	0.0		
<i>Brucella melitensis</i> ATCC 23456	5	6.6	0.16	3.6	2.9	2.5	0.3	0.5
			0.50	3.0	2.0	0.5		
	25	6.9	1.10	2.4	0.4	0.0	0.1	0.2
			0.04	4.8	2.6	1.8		
			0.23	0.8	0.4	0.0		
			0.85	0.0	0.0	0.0		
<i>Brucella suis</i> EAM562	5	6.8	0.20	2.6	1.2	0.0	0.3	0.4
			0.50	2.3	0.0	0.0		
	25	7.2	1.00	0.0	0.0	0.0	0.1	0.2
			0.15	1.1	0.0	0.0		
			0.20	0.1	0.0	0.0		
			0.60	0.0	0.0	0.0		
<i>Francisella tularensis</i> NY98	5	6.5	0.20	6.4	6.2	6.1	7.8	10.3 ^b
			0.54	6.4	6.1	5.8		
	25	6.6	1.00	6.3	5.9	4.2	2.0	3.9
			0.30	6.1	6.2	5.6		
			0.72	5.5	4.1	0.0		
			1.66	3.7	1.9	0.0		
<i>Francisella tularensis</i> LVS	5	7.0	0.23	5.5	4.9	3.9	1.5	2.4
			0.50	5.2	3.6	1.9		
	25	6.8	1.06	4.8	2.7	1.3	0.6	1.0
			0.10	5.5	5.5	5.5		
			0.36	5.4	1.9	2.1		
			0.86	3.7	0.0	0.0		
<i>Yersinia pestis</i> A1122	5	6.1	0.42	0.0	0.0	0.0	0.5	0.7
			0.98	0.0	0.0	0.0		
	25	6.4	1.84	0.0	0.0	0.0	0.4	0.6
			0.37	1.0	0.0	0.0		
			0.76	1.2	0.0	0.0		
			1.60	0.0	0.0	0.0		
<i>Yersinia pestis</i> Harbin	5	6.6	0.06	1.8	0.7	0.1	0.03	0.04
			0.31	0.0	0.0	0.0		
	25	6.6	0.84	0.0	0.0	0.0	0.03	0.04
			0.08	1.6	0.0	0.0		
			0.31	0.0	0.0	0.0		
			0.82	0.0	0.0	0.0		

^a To account for zero values, we added 1 to all datum points before conversion to the log₁₀ scale.

^b Extrapolated value.

disinfectants on pathogenic organisms but must be determined for each species and at several temperatures and pH conditions. If the water temperature were lower or the pH were higher within a particular distribution system, the Ct values would be expected to be higher for all organisms than those

calculated in the present study. Other factors that may promote survival of bacteria in chlorinated water include nutrient availability before exposure to FAC (9–11), attachment to surfaces (8), clumping and incorporation in organic materials (7), and incorporation into biofilms present in drinking water sys-

TABLE 2. Free chlorine inactivation of *Bacillus anthracis* spores at pH 7

Isolate	Temp (°C)	Initial inoculum (log ₁₀ CFU/ml)	FAC (mg/liter)	Viable spores after exposure time [mean log ₁₀ (CFU + 1)/ml, n = 3] ^a				Ct (mg min/liter)	
				1 h	2 h	3 h	4 h	2 log	3 log
<i>Bacillus anthracis</i> Sterne	5	4.1	0.88	3.8	3.8	3.6	2.5	190	271
			1.98	3.6	2.6	0.2	0.0		
			2.94	2.7	0.0	0.0	0.0		
	25	4.3	1.02	3.6	0.0	0.0	0.0	60	86
			1.96	0.0	0.0	0.0	0.0		
<i>Bacillus anthracis</i> Ames	5	4.9	0.80	4.9	4.8	ND	3.9	220	339
			2.80	4.2	0.0	ND	0.0		
	25	4.9	0.80	4.6	2.6	ND	0.0	79	102
			2.80	0.0	0.0	ND	0.0		
			4.80	0.0	0.0	ND	0.0		

^a To account for zero values, we added 1 to all datum points before conversion to the log₁₀ scale.

tems (9). Continued work to address the efficacy of FAC under various water conditions, as well as evaluation of other water treatment methods such as monochloramine, ozone, and UV light, is essential for protecting public health in the event of an intentional release of these bacterial agents into a potable water system.

We thank May Chu, CDC, for providing the *Francisella tularensis* isolates; Tanja Popovic, CDC, for providing the *Burkholderia* and *Brucella* isolates; and Richard Meyer, CDC, for providing the *Yersinia pestis* isolates.

REFERENCES

- American Public Health Association. 1995. Standard methods for the examination of water and wastewater, 19th ed. American Public Health Association, Washington, D.C.
- American Water Works Association Water Quality Division Disinfection Systems Committee. 2000. Committee report: disinfection at small systems. J. Am. Water Works Assoc. **92**:24–31.
- American Water Works Association Water Quality Division Disinfection Systems Committee. 2000. Committee report: disinfection at large and medium-sized systems. J. Am. Water Works Assoc. **92**:32–43.
- Berman, D., and J. C. Hoff. 1984. Inactivation of simian rotavirus SA11 by chlorine, chlorine dioxide, and monochloramine. Appl. Microbiol. **48**:317–323.
- Brazis, A. R., J. E. Leslie, P. W. Kabler, and R. L. Woodward. 1958. The inactivation of spores of *Bacillus globigii* and *Bacillus anthracis* by free available chlorine. Appl. Microbiol. **6**:338–342.
- Foote, H. B., W. L. Jellison, E. A. Steinhaus, and G. M. Kohls. 1943. Effect of chlorination of *Pasteurella tularensis* in aqueous suspension. J. Am. Water Works Assoc. **35**:902–910.
- LeChevallier, M. W., T. M. Evans, and R. J. Seidler. 1981. Effect of turbidity on chlorination efficiency and bacterial persistence in drinking water. Appl. Environ. Microbiol. **42**:159–167.
- LeChevallier, M. W., T. S. Hasseneauer, A. K. Camper, and G. A. McFeters. 1984. Disinfection of bacteria attached to granular activated carbon. Appl. Environ. Microbiol. **48**:918–923.
- LeChevallier, M. W., C. Cawthon, and R. G. Lee. 1988. Inactivation of biofilm bacteria. Appl. Environ. Microbiol. **54**:2492–2499.
- LeDantec, C., J. P. Duget, A. Montiel, N. Dumoutier, S. Dubrou, and V. Vincent. 2002. Chlorine disinfection of atypical mycobacteria isolated from a water distribution system. Appl. Environ. Microbiol. **68**:1025–1032.
- Lisle, J. T., S. C. Broadaway, A. M. Prescott, B. H. Pyle, C. Fricker, and G. A. McFeters. 1998. Effects of starvation on physiological activity and chlorine disinfection resistance in *Escherichia coli* O157:H7. Appl. Environ. Microbiol. **64**:4658–4662.
- Rotz, L. D., A. S. Kahn, S. R. Lillibridge, S. M. Ostroff, and J. M. Hughes. 2002. Public health assessment of potential biological terrorism agents. Emerg. Infect. Dis. **8**:225–230.
- Water Quality Division Disinfection Committee. 1992. Survey of water utility disinfection practices. J. Am. Water Works Assoc. **84**:121–128.