

Utilization of Mucus from the Coral *Acropora palmata* by the Pathogen *Serratia marcescens* and by Environmental and Coral Commensal Bacteria^{∇†}

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In recent years, diseases of corals caused by opportunistic pathogens have become widespread. How opportunistic pathogens establish on coral surfaces, interact with native microbiota, and cause disease is not yet clear. This study compared the utilization of coral mucus by coral-associated commensal bacteria (“*Photobacterium mandapamensis*” and *Halomonas meridiana*) and by opportunistic *Serratia marcescens* pathogens. *S. marcescens* PDL100 (a pathogen associated with white pox disease of *Acroporidae* corals) grew to higher population densities on components of mucus from the host coral. In an *in vitro* coculture on mucus from *Acropora palmata*, *S. marcescens* PDL100 isolates outgrew coral isolates. The white pox pathogen did not differ from other bacteria in growth on mucus from a nonhost coral, *Montastraea faveolata*. The ability of *S. marcescens* to cause disease in acroporid corals may be due, at least in part, to the ability of strain PDL100 to build to higher population numbers within the mucus surface layer of its acroporid host. During growth on mucus from *A. palmata*, similar glycosidase activities were present in coral commensal bacteria, in *S. marcescens* PDL100, and in environmental and human isolates of *S. marcescens*. The temporal regulation of these activities during growth on mucus, however, was distinct in the isolates. During early stages of growth on mucus, enzymatic activities in *S. marcescens* PDL100 were most similar to those in coral commensals. After overnight incubation on mucus, enzymatic activities in a white pox pathogen were most similar to those in pathogenic *Serratia* strains isolated from human mucosal surfaces.

Serratia is a gammaproteobacterium frequently isolated from waters, plants, and animals (7). Some isolates of *Serratia* are well-characterized symbionts of invertebrates. *Serratia marcescens* and *Serratia liquefaciens* have been identified as vertically transmitted symbionts of the sugar beet maggot (9). *Serratia* colonizes male and female reproductive tracts of the maggots, eggs, and pharyngeal filter. There, the bacteria are hypothesized to aid in metamorphosis by digesting chitinous puparial walls (9). In the gut of another insect, the diamond-back moth, strains of *S. marcescens* appear to live as commensals capable of modestly (5 to 8%) increasing growth rates of the host (8). *Serratia* strains have also been isolated from feces and cloacal swabs from clinically normal captive birds, but not from organs or carcasses of sick or diseased animals housed within the same facility (3, 20). *Serratia* spp. have also been linked to diseases of invertebrate animals and their larvae (for reviews, see references 7, 15, and 21). To cause diseases in nematodes and flies, *S. marcescens* first colonizes the intestines, degrades cells of the alimentary tract and then spreads to other organs (14, 21). There are, however, exceptions to this

mode of infection. *Serratia entomophila*, the causal agent of amber disease in grubs, grows within the alimentary tract of the animal to >10⁶ CFU. However, bacteria neither attach to nor colonize surfaces of the gut; rather, they adhere to gut contents (10) and cause the appearance of signs by producing the Sep toxin that inhibits accumulation of the insect’s digestive serine proteases and disrupts the cytoskeletal network (6). It appears, therefore, that various isolates of *Serratia* are capable of entering into a full range of interactions (from mutualistic to commensal to pathogenic) with their animal hosts (for reviews, see references 7, 15, and 21).

A strain of *S. marcescens*, PDL100, was shown to be associated with white pox disease of the threatened Caribbean coral *Acropora palmata* (22, 27). White pox disease results in coral tissue necrosis, exposing carbonate skeleton at a rate of 2.5 cm² day⁻¹ (22). It is not yet clear how *S. marcescens* PDL100 colonizes and infects corals. It is likely that to cause disease, the pathogen first needs to colonize and establish within the coral surface mucus layer.

The coral surface mucus layer contains polymers of mixed origin. Coral mucus is made in the mucocytes of the polyp, where the photosynthate produced by the coral symbiotic dinoflagellate *Symbiodinium* spp. is converted into polymers that are excreted onto the coral surface (for a review, see reference 2). A glycoprotein is the major component of coral mucus from both hard and soft corals (16, 17, 19). The com-

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TABLE 1. Bacterial strains used in the study

Strain	Description
<i>Halomonas meridiana</i> 33E7	Isolated from <i>Acropora palmata</i> mucus (23); identity confirmed by 16S rRNA gene sequencing
<i>Photobacterium mandapamensis</i> 33C12	Isolated from <i>Acropora palmata</i> mucus (23); identity confirmed by 16S rRNA gene sequencing
<i>Photobacterium leiognathi</i> 33G12	Isolated from <i>Acropora palmata</i> mucus (23); identity confirmed by 16S rRNA gene sequencing
<i>S. marcescens</i> PDL100	Isolated from a white pox lesion on <i>Acropora palmata</i> (22, 28)
<i>S. marcescens</i> MG1	Isolated from a rotten cucumber
<i>Serratia</i> sp. strain ATCC 39006	Cheesequake Salt Marsh (NJ) channel isolate, pigmented
<i>S. marcescens</i> ATCC 43422	Human throat isolate, pigmented
<i>S. marcescens</i> ATCC 43820	Human urine isolate
<i>S. marcescens</i> EL31	Isolated from Florida Keys wastewater
<i>S. marcescens</i> EL34	Isolated from Eden Pines (Florida Keys) canal water, pigmented
<i>S. marcescens</i> EL139	Isolated from Florida Keys wastewater
<i>S. marcescens</i> EL202	Isolated from Higgs Beach (Key West)
<i>S. marcescens</i> EL206	Isolated from canal water by Mote Marine Tropical Research Laboratory (Summerland Key, FL)
<i>S. marcescens</i> EL266	Isolated from Florida Keys wastewater
<i>S. marcescens</i> EL368	Isolated from a seabird (Key Largo, FL)
<i>S. marcescens</i> EL402	Isolated from a seabird (Florida Keys)

