

Genetic and Phylogeographic Structures of the Symbiotic Fungus *Tuber magnatum*†

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The quality and market price of truffles vary with the species and, traditionally, the place of origin. The premium species *Tuber magnatum* produces white truffles and has a patchy distribution restricted to Italy and some Balkan areas. We used polymorphic microsatellites to evaluate 316 specimens grouped into 26 populations sampled across the species' geographic range to determine if natural populations of *T. magnatum* are genetically differentiated. We found that the southernmost and the northwesternmost populations were significantly differentiated from the rest of the populations. The simple sequence repeat data also could be used to make inferences about the postglacial *T. magnatum* expansion pattern. This study is the first to identify a genetic and phylogeographic structure in *T. magnatum*. The presence of a genetic structure can be of practical interest in tracing truffle populations according to their geographic origin for marketing strategies. Evidence for extensive outcrossing in field populations of *T. magnatum* also is provided for the first time.

Tuber spp. are hypogeous ascomycetes that grow in ectomycorrhizal symbioses with some shrub and tree species and produce ascomata, known as truffles. Some truffles are edible and appreciated worldwide. Their quality and market price depend on the species and, traditionally, the place of origin. Natural truffle production has declined dramatically over the past century (12). This decline and the flourishing truffle market have encouraged large-scale programs for growing these fungi through the planting of nursery-produced mycorrhizal trees. These programs have been widely developed for some species, e.g., *Tuber melanosporum* Vittad. and *Tuber uncinatum* Chatin, and most of the commercial demand for these truffles is satisfied by these artificial plantations rather than natural field collections (12). How the deliberate introduction of foreign strains into native populations has affected truffle production and biodiversity remains unknown.

The complexity of the truffle life cycle, the difficulties of growing these fungi under controlled conditions, and the lack of reliable phenotypic markers to differentiate morphologically similar species have been major obstacles to understanding the distribution, propagation, and fructification of these hypogeous fungi. Molecular markers have been developed to type most of the economically important truffle species (2, 10, 15, 19, 23, 24, 25, 33, 34), but studies of the environmental and molecular determinants for different portions of the life cycle generally are lacking. The spatial distribution and ecological requirements of these symbiotic fungi vary by species. Some species are widely distributed and have pronounced morpho-

logical and molecular variability (9, 11, 22, 25, 27, 32), while others have a more restricted distribution and little intraspecific polymorphism in either morphological or genetic traits (3, 4, 8, 9, 26). *Tuber magnatum* Pico, which produces white ascomata, is harvested only in Italy and some countries on the Balkan Peninsula.

In the related species *T. melanosporum*, there is little or no genetic differentiation between the sampled populations, and the species is regarded as one that self-fertilizes for sexual reproduction and that has an ascocarp consisting primarily of diploid (dikaryotic) hyphae (3, 4, 21). The few studies of genetic structure and mode of reproduction of *T. magnatum* found very limited intraspecific allozyme variation and a complete absence of heterozygous individuals (8). Whether *T. magnatum* is normally dikaryotic and reproduces homothallically or by self-fertilization or if it is an outbreeding ascomycete remains to be determined.

Our objectives in this study were to use microsatellite markers to determine (i) if *T. magnatum* populations are genetically homogeneous, (ii) if the fungus is dikaryotic, and (iii) the degree of outbreeding in field populations. Understanding the life cycle and genetic differences between truffle populations is important for commercial truffle growth management and marketing strategies.

