

1 **Table S1. Primers for fimbrial genes used in this study.**

Fimbrial gene	Forward primer (5'→3') Reverse primer (5'→3')	Amplicon size (bp)	Reference or Gene accession #
<i>bcfD</i>	GACAACATGCGGAGTGTCAT GACATCTGTCAGCGGATCG	572	<i>Yue et al</i> (2012)
<i>csgA</i>	GGATTCCACGTTGAGCATTT GTTGTTGCCAAAACCAACCT	312	<i>Smith et al</i> (2010)
<i>fimA</i>	ACTATTGCGAGTCTGATGTTTG CGTATTTTCATGATAAAGGTGGC	508	<i>Wu et al</i> (2012)
<i>lpfD</i>	ATCCGTTTGTGGGTGAAAGT GGCATCAAAGCATGAAGGTT	597	<i>Yue et al</i> (2012)
<i>pefA</i>	ACACGCTGCCAATGAAGTGA ACTGCGAAAGATGCCACAGA	450	<i>Smith et al</i> (2010)
<i>pegD</i>	TATGTGGCAAAGACAGGAA GCAAAGAATCAATGGAGCA	524	AM933172.1* (2246402-2246925)
<i>safC</i>	TGTTCTGGCTCCTTGTTTGACG TTCTGTTTGACCTCCACCCGAG	612	<i>Townsend et al</i> (2001)
<i>sefA</i>	GATACTGCTGAACGTAGAAGG GCGTAAATCAGCATCTGCAGTAGC	488	<i>Oliveira et al</i> (2003)
<i>staA</i>	TTTAGAAGCATCGGCACG ATGGTTATGGCTATGGGTT	501	AL513382.1* (216884-217384)
<i>stbD</i>	CAGATTATCCTGCGATCGTG CATCCCGTCAACCCAAGTAG	554	<i>Yue et al</i> (2012)
<i>stcA</i>	GCTGCTATTTGGATCTAGTGTC TGAGTATGATTCAGGCGTTC	355	CP002614.1* (2246947-2247301)
<i>stdB</i>	GCCACATCAACCGAAAT CCAGAACCAGGTCTACGC	477	AM933172.1* (3077526-3078002)
<i>steB</i>	CGCACTCCATCGCCACAT GCTGCCAGTTATCGCCGTTCT	549	AM933172.1* (2986399-2986947)
<i>stfH</i>	CTTCGCAAGCGCACTAGTTA CGCATCCCAGGAGATATCAA	555	<i>Yue et al</i> (2012)
<i>stgA</i>	CGCTCTGGTTCTGTCTTCT CTGTGGTATCAATCGTGCT	443	AL513382.1* (3780619-3781061)
<i>sthE</i>	GTTATATCTGGCGCTTATTCTGC CCAAATACAGATGGGTCAGGA	572	<i>Yue et al</i> (2012)
<i>stiH</i>	TCCGCAGGGAATAAGGGTA TCAGCGAATCTGAATCTCCA	600	<i>Yue et al</i> (2012)
<i>stjA</i>	TATGAAACGGCAAAGGGAGT CGTCAGCACTTCCCCTGTA	593	<i>Yue et al</i> (2012)
<i>stkA</i>	TGCCTGTACGGTTCCTGC TTCCACGGTCTGGTTTCT	482	CP003416.1* (952532-953013)
<i>tcfA</i>	TCGCTATGTTTGCATGTGGT TTCAGGAACAGCCTCGAAGT	316	<i>Suez et al</i> (2013)

* The PCR system was established in this study with the included gene accession # and the location of target gene.

Yue M, Rankin SC, Blanchet RT, Nulton JD, Edwards RA, Schifferli DM. 2012. Diversification of the *Salmonella* fimbriae: A model of macro- and microevolution. PLoS One. doi: 10.1371/journal.pone.0038596.

Smith KP, George J, Cadle KM, Kumar S, Aragon SJ, Hernandez RL, Jones SE, Floyd JL, Varela MF. 2010. Elucidation of antimicrobial susceptibility profiles and genotyping of *Salmonella enterica* isolates from clinical cases of salmonellosis in New Mexico in 2008. World J. Microbiol. Biotechnol. **26**:1025–1031.

Yue M, Schmieder R, Edwards RA, Rankin SC, Schifferli DM. 2012. Microfluidic PCR combined with pyrosequencing for identification of allelic variants with phenotypic associations among targeted *Salmonella* genes. Appl. Environ. Microbiol. **78**:7480–7482.

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Townsend SM, Kramer NE, Edwards R, Baker S, Hamlin N, Simmonds M, Stevens K, Maloy S, Parkhill J, Dougan G, Bäumler AJ. 2001. *Salmonella enterica* serovar Typhi possesses a unique repertoire of fimbrial gene sequences. Infect. Immun. **69**:2894–2901.