position of the glycoprotein differs among coral species (4, 17). The mucus polymer of *Acropora formosa*, for example, contains 36 to 38% of neutral sugars, 18 to 22% of amino sugars, and 19 to 30% of amino acids; lipids make up 4.2% of the polymer (17). In the mucus of *A. formosa*, the oligosaccharide decorations (two to four sugar residues long) are attached to the polypeptide backbone by an O-glycosidic link to serine or threonine through the carbon 1 of mannose (16). The glycoproteins from *A. formosa* and *Pseudopterogorgia americana* corals contain terminal arabinose residues linked by a β 1 \rightarrow 2 or β 1 \rightarrow 3 bond. In the mucus of acroporid corals, arabinose, *N*-acetyl-glucosamine, mannose, glucose, galactose, *N*-acetyl-galactosamine, and fucose were the major sugars; serine and threonine were the major amino acids (4, 17). The elucidation of the chemical structure of coral mucus is complicated by the fact that the mucus contains excretions of coral mucocytes, extracellular substances produced by the associated microbiota as well as oligomers that may result from the degradation of these polymers (for reviews, see references 2 and 24).

In this study, we tested the hypothesis that *S. marcescens* PDL100 is capable of a more extensive utilization of *A. palmata* mucus than other environmental or pathogenic isolates of *S. marcescens*. This hypothesis is based on the recent discoveries that pathogenic and commensal host-associated bacteria differ in their patterns of carbon source utilization during growth on components of the mucus that lines host surfaces (5, 26). These different strategies of mucus utilization may allow pathogenic bacteria to outcompete native residents and establish within the host's mucosa (5, 13, 26). To test this hypothesis, growth of the strain PDL100 on coral mucus and enzymatic activities induced during growth on mucus were assayed and compared to those of pathogenic and environmental isolates of *S. marcescens* and three native coral-associated bacteria.

MATERIALS AND METHODS

Bacterial strains and culture conditions. Bacterial strains used in this study are listed in Table 1. *Escherichia coli* and *Serratia marcescens* isolates were grown in LB broth (Fisher Scientific, Pittsburgh, PA). Bacteria isolated from *A. palmata* mucus were routinely grown in GASW broth (356 mM NaCl, 40 mM MgSO₄, 20 mM MgCl₂ · 6H₂O, 8 mM KCl, 60 μ M K₂HPO₄, 33 μ M Tris, and 7 μ M FeSO₄, with 0.05% peptone, 0.2% yeast extract, and 2.0% glycerol [pH 7.0]) or on GASW medium solidified with 1.5% agar (GASWA) (Fisher Scientific) (23).

Unless otherwise stated, seawater used in the experiments was collected at Crescent Beach (29°45'56"N, 81°15'11"W) and sterilized through a 0.22- μ m filter. As needed for experiments, seawater samples were buffered with 10 mM HEPES to pH 7.0.

Coral mucus samples were collected from different, apparently asymptomatic *A. palmata* colonies at Looe Key Reef (24°33'75"N, 81°24'05"W) in September 2006, winter of 2007, and February 2008 with a needleless syringe (23). *Montastraea faveolata* mucus samples were collected in a similar manner at the Dry Tortugas (24°37'43"N 82°52'24"W) in May 2008. Mucus samples were stored as aliquots at -80°C . As needed, aliquots were thawed and sterilized in polypropylene tubes by UV irradiation (254 nm) for 20 min and by filtration through a glass fiber (GFC) filter and then through 0.45- μ m and 0.22- μ m filters. Mucus samples were then separated into low- and high-molecular-mass fractions with VivaSpin-15 spin dialysis assemblies (VivaScience, Stonehouse, United Kingdom), following the manufacturer's instructions. The high-molecular-mass fraction was brought up to volume in artificial seawater (Instant Ocean, Mentor, OH) as described previously (13).

To evaluate growth on mucus components, overnight bacterial cultures (grown to an optical density at 600 nm of \sim 1.9 in either LB or GASW broth) were pelleted, washed three times in phosphate-buffered saline (pH 7.0) and resuspended in phosphate-buffered saline. These inocula were mixed with seawater containing "total mucus," its high- or low-molecular-mass mucus fractions resulting in the initial population densities of 10^2 to 10^3 CFU ml⁻¹. Cells were prepared in a similar manner for coculture experiments; they were diluted to 10^2 CFU ml⁻¹ and then mixed 1:1. All cultures were incubated at 30°C with shaking at 180 rpm. Growth was monitored by serially diluting cultures and plating them onto either LB agar or GASWA plates at the time intervals indicated below. To differentiate *S. marcescens* from coral commensals in cocultures, individual colonies were picked from LB and/or GASWA plates and patched onto xylose lysine deoxycholate agar (Oxoid). Differences in bacterial growth were compared using a one-way analysis of variance and Tukey's honestly significant difference (HSD) post hoc test with type I error significance at $\alpha = 0.05$.

The extent to which the bacteria hydrolyzed polymers in mucus was examined using a 2,2'-bicinechonic acid (BCA) assay (29). Briefly, spent cultures and controls were centrifuged, and equal volumes of cell-free supernatants were mixed with solution A (0.67 M Na₂CO₃ · H₂O, 0.32 M NaHCO₃, 2.22 mM BCA) and solution B (5 mM CuSO₄ · 5H₂O, 12 mM L-serine). The reaction mixtures were heated to 100°C for 15 min, and A₅₆₀ was measured (29). The reducing end equivalents were calculated using a regression equation from a standard curve using known concentrations of L-arabinose ($y = 0.0073x + 0.0031$; $R^2 = 0.9988$).