MATERIALS AND METHODS

Sample source. Ascomata of *T. magnatum* were sampled from natural truffle grounds in the best known and most productive areas of Italy and from the Istrian peninsula (Croatia and Slovenia) with the help of local pickers. When multiple samples were provided by the same individual, but evidence for collection from physically different locations was lacking, the samples were screened for clones with simple sequence repeats (SSR), and strains with identical multilocus genotypes were reduced to a single representative for the statistical analyses. Thus, only 316 out of 351 ascocarps sampled were used in the statistical analysis. These truffles were grouped into 26 populations and then these populations were clustered into 13 groups based on geographic criteria (Table 1). Ascocarps also were harvested from single-truffle grounds (Ascoli, Certaldo,

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TABLE 1. Name and location of the studied populations and estimated diversity parameters

Geographic group	Population no. and name	Region	No. of samples	Mean no. of alleles ^a	Allelic richness ^a
1	1. Istria	Croatia-Slovenia	17	3.3 (0.61)	2.7 (0.48)
2	2. Pavia	Lombardia	11	2.6 (0.43)	2.4 (0.44)
	3. Langhe	Piemonte	15	3.3 (0.81)	2.9 (0.62)
	4. Asti	Piemonte	9	2.7 (0.42)	2.6 (0.37)
	5. Alba	Piemonte	7	3.0 (0.49)	2.9 (0.47)
3	6. Pianoro	Emilia Romagna	9	2.7 (0.57)	2.6 (0.52)
	7. Val di Zena	Emilia Romagna	10	3.1 (0.59)	2.8 (0.48)
4	8. Firenze	Toscana	32	4.4 (0.97)	3.0 (0.46)
5	9. Pietralunga	Umbria	7	2.6 (0.43)	2.5 (0.43)
	10. Montemaggiore	Umbria	12	2.4 (0.57)	2.2 (0.53)
	11. Città di Castello	Umbria	12	3.1 (0.74)	2.7 (0.60)
	12. Gubbio	Umbria	25	4.3 (0.84)	2.9 (0.46)
	13. Appennino Umbro	Umbria	9	2.6 (0.53)	2.4 (0.50)
6	14. Fabro	Umbria	11	2.9 (0.46)	2.6 (0.39)
	15. Montecastrilli	Umbria	9	3.0 (0.62)	2.8 (0.57)
7	16. Ascoli	Marche	10	3.0 (0.58)	2.7 (0.55)
	17. Chieti	Abruzzo	5	2.4 (0.43)	2.4 (0.46)
8	18. Valle Roveto	Abruzzo	7	2.4 (0.43)	2.4 (0.44)
9	19. Quadri, Rosello	Abruzzo	12	3.1 (0.59)	2.8 (0.42)
	20. R. del Sangro, M. del. Sannio	Abruzzo-Molise	10	3.1 (0.46)	2.8 (0.36)
	21. Ateleta, S.P. Avellana	Abruzzo-Molise	18	3.9 (0.59)	3.0 (0.50)
	22. Castel del Giudice	Molise	16	3.1 (0.46)	2.7 (0.37)
10	23. Agnone	Molise	10	3.7 (0.75)	3.3 (0.65)
11	24. Isernia, Campobasso	Molise	16	3.6 (0.57)	3.1 (0.42)
12	25. Benevento, Avellino	Campania	5	2.4 (0.37)	2.4 (0.40)
13	26. Potenza	Basilicata	12	1.9 (0.34)	1.7 (0.26)

^a Standard errors of the means are shown in parentheses.

and Pietralunga) and analyzed to determine the amount of variation within a local population (see Table SA1 in the supplemental material).