Oliveira SD, Rodenbusch CR, Cé MC, Rocha SL, Canal CW. 2003. Evaluation of selective and non-selective enrichment PCR procedures for *Salmonella* detection. Lett. Appl. Microbiol. **36**:217–221.

Suez J, Porwollik S, Dagan A, Marzel A, Schorr YI, Desai PT, Agmon V, McClelland M, Rahav G, Gal-Mor O. 2013. Virulence gene profiling and pathogenicity characterization of non-typhoidal *Salmonella* accounted for invasive disease in humans. PLoS One. doi: 10.1371/journal.pone.0058449.

5 **Table S2. Poultry *Salmonella* isolated from 51 farms.**

Number positive / number tested (%)				
Chicken	Duck	Goose	Pigeon	Turkey
11/87 (12.6)	3/87 (3.4)	5/31 (16.1)	0/33 (0)	8/93 (8.6)
21/193 (10.9)	7/70 (10.0)	7/95 (7.4)	6/89 (6.7)	8/64 (12.5)
5/82 (6.1)	7/66 (10.6)	10/79 (12.6)	4/50 (8.0)	9/52 (17.3)
16/143 (11.2)	2/52 (3.8)	6/30 (20.0)	1/54 (1.9)	5/47 (10.6)
20/180 (11.1)	5/88 (5.7)	7/65 (10.8)	3/55 (5.5)	2/53 (3.8)
6/39 (15.4)	4/49 (8.2)	4/29 (13.8)	0/16 (0)	
3/35 (8.6)	5/67 (7.5)	4/38 (10.5)	2/57 (3.5)	
14/169 (8.3)	5/73 (6.8)	11/42 (26.2)	1/63 (1.6)	
3/93 (3.2)	3/49 (6.1)	5/42 (11.9)		
6/68 (8.8)		3/35 (8.6)		
3/58 (5.2)		4/47 (8.5)		
9/123 (7.3)				
4/55 (7.3)				
8/64 (12.5)				
20/137 (14.6)				
7/59 (11.9)				
9/82 (11.0)				
2/39 (5.1)				

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8 **Table S3. Poultry *Salmonella* serovars in 12 provinces of China.**

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Serovars	Number of isolates											
	Jian gsu	Anh ui	Zhe jian g	Shan dong	Sha ngh ai	He bei	Bei jing	Si chuan	He nan	Xinjiang	Guan gdon g	Heilong jiang
<i>S. Pullorum</i>	20	2	3	8	0	4	3	0	3	10	0	2
<i>S. Typhimurium</i>	24	1	5	4	1	3	0	2	4	6	0	0
<i>S. Enteritidis</i>	20	5	6	0	0	0	0	3	2	3	0	0
<i>S. Indiana</i>	11	3	2	4	0	0	0	0	2	3	0	0
<i>S. Heidelberg</i>	4	7	8	0	0	0	0	0	1	0	0	2
<i>S. Potsdam</i>	5	3	2	4	0	0	0	0	2	7	0	0
<i>S. Kentucky</i>	9	0	0	4	0	3	0	0	0	2	1	0
<i>S. Thompson</i>	4	1	0	0	0	1	1	0	0	2	0	0
<i>S. Saintpaul</i>	6	0	1	0	3	0	0	0	0	1	5	0
<i>S. Kottbus</i>	1	2	0	1	0	0	0	0	0	0	0	0
<i>S. Agona</i>	0	0	0	0	0	0	0	0	3	0	0	0
<i>S. Gallinarum</i>	6	0	3	0	0	0	0	1	0	0	0	2
<i>S. Blockley</i>	1	9	0	0	0	0	0	0	1	2	0	0
<i>S. Bazenheid</i>	2	6	0	2	0	0	0	0	0	0	0	0
<i>S. Anatum</i>	3	0	2	0	0	0	0	0	0	0	0	0
<i>S. Montevideo</i>	0	0	0	4	0	1	0	0	0	0	0	0
<i>S. Derby</i>	3	0	0	0	0	0	0	4	0	0	0	0
<i>S. Reading</i>	0	0	1	2	0	0	0	0	0	0	0	0
<i>S. Senftenberg</i>	0	0	0	1	0	0	0	0	0	0	0	0
<i>S. Meleagridis</i>	0	2	0	0	0	0	0	0	0	0	0	0
Total	119	41	33	34	4	12	4	10	18	36	6	6

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Fig. S1. Map of the poultry *Salmonella* spp. detection regions. For this study, 3,566 rectal swab samples were obtained from 7 regions of 12 provinces in China. Below each region label is the total number of *Salmonella*-positive samples over the total number of samples received from poultry farms in that region.