Enzymatic assays. Two overnight cultures of each isolate were grown in LB or GASW broth to an optical density at 600 nm of 2.0. Cells were pelleted, washed in filter-sterilized seawater, resuspended in the same volume of buffered seawater, and starved at 30°C with shaking for 3 days. Following starvation, 1 ml of cells was added to 2 ml of either 1 \times coral mucus or 10 mM HEPES-buffered seawater. Treatments of the negative control (coral mucus alone) and a blank (buffered seawater) were carried out in parallel. Enzymatic assays were conducted after 2 or 18 hours of incubation in mucus (at 30°C), by following established protocols (12, 18). Corresponding *p*-nitrophenyl enzymatic substrates were purchased from Sigma-Aldrich. All enzymatic reactions were conducted for exactly 24 h. Cellular

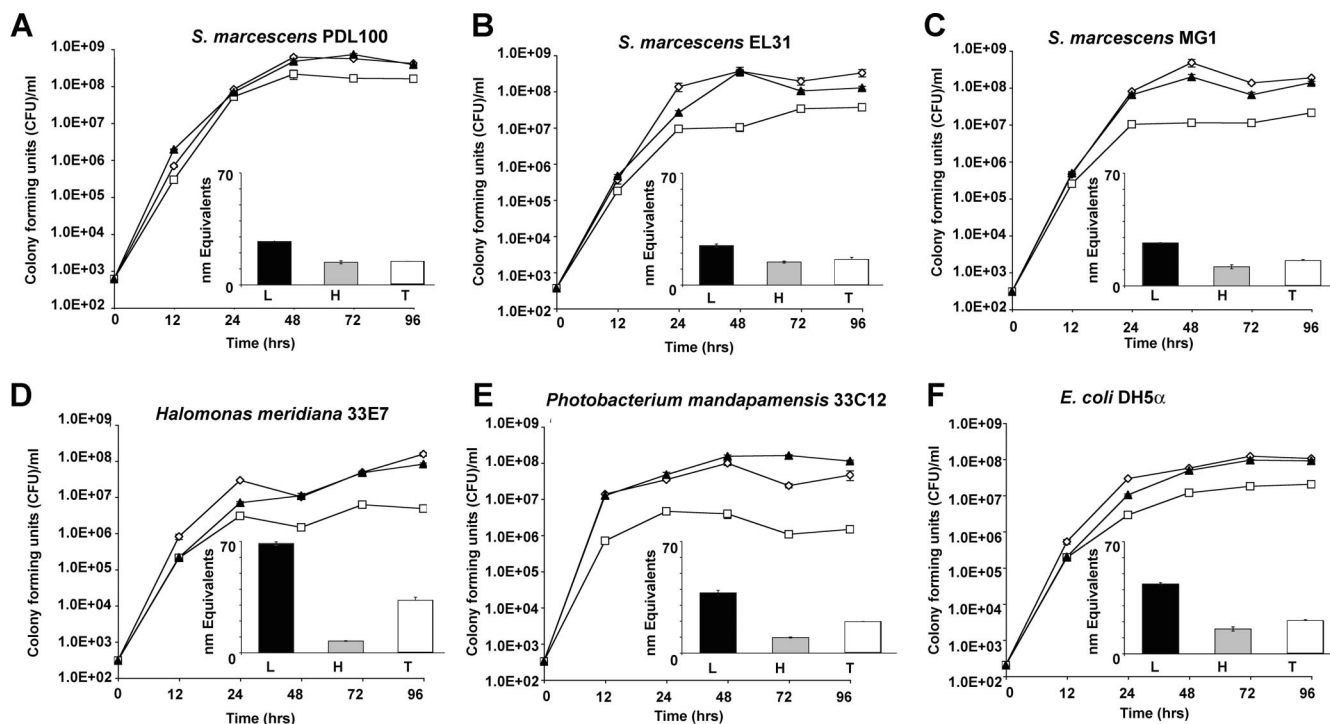


FIG. 1. Bacterial growth on the mucus of *Acropora palmata*. *Serratia marcescens*, coral commensal bacteria, and *E. coli* DH5 α were grown for 96 h in seawater containing approximately 60 mg ml⁻¹ (wt/vol) total mucus (open diamonds) or its high-molecular-mass (>15 kDa; open squares) or low-molecular-mass (<15 kDa; closed triangles) fractions. (Insets) The extent to which the bacteria hydrolyzed substrates in mucus was examined using a BCA assay as described in Materials and Methods. Black bars indicate accumulation of reducing group equivalents in the low-molecular-mass fraction of mucus; gray bars indicate accumulation of reducing group equivalents in the high-molecular-mass fraction of mucus; and white bars indicate accumulation of reducing group equivalents in total mucus. (A) Growth of *S. marcescens* PDL100, a pathogen associated with white pox disease of *A. palmata*. PDL100 reached $\sim 2 \times 10^8$ CFU ml⁻¹ on all mucus treatments, which was significantly higher than the densities of all of the other strains (Tukey's HSD P value of <0.0025). (B) Growth of *S. marcescens* EL31 isolated from wastewater in the Florida Keys. (C) Growth of *S. marcescens* MG1 isolated from a rotten cucumber. (D) Growth of *H. meridiana* 33E7 isolated from *A. palmata* mucus. (E) Growth of *P. mandapamensis* 33C12 isolated from *A. palmata* mucus. (F) Growth of *E. coli* DH5 α on mucus and its components. Graphs show the results of a representative experiment; error bars indicate standard errors from three parallel independent cultures.

debris and unused enzymatic substrate were pelleted at a relative centrifugal force of 16,000 $\times g$. The supernatants were transferred to a clear polystyrene 96-well plate, and A_{405} was measured on a Victor3 plate reader (Perkin Elmer, Shelton, CT). Enzymatic activities in the mucus-only control were measured and subtracted from those of all other treatments; the activity was then calculated using modified Miller units (A_{405}/A_{590}).