DNA isolation and SSR characterization. Sample preparation and DNA isolation from all ascocarps (24) and amplification of the microsatellite loci (35) were carried out as previously described. The MA2 and MA5 loci are genetically unlinked, but each contain an (AC)_n-(TC)_n compound microsatellite motif. The total length of these compound loci has been previously determined following PCR with a locus-specific primer pair designed on the flanking regions of MA2 and MA5 (35). To reduce the bias due to size homoplasy, selective amplification of only one of the two motifs (MA2-1 and MA5-1) within each of the two compound loci was made with a new primer set, consisting of a nested primer (5'-AGAGAGAGAGTGTGTGTG-3') designed at the junction between the two repeated motifs, in combination with the locus-specific flanking primers 5'-CGCAGTGCAAAAAGGAAGCATCA-3' and 5'-CGTCTATTCCCCGCG AGATAACAAC-3' for MA2 and MA5, respectively. PCR conditions were as previously described (35). The resulting DNA fragments were resolved with an ABI 310 genetic analyzer (Applied Biosystems, Foster City, CA) to estimate allele size for information for statistical analyses. The 15 most common alleles of MA2-1 and MA5-1 and a representative of each of the alleles of the other microsatellite loci, MA4, MA7, MA12, MA14, and MA19 (35), were sequenced. Sequencing reactions were performed directly on purified PCR products (Jet Quick spin kit; Genomed, Bad Oeynhausen, Germany) with a Big Dye terminator sequencing kit (Applied Biosystems). Estimated allele sizes were used to determine the number of repeated units.

Data analysis. The mean number of alleles and the allelic richness, over all loci, were calculated for each population. Allelic richness, computed for each

population, was weighted to the smallest sample size ($n = 5$) using a rarefaction method (29).

Two-locus linkage disequilibrium analysis was performed with GENEPOP, version 3.2a (30), within each population and each geographic group and across all populations. The index of association (I_A) used to test for multilocus linkage disequilibrium was calculated as described by Maynard Smith et al. (18) with the program MULTILOCUS, version 1.2 (1). The same software was used to determine if I_A differed significantly from zero, by performing 1,000 randomizations of the data set. The randomization procedure was first performed by shuffling alleles among all individuals. To determine if linkage disequilibrium results from the reproductive isolation of geographically distant populations, randomizations by shuffling alleles among individuals within populations and within geographical groups also were performed (17).

To test the effects of sample size reduction on linkage disequilibrium (LD) values, two locus and multilocus analyses also were performed by considering a data set of 275 individuals pooled as a single population and obtained by excluding the southernmost and northwesternmost populations (3, 4, 25, and 26).

The extent of population subdivision was evaluated by calculating Wright's F_{ST} (39) and the related R_{ST} (37) indexes, with the latter taking explicit account of the mutation process that can occur at microsatellite loci. These two indexes were estimated across populations and between pairs of populations, according to Weir and Cockerham (38) and Michalakis and Excoffier (20), by using the program SPAGED1, version 1.1 (14).

A genetic distance matrix of pairwise F_{ST} values also was used to perform a hierarchical analysis of molecular variance (AMOVA) (7) with the ARLEQUIN

program (version 2.0; Genetics and Biometry Laboratory, University of Geneva, Switzerland [http://lgb.unige.ch/arlequin/]).

The possible presence of geographic structure of genetic variation in nuclear microsatellites in truffles was evaluated with four tests. First, we tested for the presence of phylogeographic structure by comparing R_{ST} estimates to R_{ST} values computed after 10,000 random permutations of allele types among alleles (13) were calculated using the SPAGEDI program. If R_{ST} is $>R_{ST}$ (permuted), then there is phylogeographic structure, i.e., on average, phylogenetically similar alleles are found in the same population more often than are randomly chosen alleles.

Second, we tested for a pattern of isolation by distance (31). A Mantel test with 1,000 random permutations was performed with the matrix of pairwise genetic differentiation between populations, using $R_{ST}/(1 - R_{ST})$, and a matrix of the ln(geographic distance) with the software GENALEX (version 5.1; Australian National University, Canberra, Australia [http://anu.edu.au/BoZo/GenALEX/]).

Third, a spatial autocorrelation analysis was performed with SPAGEDI. This analysis used R_{ST} and evaluated 18 spatial distance classes with similar sample sizes. The 95% confidence intervals were estimated for each distance class by 10,000 random permutations.