To compare the totality of enzymatic activities induced during the growth of each isolate on *A. palmata* mucus, the activities were scaled to those of a starved control for each strain, and joining tree analysis was performed on the natural log of the resulting induction value. Hierarchical clustering was done separately at each time period, using Ward's method (30). The dendrograms were constructed using Euclidean distance metric, which serves to emphasize the similarities based on absolute (rather than relative) values of induction.

Nutrient utilization profiling with Biolog EcoPlates. Isolates were grown in either LB or GASW broth overnight at 30°C with shaking. Cells were then pelleted and washed twice with 0.22- μ m-filter-sterilized seawater. Cells were then resuspended in 2 volumes of filter-sterilized seawater and starved for 24 h at 30°C and then inoculated into EcoPlates (Biolog Inc., Hayward, CA). A_{590} was monitored using a Victor3 plate reader every 24 h for 72 h. For each substrate, the average well color development (AWCD) was calculated after the final reading and then converted into binary format. To evaluate the degree of similarity between *S. marcescens* PDL100 and each of the tested isolates, the Jaccard similarity coefficient and the odds ratio were used. Given the small sample size (contingency tables comprised of binary responses from 31 substrates and a control), both the odds ratio and the associated 95% confidence intervals were estimated after employing a modified Gart interval sample size correction (1). All statistical analyses were performed using Statistica version 8.0 (StatSoft, Inc., Tulsa, OK).

RESULTS AND DISCUSSION

Bacterial growth on mucus of *A. palmata*. All tested strains utilized coral mucus as a growth substrate (Fig. 1). When grown on the crude preparation of "total mucus," the white pox pathogen, *Serratia marcescens* PDL100, reached final population densities of 7×10^8 CFU ml⁻¹, while other *S. marcescens* isolates reached somewhat lower final densities of 1.5×10^8 to 5×10^8 CFU ml⁻¹ (Tukey's HSD P value of <0.0025). Two unrelated bacteria ("*Photobacterium mandapamensis*" 33C12 and *Halomonas meridiana* 33E7) that were previously characterized as dedicated *A. palmata* commensals (23) reached 1.6×10^8 CFU ml⁻¹ and 8×10^7 CFU ml⁻¹ (respectively). These observations are generally consistent with the report that crude preparations of coral mucus supported bacterial cultures with 10^8 to 10^9 CFU ml⁻¹ (26). *Escherichia coli* DH5 α , used as a negative control in this work, reached 9.4×10^7 CFU ml⁻¹ when grown on crude *A. palmata* mucus (Fig. 1F). The observation that *E. coli* DH5 α , not known for its ability to extensively utilize complex carbohydrate polymers, grew on mucus as well as coral-associated bacteria suggested either that the mucus sample contained a significant amount of monomers or that proteinaceous components of mucus were

sufficient to support growth of various bacteria, including a laboratory strain of *E. coli*.

Bacterial growth on size-fractionated mucus of *A. palmata*.

To test whether the mucus sample contained a significant proportion of low-molecular-mass components that could support growth of generalists, it was dialyzed to separate low-molecular-mass (<15 kDa) and high-molecular-mass (>15 kDa) fractions. All tested strains of *S. marcescens* reached the final densities of 2×10^8 to 6×10^8 CFU ml⁻¹ when grown on the low-molecular-mass fraction of *A. palmata* mucus (Fig. 1). *H. meridiana*, *P. mandapamensis*, and *E. coli* DH5 α reached 1×10^8 to 1.5×10^8 CFU ml⁻¹. This is consistent with the hypothesis that low-molecular-mass compounds in coral mucus are easily metabolizable by most bacteria.

We hypothesized that the high-molecular-mass fraction of mucus consists of polymers that are not readily available to all microbes. As shown in Fig. 1A, *S. marcescens* PDL100 reached the final density of 2×10^8 CFU ml⁻¹, which is approximately seven times higher (Tukey's HSD *P* value of <0.0025) than the final population densities of the two other *S. marcescens* isolates (MG1, isolated from a rotten cucumber, and EL31, isolated from Florida Keys wastewater). Similarly to the *S. marcescens* strains MG1 and EL31, *E. coli* DH5 α reached 3×10^7 CFU ml⁻¹. *H. meridiana* 33E7 and *P. mandapamensis* 33C12 grew only to 1×10^6 to 6×10^6 CFU ml⁻¹, approximately 50 to 100 times less than *S. marcescens* PDL100 (Fig. 1D and E). In general, the extent to which coral commensals utilized mucus is consistent with the studies of the degradation of the high-molecular-mass component (6 to 8 kDa) of coral mucus, which demonstrated that bacterial populations in situ reached $\sim 10^6$ CFU ml⁻¹ when grown on *Acropora* coral mucus in size exclusion cages (28).

Patterns of mucus utilization by bacteria. BCA assays were carried out for each mucus fraction (low-molecular-mass, high-molecular-mass, and total mucus) after bacterial growth for 96 h. The BCA assay would determine both the protein content and the presence of reducing ends of sugars (29). As expected, the fractions of mucus differed in the numbers of the BCA-reactive species prior to incubation with bacteria; the number of BCA-reactive species was highest in the low-molecular-mass fraction of mucus (77 equivalents in low-molecular-mass fractions, 13 equivalents in high-molecular-mass fractions, and 44 equivalents in the crude preparation of "total mucus"). This supports the observations that in addition to the sulfated glycoprotein polymer, mucus samples contain a significant amount of low-molecular-mass compounds (16, 17).

Upon incubation with bacteria, the number of BCA-reactive species in mucus and its low-molecular-mass fraction decreased two- to threefold in the spent cultures of all *S. marcescens*, *E. coli*, and *P. mandapamensis* isolates. The number of BCA-reactive species also decreased (although to a more modest degree) after growth of *H. meridiana*. These results indicate that the components of the low-molecular-mass fraction were generally mineralized in similar fashions by the different bacteria used in this experiment. The reduction in the number of BCA-reactive species in the low-molecular-mass fraction may suggest that carbohydrate mono- and oligomers as well as short peptides that may be present in the low-molecular-mass fraction were mineralized.

As it relates to carbohydrate-degrading activities, an in-

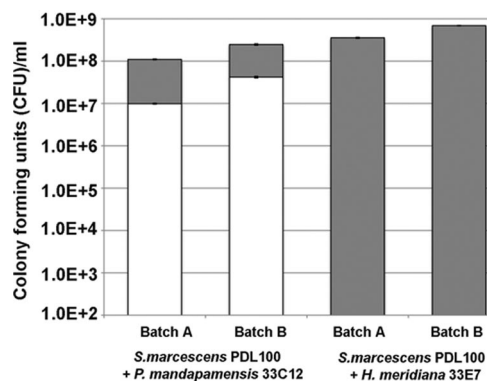
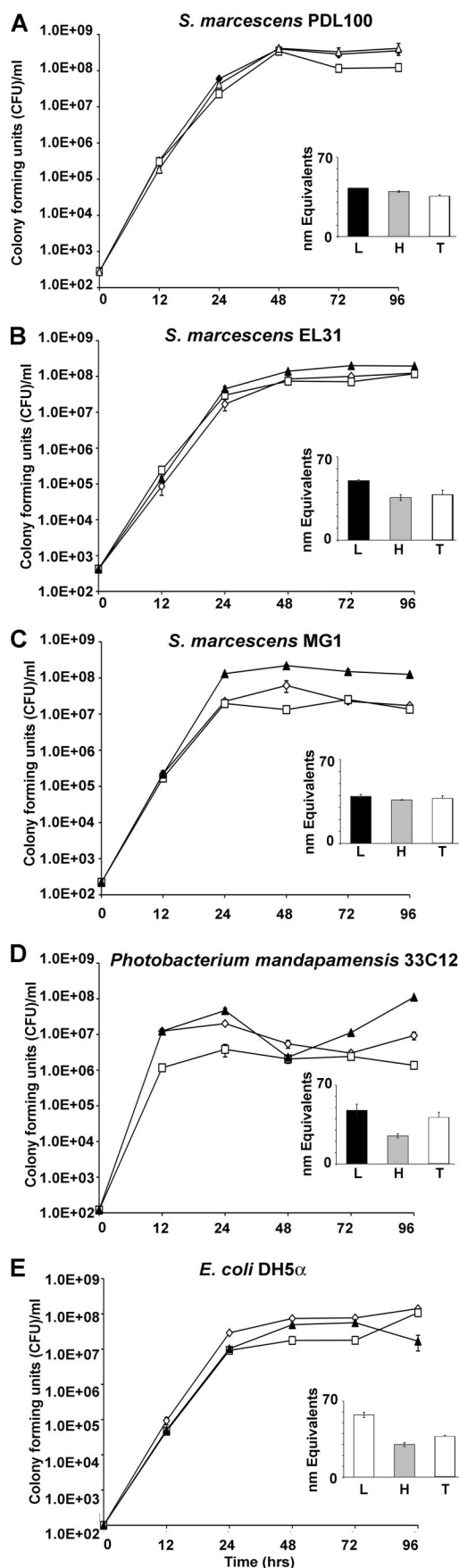


FIG. 2. Growth of *S. marcescens* PDL100 and coral commensals in an in vitro coculture on mucus. Starved washed cultures of *S. marcescens* PDL100 were coinoculated into sterilized mucus with either *P. mandapamensis* 33C12 or *H. meridiana* 33E7. Two batches of mucus samples harvested from different *A. palmata* colonies in the winter of 2007 were used in these assays. Initial inocula were ~ 100 CFU ml⁻¹ for all bacteria. After 96 h and 7 days, aliquots were dilution plated onto GASW and LB agar and then patched onto xylose lysine deoxycholate agar (Oxoid) to distinguish between *S. marcescens* and coral commensals. Growth of the coral commensals is indicated by white bars; growth of *S. marcescens* PDL100 is indicated by gray bars. Error bars indicate standard errors of three technical replications.

crease in the number of BCA-reactive species in the sample with the high-molecular-mass fraction of mucus could be indicative of an endoglycanase activity, while exoglycanases could be identified by no significant change in the number of reducing group equivalents (assuming that each cleaved monomer is then metabolized). The number of BCA-reactive species did not change significantly in the spent cultures of *S. marcescens* PDL100, MG1, or EL31 or *E. coli* DH5 α grown on the high-molecular-mass fraction of mucus as the sole carbon source (Fig. 1). The number of BCA-reactive species was strongly reduced after the two coral isolates were grown on high-molecular-mass mucus. The results of BCA assays of the spent high-molecular-mass fraction suggest that *Serratia* isolates break down mucus through the action of exoglycanases, while the two native coral isolates may be preferentially utilizing the protein backbone of the mucus glycoprotein or lipids found in coral mucus (4, 17).