Finally, a simulated annealing procedure implemented in the spatial analysis of molecular variance (SAMOVA) algorithm (6) was used to define groups of populations that are geographically homogeneous and maximally differentiated from each other. The program repeatedly seeks the composition of a user-defined number, K , of groups of geographically adjacent populations that maximizes F_{CT} , the proportion of total genetic variance due to differences among groups of populations. The program was run for 10,000 iterations for $K \in \{2, \dots, 12\}$ from each of 250 random initial conditions. The simulated annealing process was repeated 250 times to ensure that the final configuration of the K groups is not affected by a given initial configuration (6). For each K , the configuration with the largest F_{CT} values after the 250 independent simulated annealing processes were completed was retained as the best grouping of populations.

RESULTS

Linkage disequilibrium analysis. When all 316 truffles were included, the two-locus analysis had significant linkage disequilibrium between many pairs of loci (15 out of the 21 two-locus combinations).

There also was a significant departure from linkage equilibrium ($I_A = 0.109$, $P < 0.001$) in the multilocus analysis. Linkage disequilibrium can result from reduced gene flow due to geographic separation. To assess the potential geographic bias in our combined analysis, these tests also were performed with samples in smaller geographic groups. When the two-locus LD was calculated for each of the 13 geographic groups, only one of the two-locus combinations had a significant LD, and no significant two-locus LD was detected when the 26 local populations were considered. Similarly, I_A was not significant when the randomizations were performed by shuffling alleles among samples within the 13 geographical groups and within the 26 local populations (P values of 0.065 and 0.119, respectively).

To evaluate the effect of sample size on LD values, the two-locus LD and I_A tests also were calculated for a data set of 275 individuals obtained by excluding from the whole data set

TABLE 2. AMOVA based on 316 truffles using seven SSR loci

Source of variation	df	Sum of squares	Variance component	% of total	P
Among regional groups	11	95	0.216	10	<0.01
Among populations within groups	14	42	0.112	5.4	<0.01
Within populations	290	509	1.75	84	<0.01
Total	315	646	2.08		

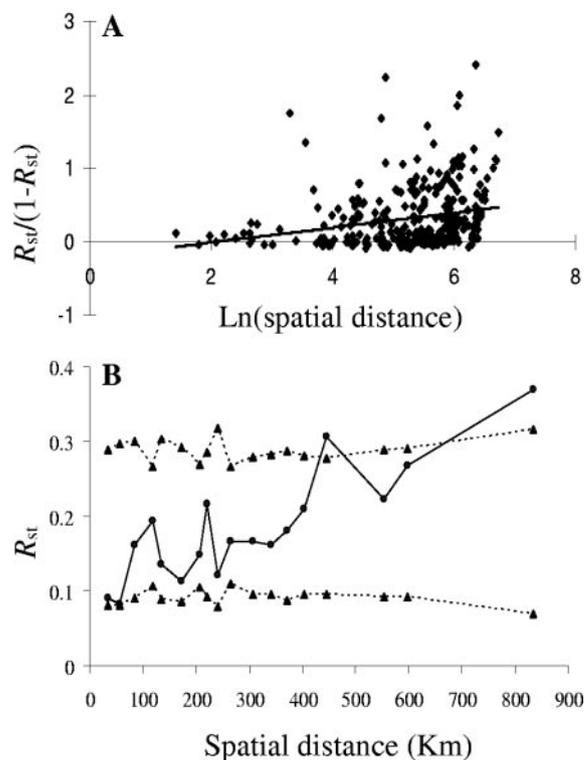


FIG. 1. (A) Correlation between genetic and geographical pairwise distances. (B) Average genetic distance (R_{ST}) for 18 spatial distance classes. Broken lines indicate the boundaries of the 95% confidence intervals estimated by permutation.

the populations 3, 4, 25, and 26 that SAMOVA proved to be genetically differentiated (see below) and thus were expected to contribute largely to LD. In this situation, only 5 out of 21 two-locus combinations resulted in significant LD ($P = 0.05$), and the I_A was not significant ($I_A = 0.036$, $P = 0.062$), suggesting that the decrease in LD values when populations are analyzed separately is not due to the reduced sample size.