***S. marcescens* PDL100 dominates mucus microcosms in vitro.** To test whether the growth to higher final cell numbers on coral mucus corresponds with the ability to outcompete other bacteria, *S. marcescens* PDL100 was coinoculated with either *H. meridiana* 33E7 or *P. mandapamensis* 33C12 into mucus in seawater. In cocultures on two different batches of *A. palmata* mucus samples (both collected in the winter of 2007), after 96 h, *S. marcescens* PDL100 outgrew *P. mandapamensis* at a ratio of 8 to 2 (Fig. 2). In cocultures with *H. meridiana*, the coral commensal was not detectable after 96 h (Fig. 2). In monocultures on mucus, *S. marcescens* PDL100 reached 4.4×10^8 to 6.5×10^8 CFU ml⁻¹, *P. mandapamensis* reached 1.8×10^7 to 2.4×10^7 CFU ml⁻¹, and *H. meridiana* reached 3.5×10^7 to 4.4×10^7 CFU ml⁻¹, generally consistent with the growth on another batch of mucus samples (Fig. 1). These results suggest that in the



absence of other biotic factors, this opportunistic coral pathogen can outgrow representative coral microbiota.

Growth and utilization of mucus from a nonhost coral by the *Serratia* isolates. To test whether *S. marcescens* PDL100 would also grow better on mucus from a nonhost coral, *Montastraea faveolata*, experiments similar to those shown in Fig. 1 were carried out. All *Serratia* strains and a coral commensal tested reached 10^8 CFU ml⁻¹ when grown on a “total mucus” sample of *M. faveolata*, significantly higher than the density of *E. coli* DH5α (Tukey’s HSD *P* value of <0.0025). As shown in Fig. 3, on the high-molecular-mass fraction of the *M. faveolata* mucus, *E. coli* DH5α and *S. marcescens* PDL100 and EL31 reached the same final population density. Growth rates of both PDL100 and EL31 were similar on all mucus components, reaching 10^8 CFU ml⁻¹, whereas MG1 reached densities that were 7- to 10-fold lower on the low- and high-molecular-mass fractions (Fig. 3). The *A. palmata* commensal, *P. mandapamensis* 33C12, reached densities approximately 10- to 100-fold lower than those of PDL100 on low- and high-molecular-mass *M. faveolata* mucus components.

BCA assays determined whether there were differences in the patterns of substrate degradation during growth on *M. faveolata* mucus. Similarly to results with *A. palmata* mucus, the amount of BCA-active species was large in the low-molecular-mass fraction of mucus (43 equivalents in low-molecular-mass, 22 equivalents in high-molecular-mass, and 48 equivalents in crude “total” mucus). *Serratia* isolates had similar abilities to generate the reducing group equivalents when grown on the low-molecular-mass fraction; however, the number of BCA-reactive species was not decreased as observed with *A. palmata* mucus, suggesting that this fraction contained oligomers that were degraded by exoglycanases. There was an increase in the number of BCA-reactive species for all bacteria grown on the high-molecular-mass fraction of *M. faveolata* mucus, which could be indicative of an endoglycanase activity.

S. marcescens PDL100 was similar to other *Serratia* strains and *E. coli* in its growth on mucus from a nonhost coral, *M. faveolata*. This observation suggests that the ability to cause disease in acroporid corals may be due, at least in part, to the ability of the strain PDL100 to build to higher population numbers within the mucus surface layer of its acroporid host.

FIG. 3. Bacterial growth on mucus of *Montastraea faveolata*. Isolates of *Serratia marcescens*, *P. mandapamensis* 33C12, and *E. coli* DH5α were grown on mucus from a nonhost coral, *M. faveolata*. Bacteria were seeded into seawater containing 60 mg ml⁻¹ (wt/vol) total mucus (closed triangles) or high-molecular-mass (>15 kDa; open squares) or low-molecular-mass (<15 kDa; open diamonds) fractions. Growth was monitored by dilution plating. (Insets) The extent to which the bacteria hydrolyzed substrates in mucus was examined using a BCA assay as described in Materials and Methods. White bars indicate accumulation of reducing group equivalents in the low-molecular-mass fraction of mucus; gray bars indicate accumulation of reducing group equivalents in the high-molecular-mass fraction of mucus; and black bars indicate accumulation of reducing group equivalents in total mucus. (A) Growth of *S. marcescens* PDL100. (B) Growth of *S. marcescens* EL31. (C) Growth of *S. marcescens* MG1. (D) Growth of *P. mandapamensis* 33C12. (E) Growth of *E. coli* DH5α. Graphs show the results from a representative experiment; error bars indicate standard errors from three parallel independent cultures.

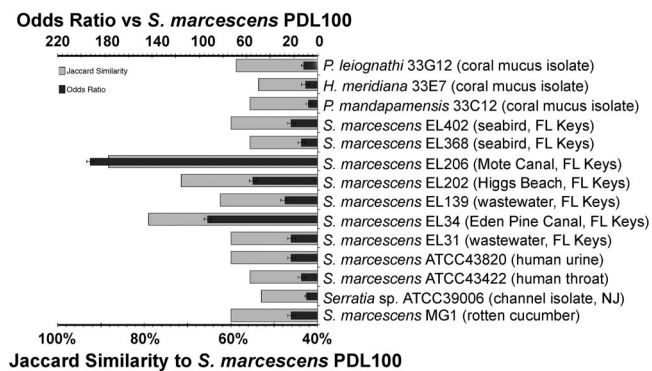


FIG. 4. Metrics of correspondence between Biolog EcoPlate assays for *S. marcescens* PDL100, other *S. marcescens* isolates, and three coral commensals. The results of the Biolog assay were converted into binary data (see Table S2 in the supplemental material), and values for the strain PDL100 were compared with those for other *S. marcescens* strains, using the Jaccard similarity coefficient and the odds ratio. Isolates with similar metabolic capabilities have high Jaccard similarity and odds ratio. Statistical analyses were carried out using Statistica version 8.0 (StatSoft, Inc., Tulsa, OK) as described in Materials and Methods.

Nutrient utilization profiling. To test the hypothesis that differences in the abilities to grow on *A. palmata* coral mucus were due to the general metabolic capabilities of the strains, their abilities to utilize carbon and nitrogen sources were compared using Biolog EcoPlates. The EcoPlate contains *N*-acetyl-D-glucosamine, L-asparagine, L-serine, and L-threonine—all known to occur in the mucus of acroporid corals (4, 17). To broaden the range of tested strains, additional environmental isolates of *S. marcescens* collected from various locations in the Florida Keys and from the American Type Culture Collection and an additional coral commensal were also included in this test (Table 1; also see Table S2 in the supplemental material).

All tested isolates respired when incubated on *N*-acetyl-D-glucosamine and L-asparagine. All strains (with the exception of *Serratia* sp. strain ATCC 39006) respired when incubated on L-serine and L-threonine (see Table S2 in the supplemental material), the two amino acids that make up the protein backbone of the mucus glycoprotein in acroporids. Consistent with the earlier report (22), *S. marcescens* PDL100 utilized acetate, citrate, D-fructose, D-glucose, maltose, and D-sorbitol and was unable to ferment α -keto butyrate, cellobiose, and α -D-lactose. The substrate utilization profile of PDL100 corresponded most strongly with those of the two Florida Keys canal water isolates, EL206 and EL34, and a beach water isolate, EL202 (Fig. 4; also see Table S2 in the supplemental material). The profile of PDL100 correlated the least with those of the three coral commensals and a Cheesequake Salt Marsh (NJ) channel isolate, *Serratia* sp. strain ATCC 39006 (Fig. 4).

The rate of AWCD in Biolog Ecoplates (see Fig. S1 in the supplemental material) was used to test whether differences in the general metabolic rates could explain higher growth rates and higher population densities reached by *S. marcescens* PDL100 on the mucus of *A. palmata*. As shown in Fig. S1 in the supplemental material, the rate and the final AWCD were the highest in *S. marcescens* EL202. The rates and the final AWCDs were similar in the white pox isolate *S. marcescens* PDL100 and in MG1. These results further support the con-

clusion that the ability of *S. marcescens* PDL100 to more efficiently utilize mucus from its host coral cannot be simply explained by generally higher metabolic rates in this pathogen or by the general ability of this strain to utilize a more diverse set of carbon and nitrogen sources.

Enzyme induction in response to growth on coral mucus. Because the ability of *S. marcescens* PDL100 to grow on *A. palmata* mucus to higher final population densities was not explained by either its metabolic rates or its general ability to grow on generic carbon and nitrogen sources, the subsequent experiments focused on identifying the potentially specific mechanisms underlying interactions of this pathogen with mucus from its coral host. An earlier study demonstrated that the in situ degradation of coral mucus involved activities of as-yet-uncharacterized proteases, esterases, and glycosidases (28). A hypothesis that *S. marcescens* PDL100 relies on a different suite of mucus-degrading enzymatic activities or that these activities are stronger than those of other bacteria was tested. This study focused on the glycosidases, because carbohydrates are a major part of mucus and account for approximately 60% of the mucus glycoprotein in acroporid corals (17).

Incubation with coral mucus induced the broadest range of enzymatic activities in *S. marcescens* MG1 and *Serratia* sp. strain ATCC 39006; their enzymatic activities were also the strongest (see Table S1 in the supplemental material). Even though only few activities in *S. marcescens* PDL100 were induced after incubation with coral mucus (see Table S1 in the supplemental material), this strain appeared to have strong constitutive α -D-glucopyranosidase and *N*-acetyl- β -D-galactosaminidase activities that were 2 to 10 times higher than in other isolates. In *S. marcescens* PDL100, a 2-hour incubation with coral mucus repressed either the accumulation or the function of α -D-galactopyranosidase, α -D-xylopyranosidase, β -D-fucopyranosidase, β -D-glucopyranosidase, and β -L-arabinopyranosidase. Some of the same enzymes were also downregulated in other isolates. We do not yet know whether the reduced activity of these enzymes was due to the absence of the potential substrates in mucus or due to the presence of substances that may inhibit induction, accumulation, or activities of these enzymes.

There were differences in temporal pattern of enzymatic induction; different enzymatic activities were induced after 2 and 18 hours of incubation on coral mucus. As shown in Table S1 in the supplemental material, after 2 hours of incubation on mucus, β -L-arabinofuranosidase was strongly induced in seven *S. marcescens* isolates, β -D-fucopyranosidase was upregulated in five *S. marcescens* isolates, and α -L-fucopyranosidase was induced in four *S. marcescens* isolates and in the coral commensal *H. meridiana*; xylopyranosidases were induced in four *S. marcescens* isolates and in the coral commensal *P. mandapamensis*. These same enzymatic activities were strongly decreased after 18 hours of incubation on mucus. β -D-Galactopyranosidases and β -D-glucopyranosidases were induced in three *S. marcescens* isolates after 18 h of incubation on mucus. This extended (18 h) incubation of coral commensals on mucus led to the induction of D-galactopyranosidases, β -D-glucopyranosidase, arabinofuranosidases, β -D-fucopyranosidase, β -D-xylopyranosidase, and *N*-acetyl-galactopyranosidase (see Table S1 in the supplemental material).