Patterns within and among population variability. The seven SSR loci had 2 to 18 alleles per locus. Within each of the 351 truffles analyzed, only a single allele per locus was detected. Each multilocus genotype was then treated as a haplotype on the assumption that *T. magnatum* is a haploid fungus (see Discussion). The 26 populations had a mean number of alleles of 3.03 ± 0.12 , ranging from 1.86 ± 0.34 to 4.43 ± 0.97 , while the average allelic richness was 2.67 ± 0.06 , ranging from 1.74 ± 0.26 to 3.30 ± 0.65 (Table 1).

Based on AMOVA, most of the genetic variability is located at the within-population level, which accounts for about 84% of the total molecular variance, while 6% and 10% of the genetic variation were attributable to differences among populations within geographical groups and among the 13 geographical groups, respectively (Table 2). Overall, 263 haplotypes were detected within the 316 samples. Some haplotypes were found at multiple sites in the same region, and some were found in multiple populations or groups (see supplemental Table SA1).

Each population contains a relatively large number of haplotypes. Within an individual truffle ground, the genetic varia-



FIG. 2. Sampling localities and geographic distribution of genetic variability. Numbers refer to populations listed in Table 1. Diameters of the circles are proportional to the sample sizes. Differently shaded circles show genetically different population groups according to SAMOVA based on R_{ST} index and a K of 4.

tion decreased slightly, because some truffles probably originated from the same vegetative mycelium. For example, of the nine ascocarps harvested in Certaldo, only three haplotypes were found. However, from the 16 and 10 ascocarps from Ascoli and Pietralunga, 10 and 7 haplotypes were identified, respectively.

Most of the alleles were not unique to a population, but some alleles could be used to characterize truffles according to their geographic origin because of their differential frequencies in some groups of populations. For example, the frequency of allele 17 at the MA7 locus was high in southern populations 23, 24, 25, and 26 (0.2, 0.4, 0.4, and 1.0, respectively), while alleles 14, 24, 34, and 35 of MA4 were found only in individuals from populations 3, 4, and 5 from Piemonte, the northwesternmost area of sampling.

A marked and highly significant difference between populations was suggested by the F_{ST} and R_{ST} values, which were 0.15 and 0.22, respectively. A phylogeographic signal also was detected with permutation procedures in the total sample [R_{ST} of 0.22 versus $R_{ST}(\text{permuted})$ of 0.12, $P = 0.0005$]. The test of isolation by distance showed that among-population differentiation increased significantly with $\ln(\text{geographical distance})$ (Mantel test; $P < 0.006$), although the regression explains only 5.4% of the total variance ($R^2 = 0.054$) (Fig. 1A).

The spatial autocorrelation analysis allowed us to examine the extent of association in different geographic distance classes. This analysis confirmed the geographic pattern of genetic variation, with populations separated by >450 km being significantly further apart genetically than would be expected by chance (Fig. 1B). The results of SAMOVA confirmed the existence of geographic structure in the *T. magnatum* populations. In particular, when R_{ST} was used, southern populations

25 and 26 and northwestern populations 3 and 4 were clearly distinguishable from the others (Fig. 2).

DISCUSSION

Although a decline in production has triggered research into truffle cultivation, many basic aspects of truffle biology, such as the reproductive mode and the amount and distribution of population genetic variation, are not well understood. The limited saprophytic capabilities of truffle mycelia and the inability to obtain monosporic cultures or to cross mycelial strains under controlled conditions have slowed the direct assessment of the reproductive mode in these organisms.

We used codominant markers (polymorphic microsatellite loci) and strains sampled across the entire known range of the species to infer the genetic structure, population dynamics, and reproductive mode of *T. magnatum*, the most prestigious of the edible species in the genus *Tuber*. We found only a single allele per locus in each of the 351 truffles analyzed. These results are consistent with those of Bertault et al. (3) for *T. melanosporum*. They previously interpreted the absence of heterozygotes to mean that these fungi are functionally diploid with a strictly self-fertilizing reproductive system. We interpret these data to mean that the ascocarps of *T. magnatum* are haploid maternal tissue, as found for many ascomycetes. Two-locus and multilocus linkage disequilibrium analyses indicate that within both the *T. magnatum* geographical groups and the populations, extensive genetic exchange occurs. The relatively high number of multilocus genotypes within populations suggests that these fungi are not highly inbred. Within the same truffle grounds, differences between haplotypes usually occurred at multiple loci rather than at a single locus (see supplemental Table SA1), a pattern consistent with recombination in addition to mutation as a source for the variation observed in the haplotypes.

The truffle ascocarps are presumed to be primarily maternal tissue and thus would be haploid. Paternal DNA might not be as easily recoverable since it would be present only in the ascospores, and these spores are not usually broken during the DNA extraction process that we use. Thus, haploid maternal tissue will be the dominant component of the truffle and presumably represents the genotypes we analyzed in this study.

Significant linkage disequilibrium was found only when the entire truffle data set was evaluated as a whole and could be explained if there was significant geographic population subdivision. The F_{ST} and R_{ST} statistics and the correlation between genetic and geographic distance are consistent with genetic differentiation of populations, while the results of the R_{ST} permutation analysis are consistent with phylogeographic structure. If individual populations are removed from the analysis, then removal of the southernmost and two northwesternmost populations allows the rest of the populations to be grouped together.

The geographic distribution of *T. magnatum* tracks the post-glacial expansion of the species with which this fungus can establish a mutualistic symbiosis. Refugia for some *T. magnatum* host species (e.g., *Quercus*, *Corylus*, *Tilia*, and *Carpinus*) were located in central and southern Italy and spread northward as the ice receded (5, 28). A refugium for *T. magnatum* in central Italy, from which the northernmost and southernmost populations originated, could explain the results observed.

The genetic variation and geographic distribution of the SSR markers could enable the tracing and identification of *T. magnatum* populations. The geographic origin of truffles often is important to associations of truffle harvesters and local governments which are promoting economic and social development of rural and marginal areas. The genetic structure of populations of *T. magnatum* also has important ecological implications. Artificial fungal plantations often are established in naturally productive areas to counterbalance a sharp decline in wild truffle harvests. Potential problems related to microbial competition are largely unexplored with respect to truffle cultivation, and the consequences of the deliberate introduction of nonnative strains into an indigenous population have been serious for other edible mushrooms (16, 36). Hence, the profitable commercial practice of artificial fungal plantations may cause a loss of fungal biodiversity, and local strains with unique commercial characteristics might become extinct. Thus, when white truffle cultivation is planned in naturally productive areas, ecologically adapted truffle strains that are similar to those already present in the region should be used. Similar cautions also apply to *T. melanosporum* (21). The SSR markers that we used could be used to type strains of *T. magnatum* that are inoculated on commercially produced seedlings as well as for assessing the impact the introduced strains might have on native ectomycorrhizal populations.

The presence of genetic structure in populations of both of the most prestigious truffle species argues against the hypothesis that phenotypic and organoleptic differences among truffles of different origin are due solely to environmental factors (3). If the SSR loci examined are linked to the genes controlling these commercially important traits, then the SSR markers could be used to select strains with desirable characteristics for future commercial use. As our data suggest that truffles are not strictly self-fertilizing, the possibility of improving these mushrooms through conventional breeding protocols needs to be evaluated with more care and in greater detail. This study provides the first evidence suggesting that truffles are not strictly self-fertilizing organisms and should result in a significant reevaluation of the life cycle and biology of *T. magnatum* and other *Tuber* spp.

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