To further compare patterns of enzymatic induction during

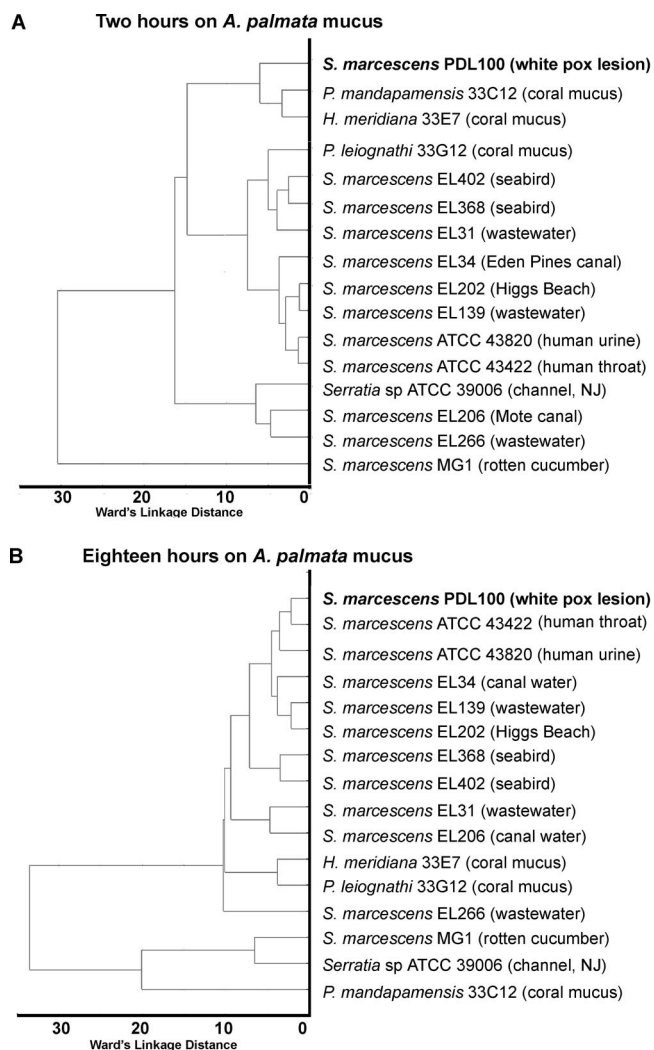


FIG. 5. Clustering of strains based on enzymatic activities induced during growth on coral mucus. The hierarchical tree plot was constructed using Ward's linkage as the joining rule and Euclidean distance metric. (A) The hierarchical tree plot of enzymatic activities induced in *S. marcescens* isolates and coral commensals after 2 hours of incubation on the total mucus of *A. palmata*. (B) The hierarchical tree plot of enzymatic activities induced in *S. marcescens* isolates and coral commensals after 18 hours of incubation on the mucus of *A. palmata*. Statistical analyses were carried out using Statistica version 8.0 (StatSoft, Inc., Tulsa, OK) as described in Materials and Methods.

growth on coral mucus, the data in Table S1 in the supplemental material were analyzed using Ward's linkage as the joining rule and a Euclidean distance metric, as described in Materials and Methods. As shown in Fig. 5, during the initial 2-h incubation on the mucus of *A. palmata*, *S. marcescens* PDL100 was most similar to the two other *A. palmata* isolates, *P. mandapamensis* 33C12 and *H. meridiana* 33E7 (Fig. 5A), with the rotten-cucumber isolate (*S. marcescens* MG1) being an extreme outlier. After 18 h of incubation on coral mucus, the pattern of enzymatic induction in *S. marcescens* PDL100 was most similar to those of the human pathogens *S. marcescens* ATCC 43422 and ATCC 43820 (Fig. 5B). In its interactions with coral mucus, *S. marcescens* PDL100 was distinct from

other *S. marcescens* isolates isolated from throughout the Florida Keys from wastewaters, canals, beaches, or seabirds.

Conclusions. (i) Differences in mucus utilization reveal an ecophysiological basis of white pox. Of the strains tested in this study, *S. marcescens* PDL100 grew to the highest final population densities on the mucus of *A. palmata* and on its low- and high-molecular-mass components (Fig. 1). The pattern of mucus-induced enzymatic activities in PDL100 was distinct from those of other environmental *Serratia* isolates (Fig. 5). A comparison of the totality of mucus-induced enzymes in *S. marcescens* PDL100 with those induced in other bacteria suggests that during early stages of mucus colonization, the white pox pathogen was most similar to coral commensals. These differences in utilization of the available carbon sources between the pathogens and commensals occupying the same ecological niche have also been observed in pathogenic and commensal *E. coli* isolates (5).

(ii) Patterns of mucus utilization by bacteria shed light on mucus composition. The compositions of mucus differ among coral species (4, 17, 19), although they appear to be consistent in corals of the same species harvested in different locations and at different depths (11). The structures of the polymers from *A. palmata* mucus are not yet known. In the glycoproteins of *A. formosa* and the soft coral *P. americana*, β -arabinose was the terminal residue. If the composition of *A. palmata* mucus is similar, it is reasonable to expect arabinosidases to be strongly induced during the initial stages of growth on mucus. Consistent with this hypothesis, β -arabinosidases were induced in 9 out of 12 *Serratia* isolates during the first 2 hours of exposure to mucus. In the coral commensal bacteria, arabinosidases were induced after 18 h of incubation on mucus (see Table S1 in the supplemental material).

It is becoming increasingly clear that nutrients and signals found in the coral surface mucopolysaccharide layer dictate, at least in part, the structure and the composition of the associated microbiota (23, 25, 26). Elucidation of the differences in the strategies for coral commensals and opportunistic pathogens will help define the mechanisms of coral disease and may lead to research on defining potential approaches for managing or treating coral diseases.

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